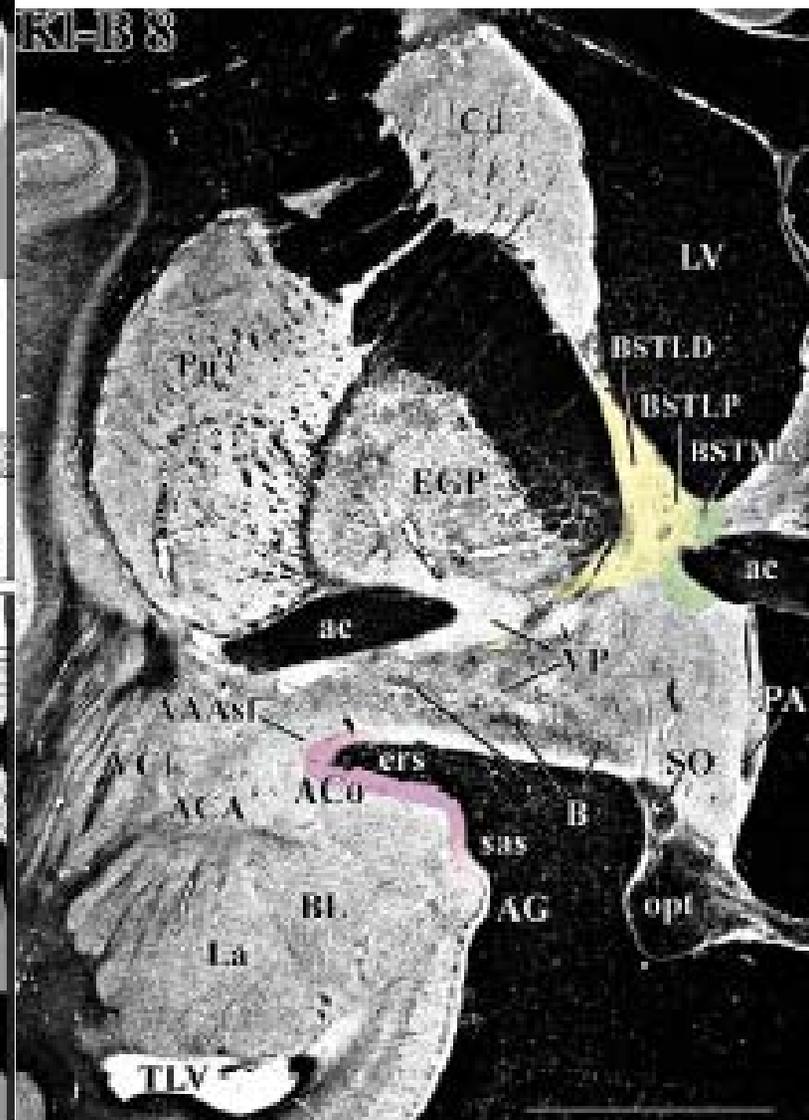
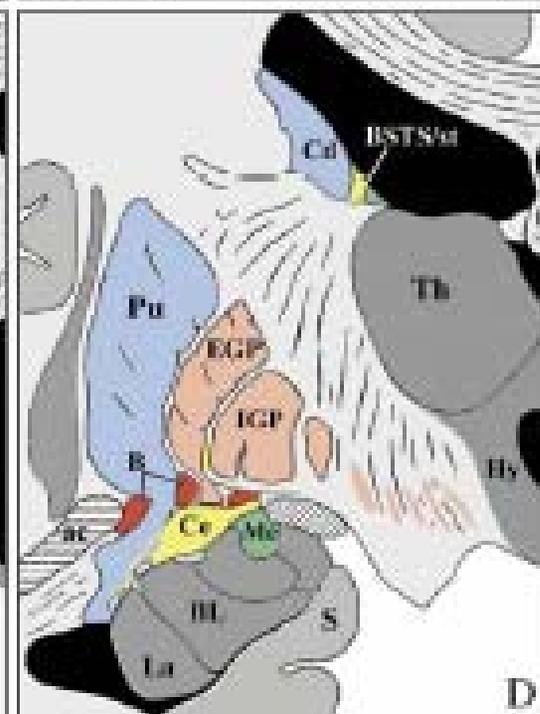
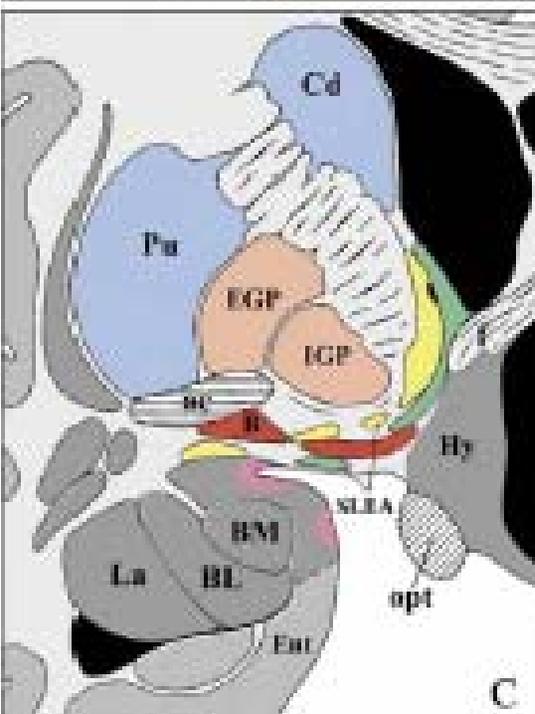
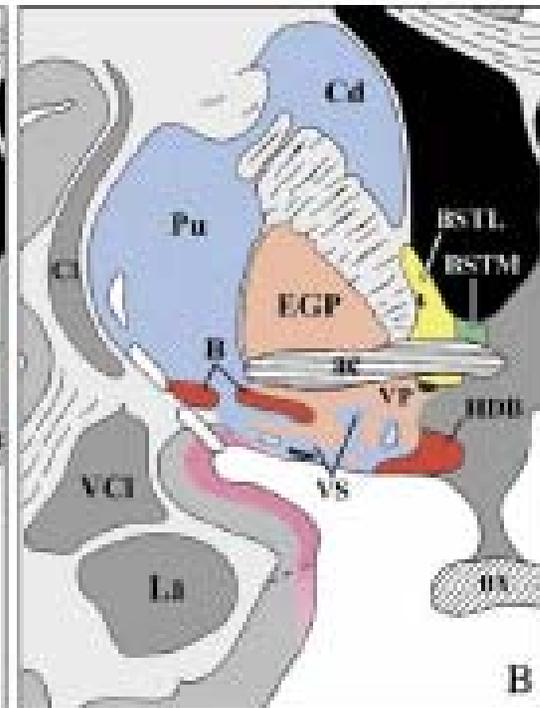
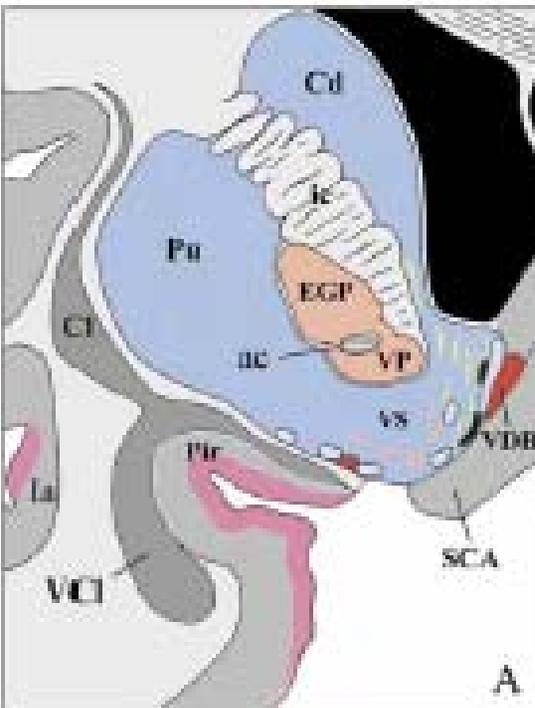
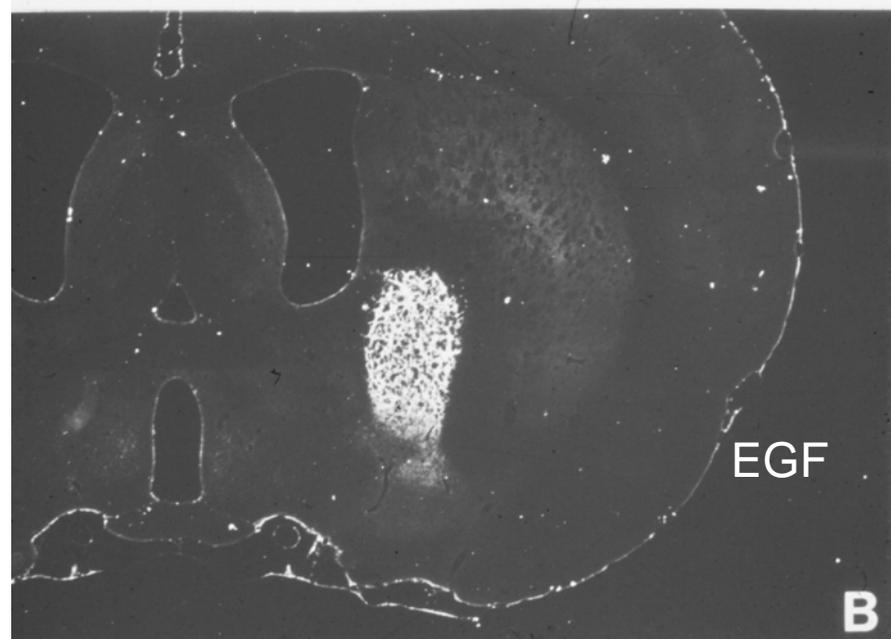
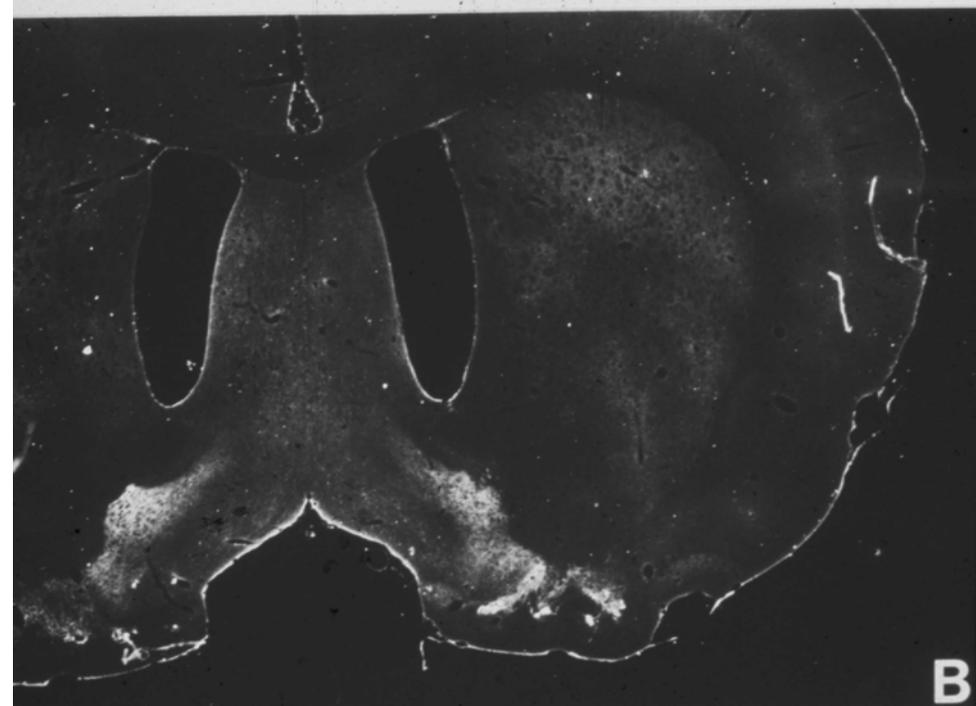
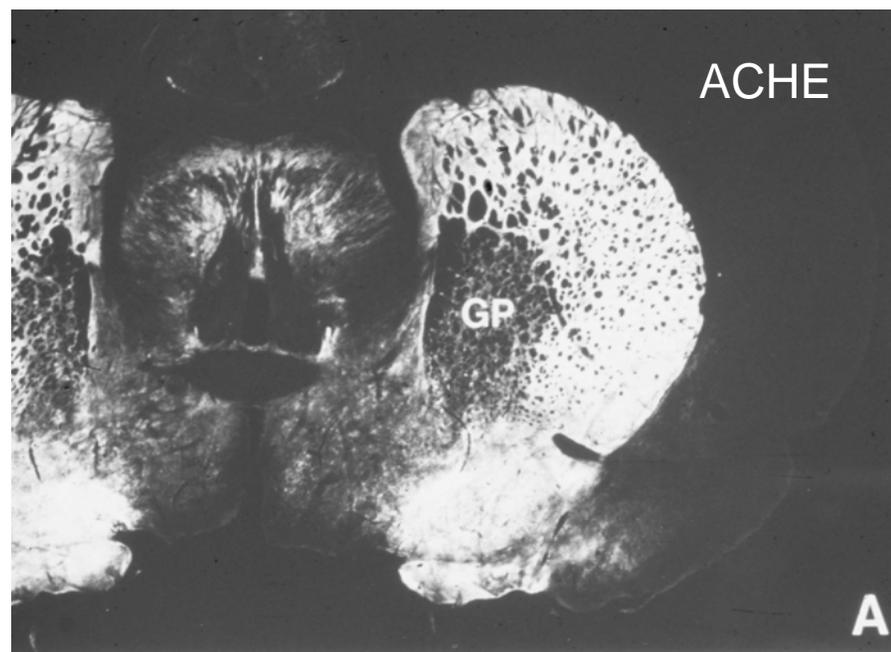
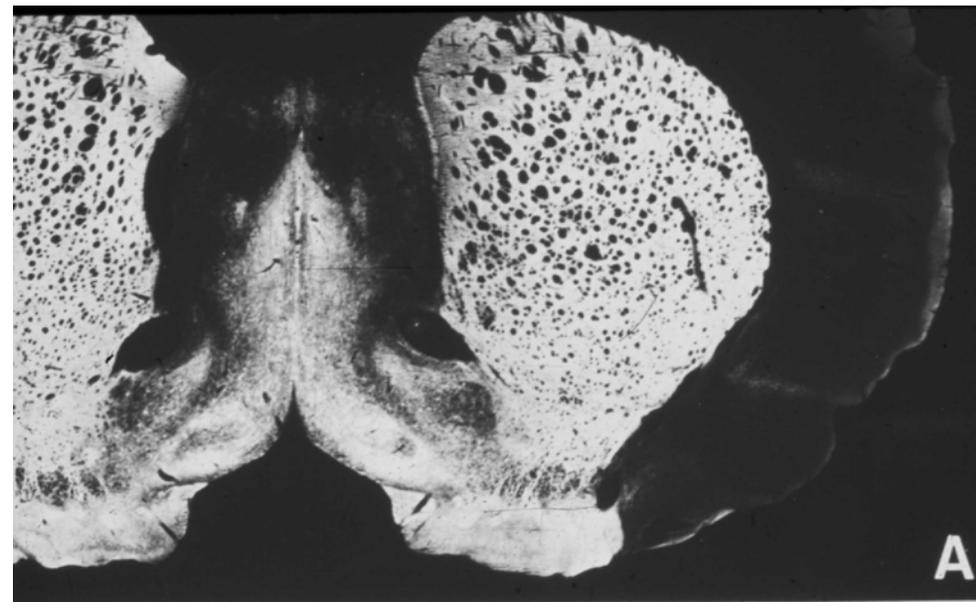


BASAL FOREBRAIN

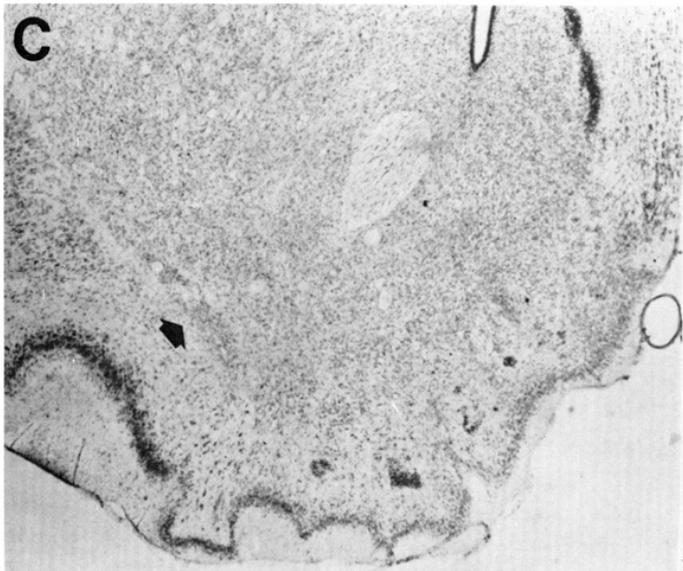
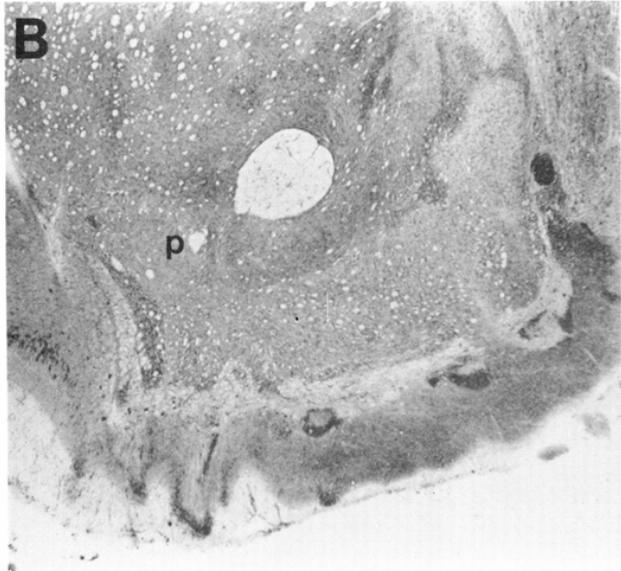
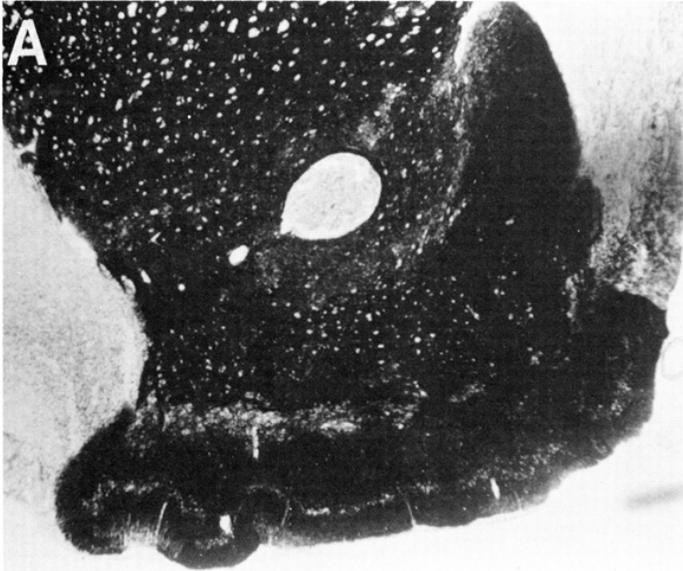
BASAL FOREBRAIN –FUNCTIONAL ANATOMICAL ‘MACROSYSTEMS’



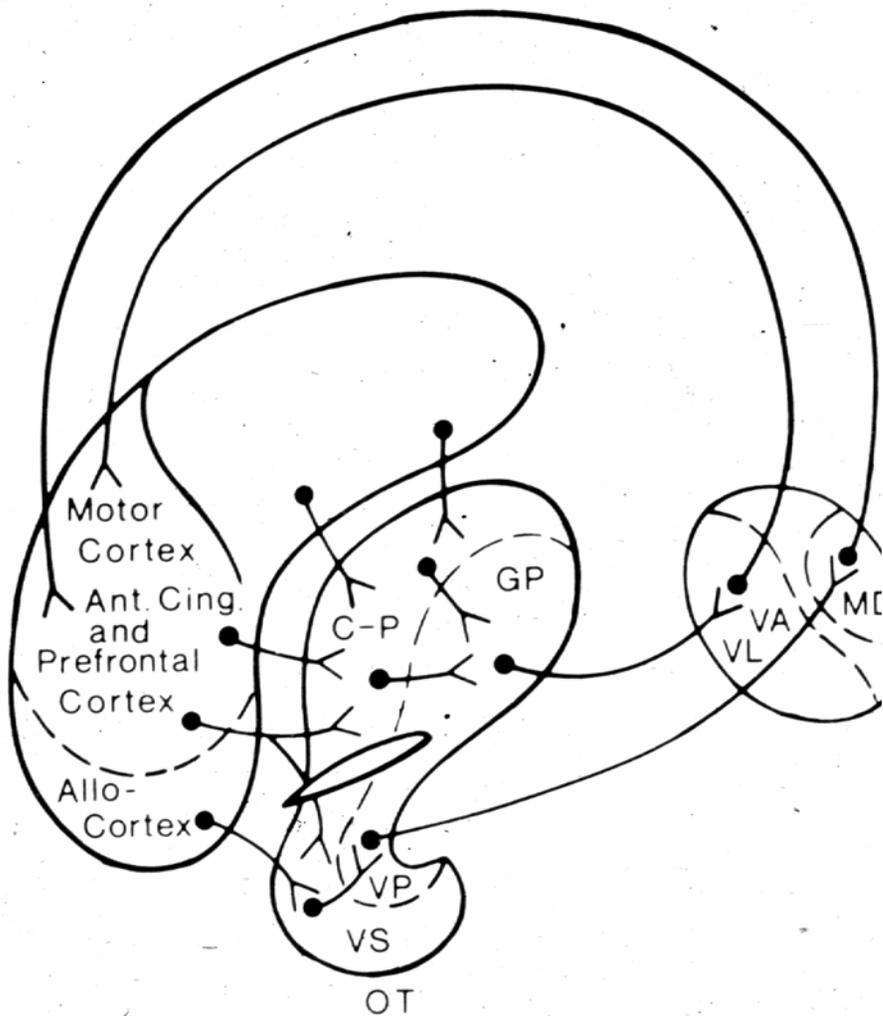
1. VENTRAL STRIATO-PALLIDAL SYSTEM IN RAT



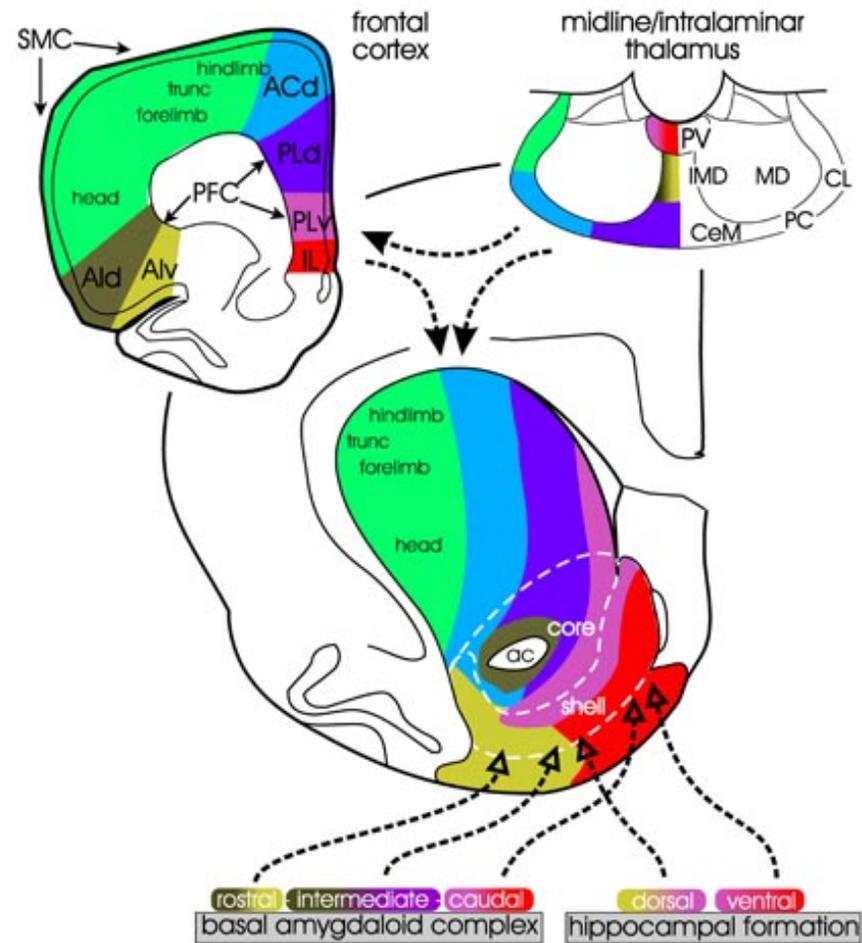
ACCUMBENS: Core and Shell



CIRCUITRIES OF THE VENTRAL STRIATO-PALLIDAL SYSTEM



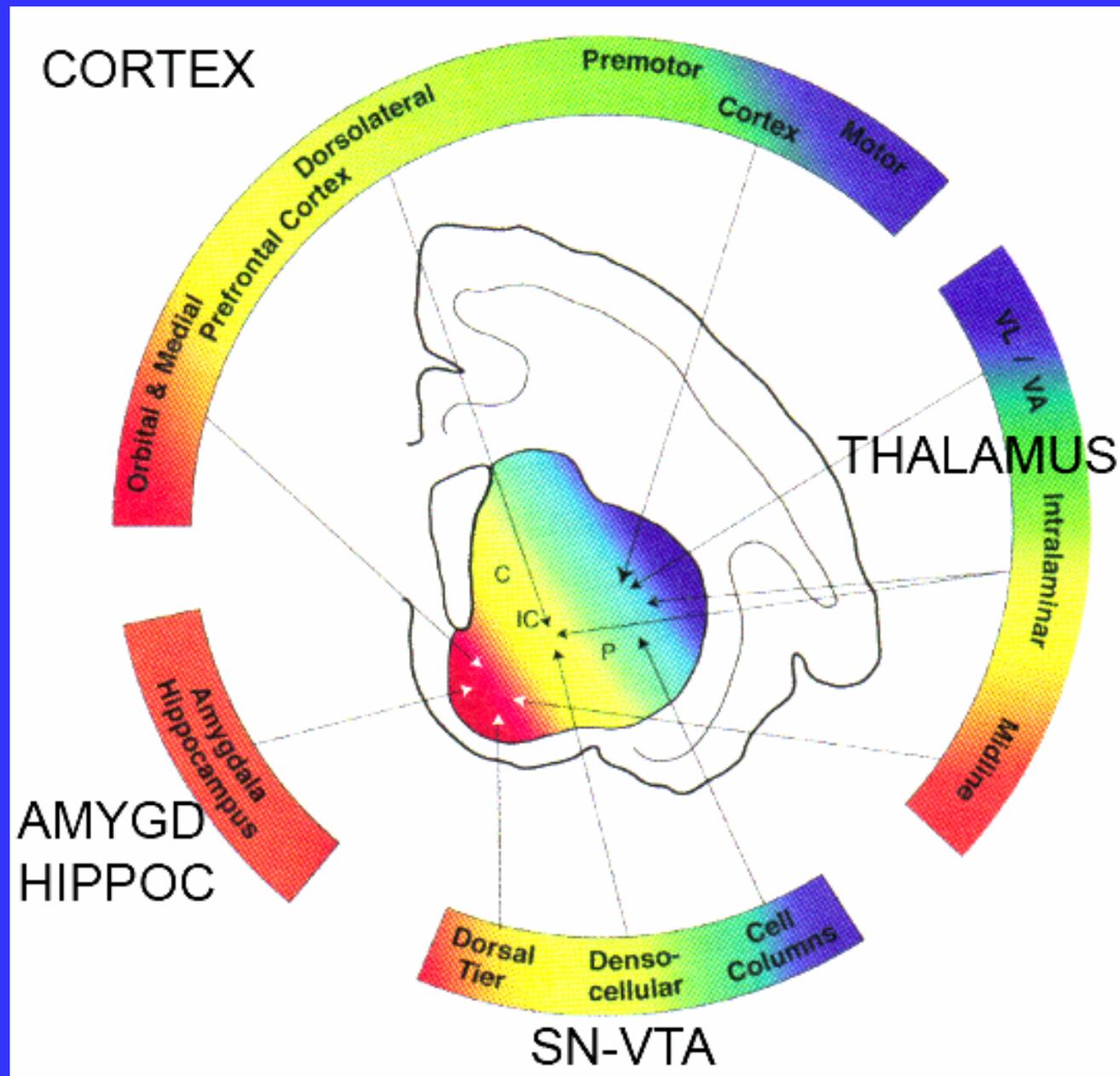
Heimer et al., 1981



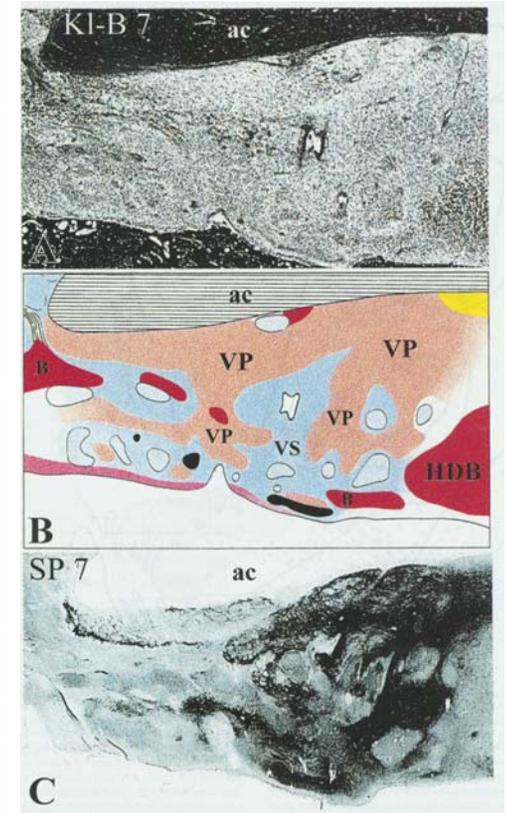
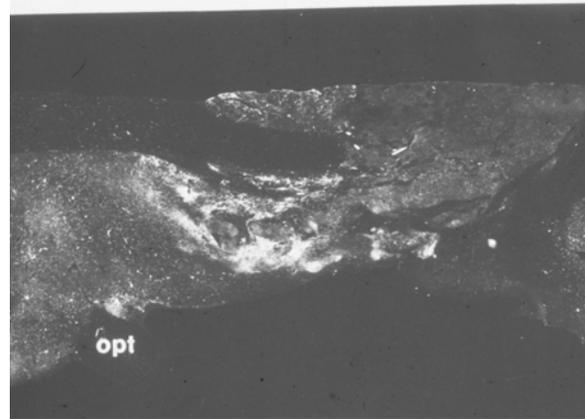
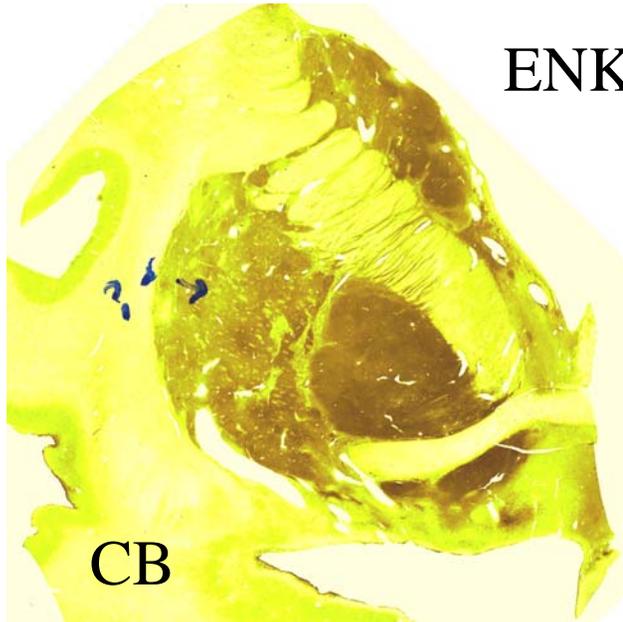
Voorn et al., 2004

CIRCUITRIES OF THE DORSAL AND VENTRAL STRIATUM

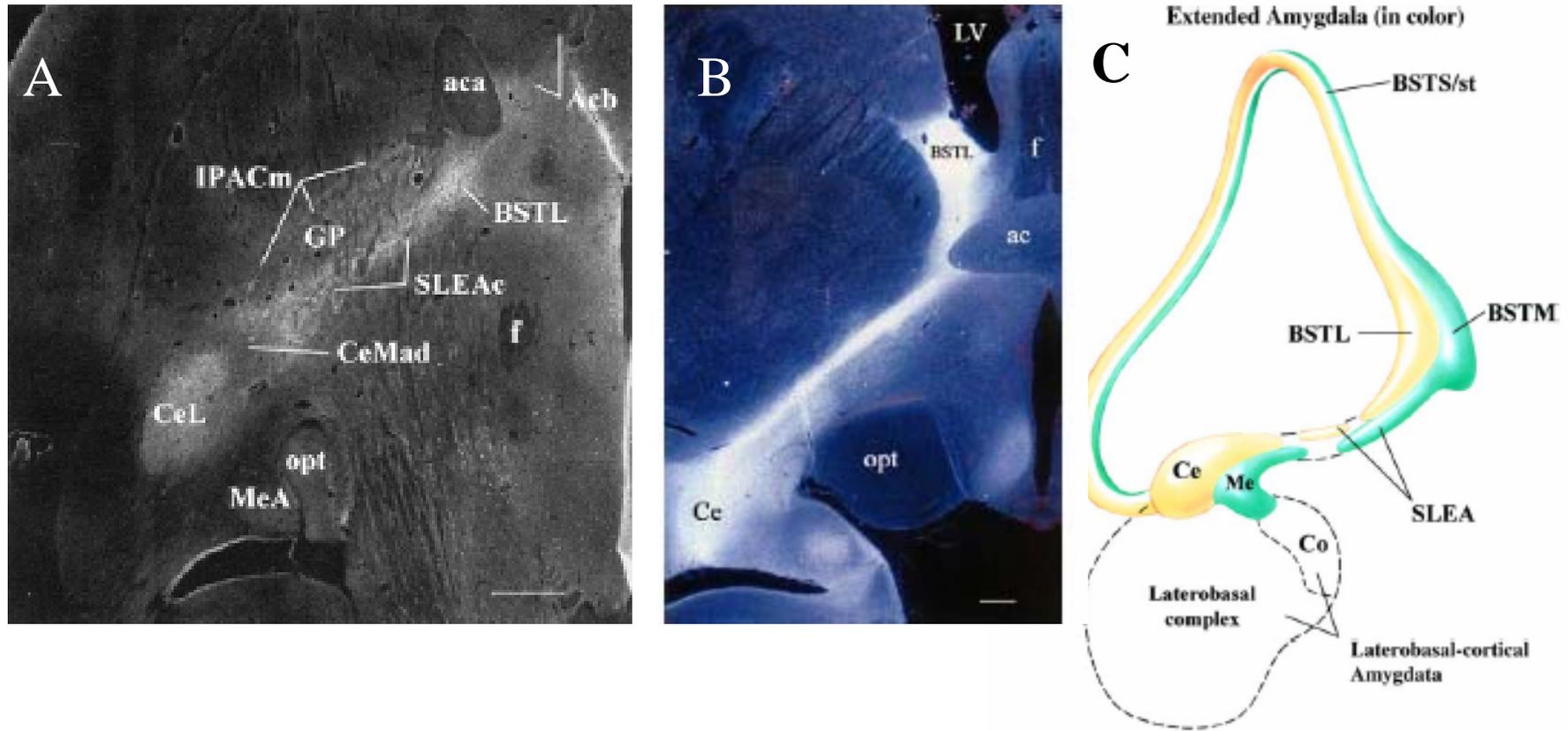
The colors denote functional distinctions. Blue: motor ctx, executive of motor actions; green premotor ctx, planning of movements; yellow dorsal and lateral PFC: cognitive and executive functions; orange: orbital PFC: goal-directed behaviors and motivations, red: mPFC, goal directed behaviors and emotional processing (Haber and Gdowski, 2004)



1. VENTRAL STRIATO-PALLIDAL SYSTEM IN HUMANS

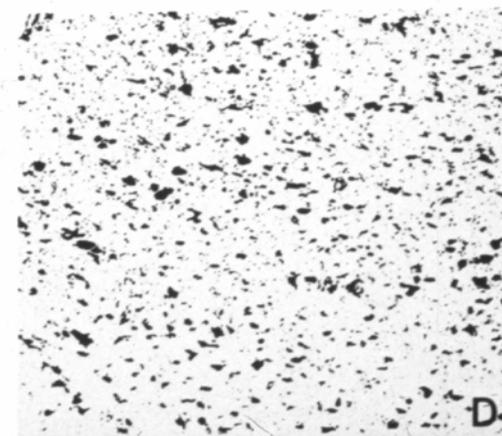
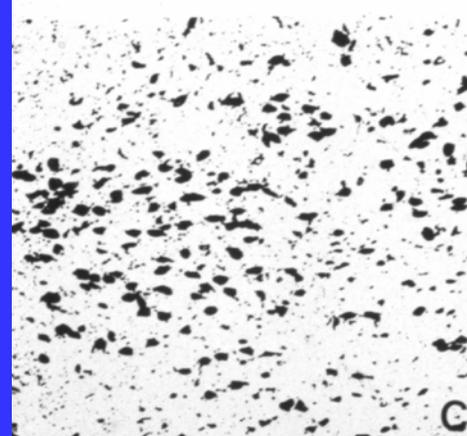
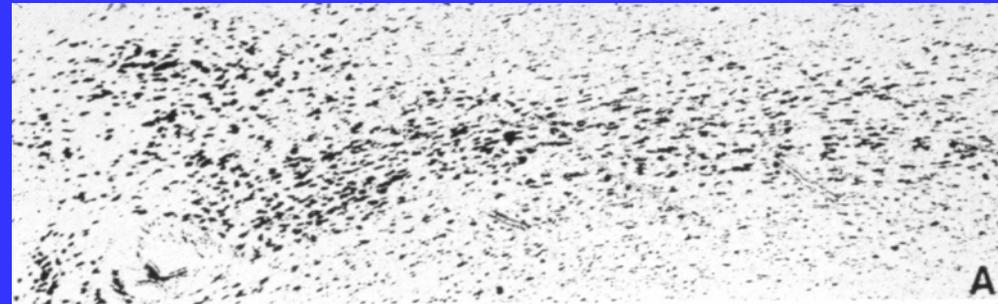


2. EXTENDED AMYGDALA IN RAT AND PRIMATES



A: Dark field image of normal cupric silver staining of the central division of the extended amygdala in rat in horizontal section. **B:** Dark field image of the sublenticular area of the extended amygdala in rhesus monkey following an injection of 3H-amino acid in the region of the central amygdaloid nucleus (Ce). **C:** The extended amygdala shown in isolation from the rest of the brain, with the extension of the Ce and Medial (Me) amygdaloid nuclei within the stria terminalis (st) and through the sublenticular region of the bed nucleus of the stria terminalis (BST). From Heimer and Hoesen (in press)

3. DIAGONAL BAND-NUCLEUS BASALIS MEYNERI



1809: J. C. Reil “unnamed substance”
1861: K. B. Reichert “substantia innominata”
1872: T. Meynert “nucleus basalis”
1896: V. Kolliker “topographical description”
1873-1900 - Golgi and Nissl “staining”
1910-1940: Beccari, Ramon y Cajal, Ayala,
Foix and Nicolesco, Kodama and
others “cellular description”
1942: Brockhaus “basal nuclei complex”
1949: Moruzzi and Mogoun “energizing system”
Koelle and Friedenwald “AChE”
1960's: Gorry, Leontovitch and Zhukova, Ramon
Moliner and Nauta, Lewis and Shute
“BF is an extension of the isodendritic
core” and “cholinergic system”
1980: Eckenstein and Thoenen “ChAT”
1983: Mesulam “Ch nomenclature”

Cholinergic hypothesis of AD

Bowen et al., 1976; Davies and Maloney,
1976; Whitehouse et al., 1981

Basal forebrain in reward

De Long, 1971; Rolls et al., 1976

Basal forebrain in cortical activation

Detari and Vanderwolf, 1987

Basal forebrain in cortical plasticity

Silito and Kemp, 1983; Sato et al., 1987;
Juliano et al, 1989; Weinberger, 1991;
Merzenich, 1998

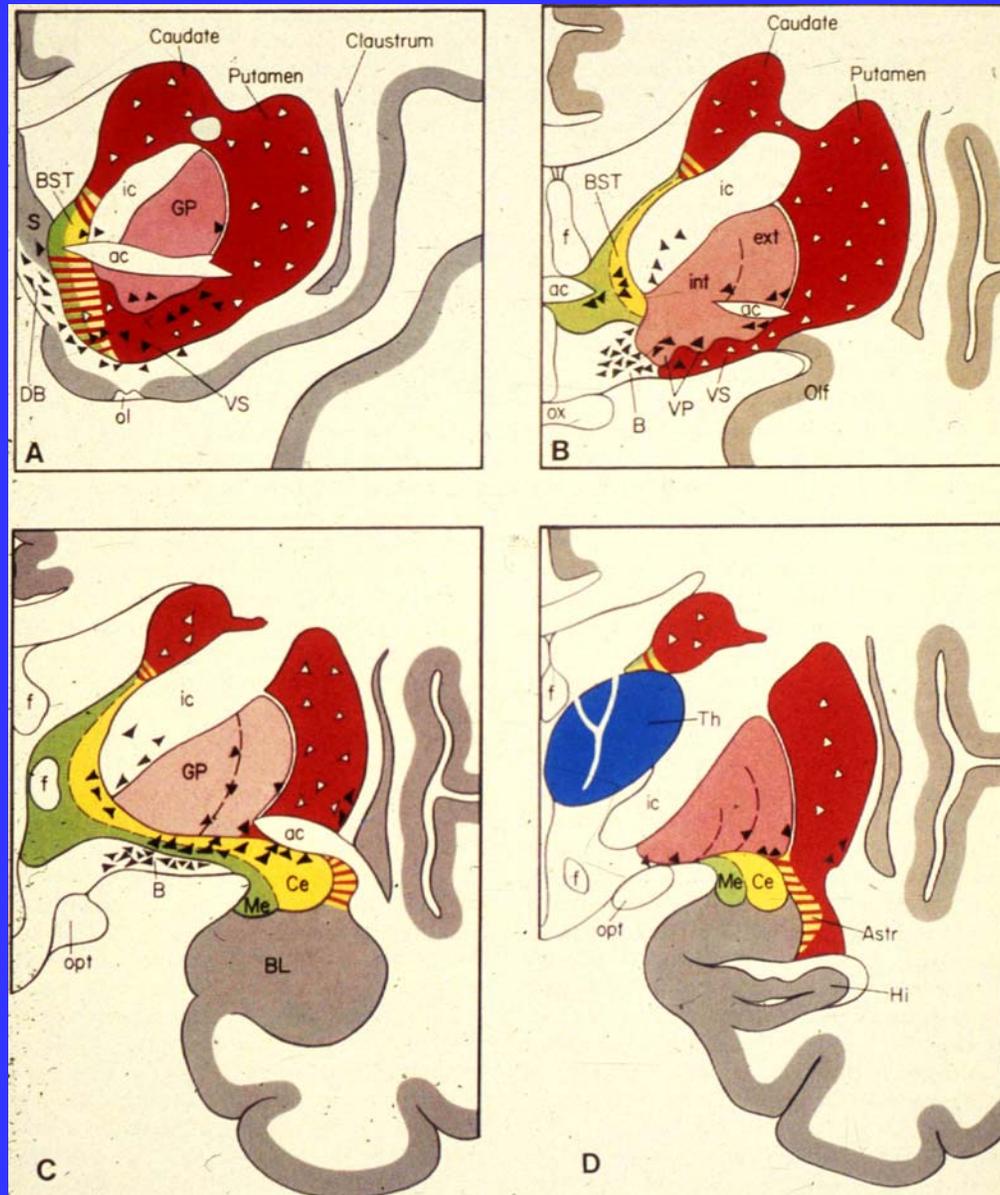
Basal forebrain in learning and memory

Olton, 1991

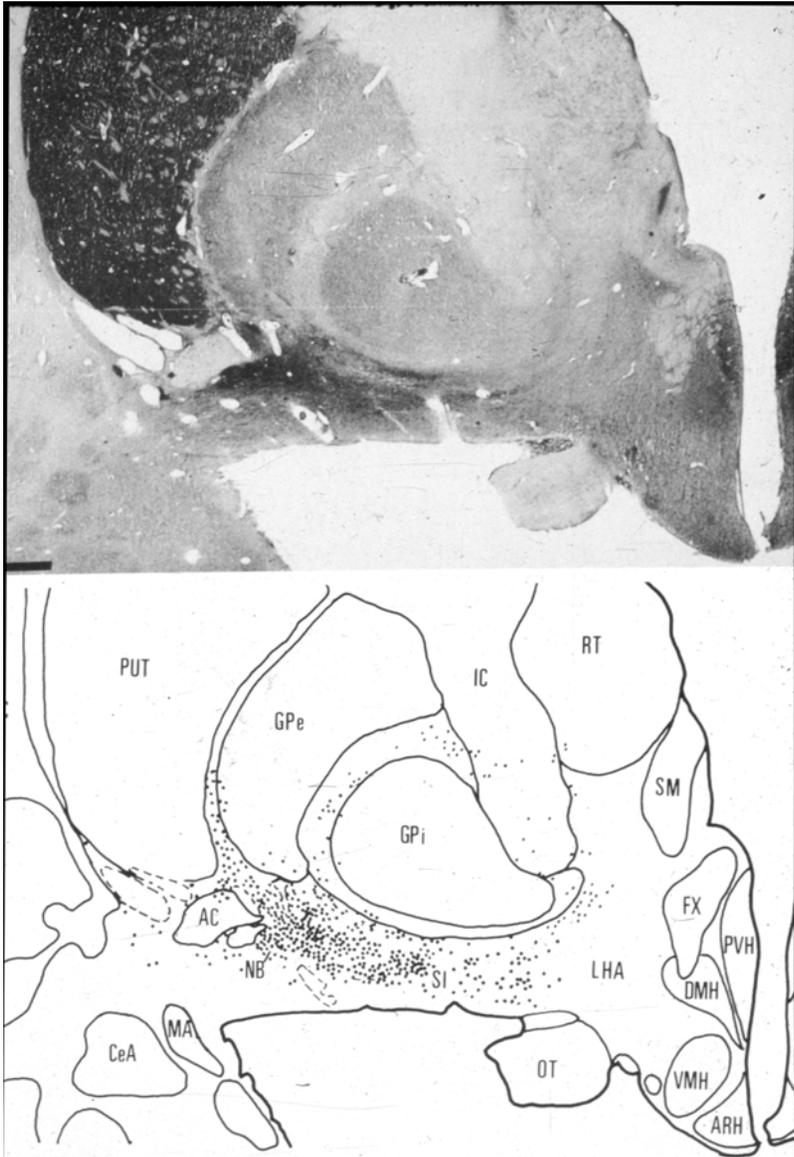
Basal forebrain in attention

Dunnett et al., 1991; Everitt and Robbins,
1997; Gallagher, 1995; Paus et al., 1997;
Sarter and Bruno, 1997

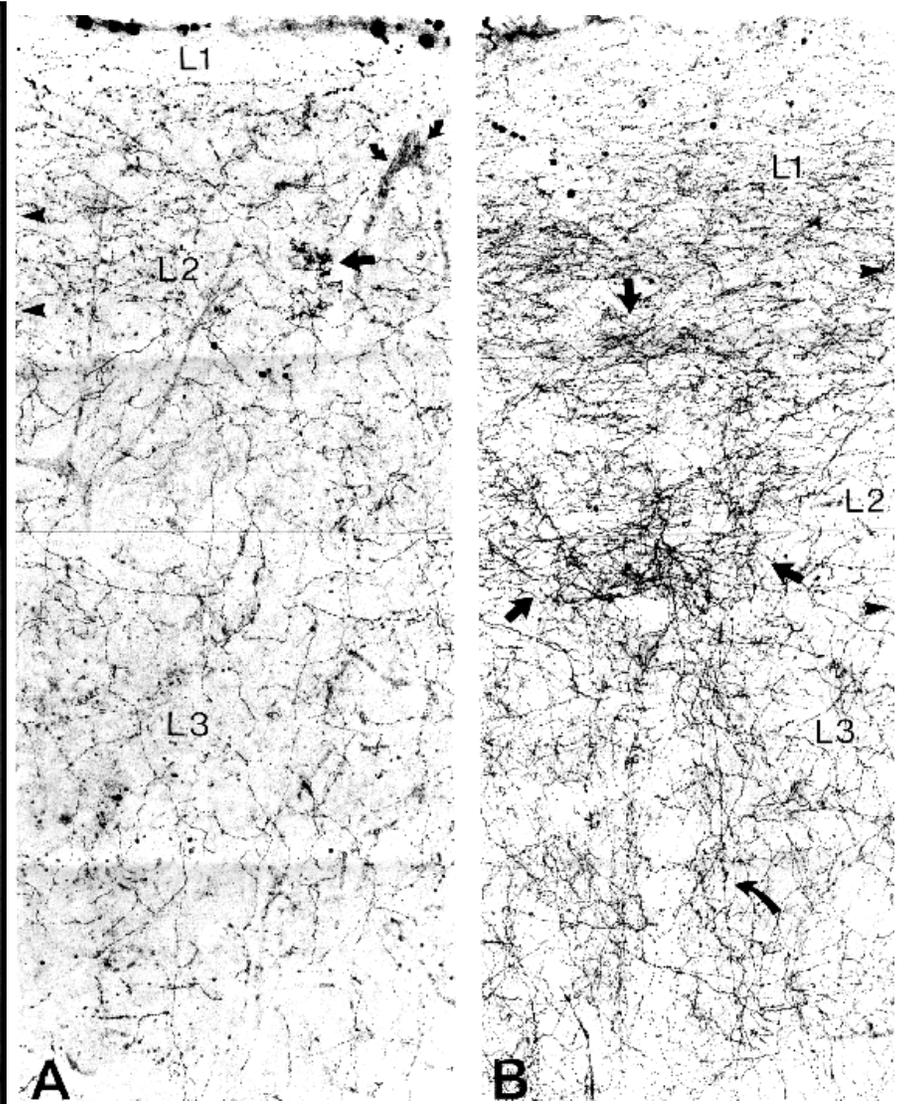
RELATION OF CORTICOPETAL CELLS IN THE BF TO SEPTUM-SUBSTANTIA INNOMINATA-AMYGDALA-BST CONTINUUM



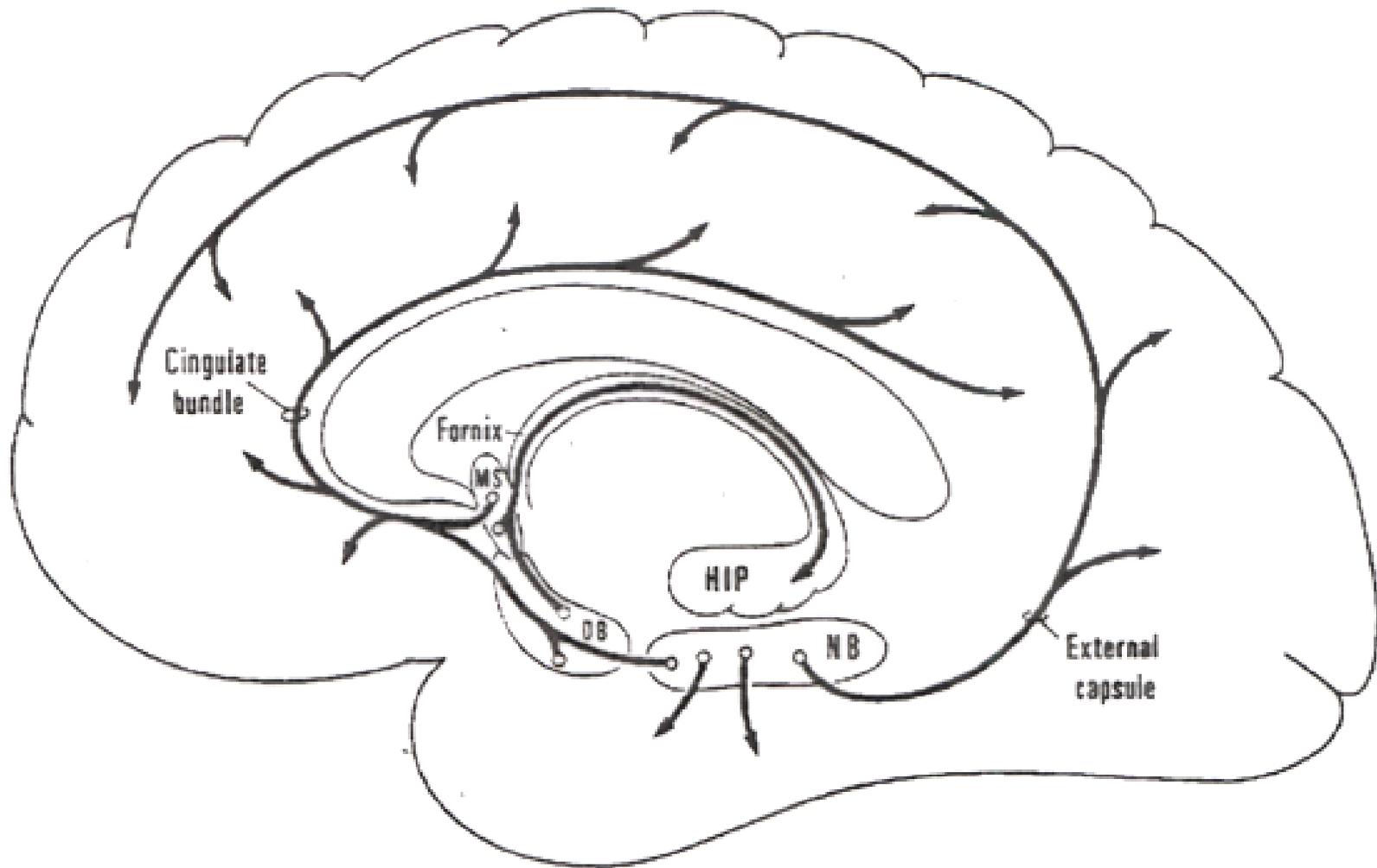
ACHE cell staining (Saper)



ChAT terminals (Mesulam)

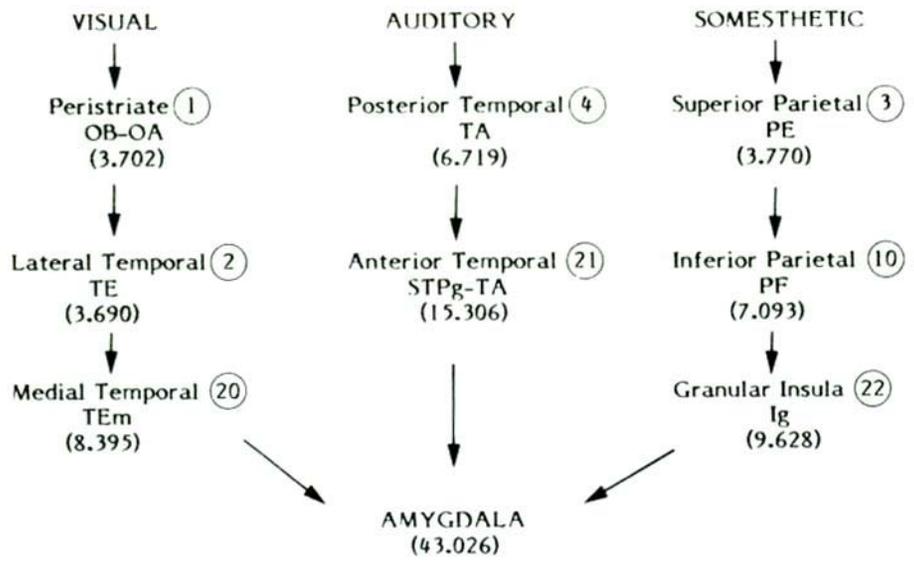
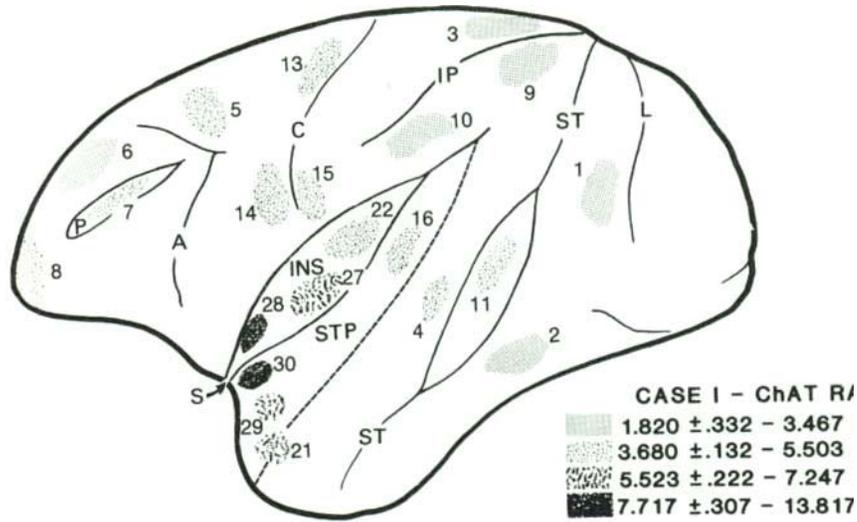


Basal forebrain cholinergic system in human

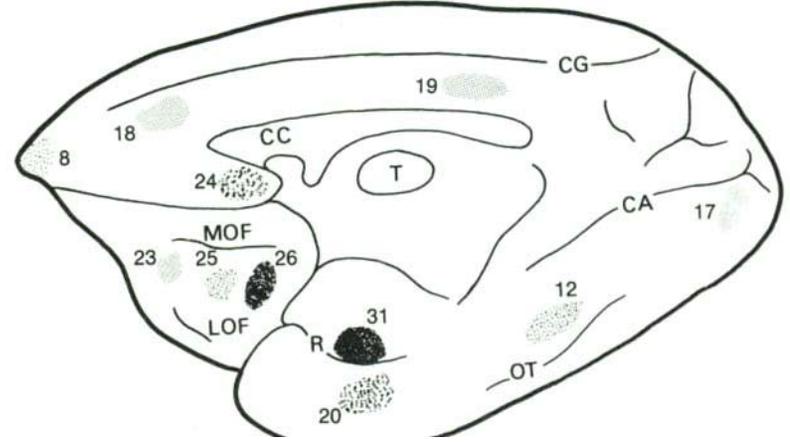


Summary of the major pathways for cholinergic innervation of the cortical mantle by the magnocellular basal complex (Saper, 1990).

REGIONAL CHAT ACTIVITIES IN RHESUS MONKEY

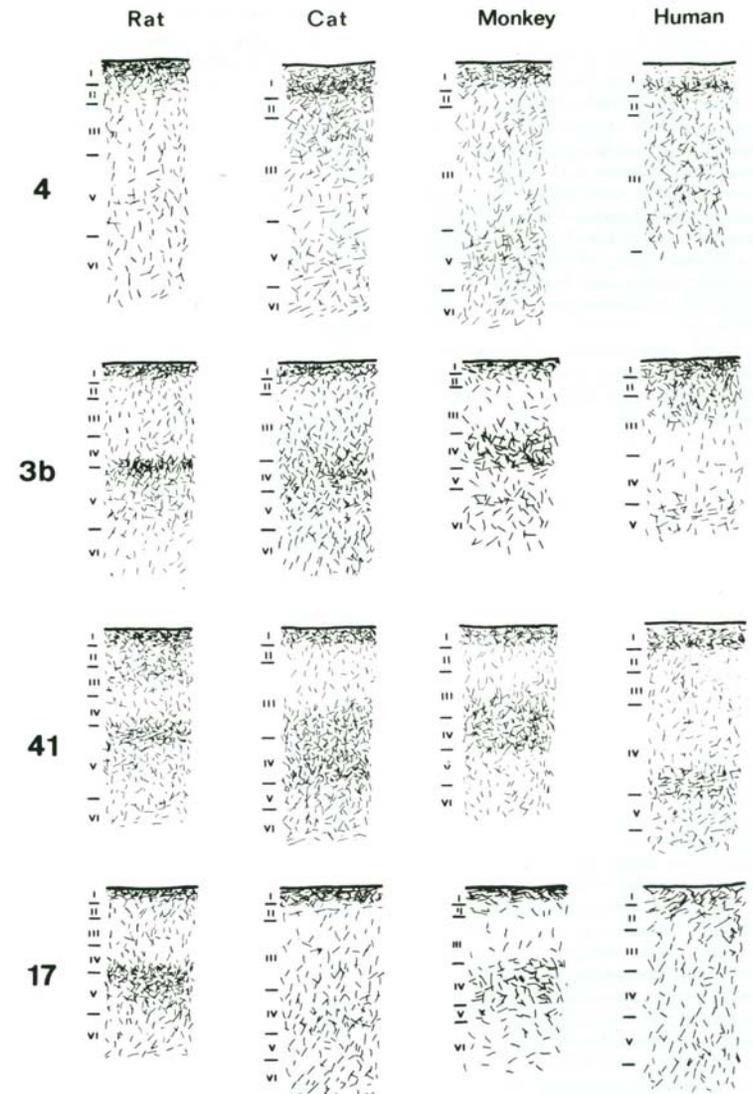
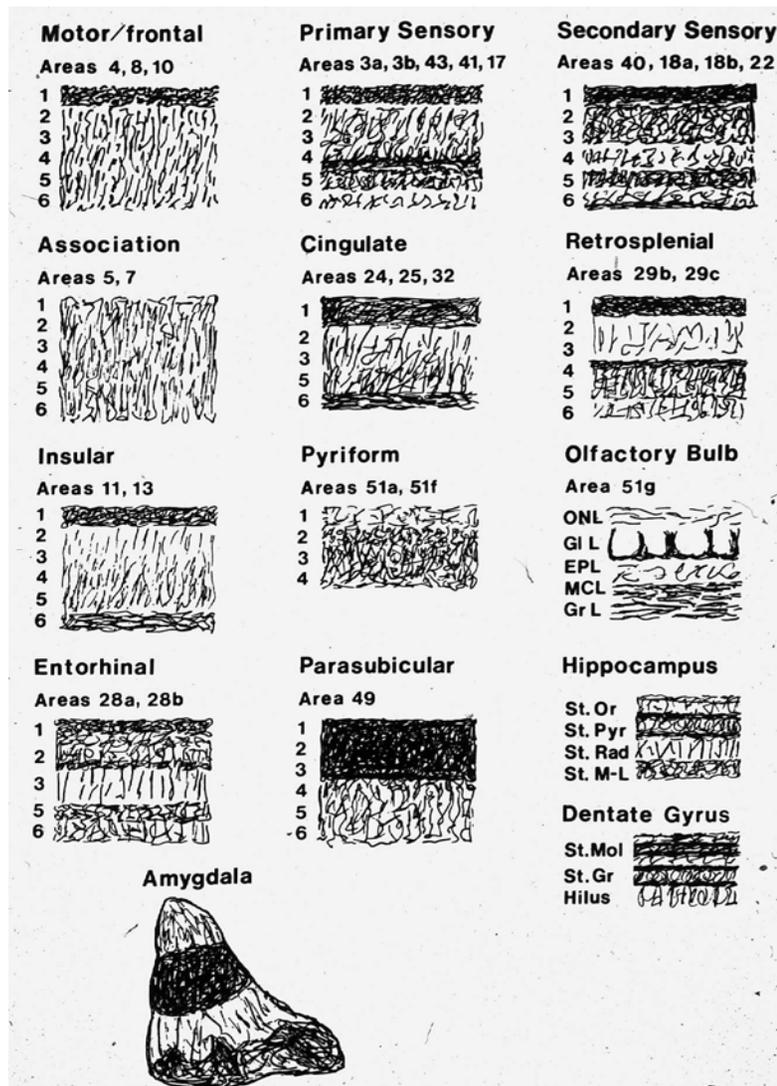


A



Mesulam et al., 1986

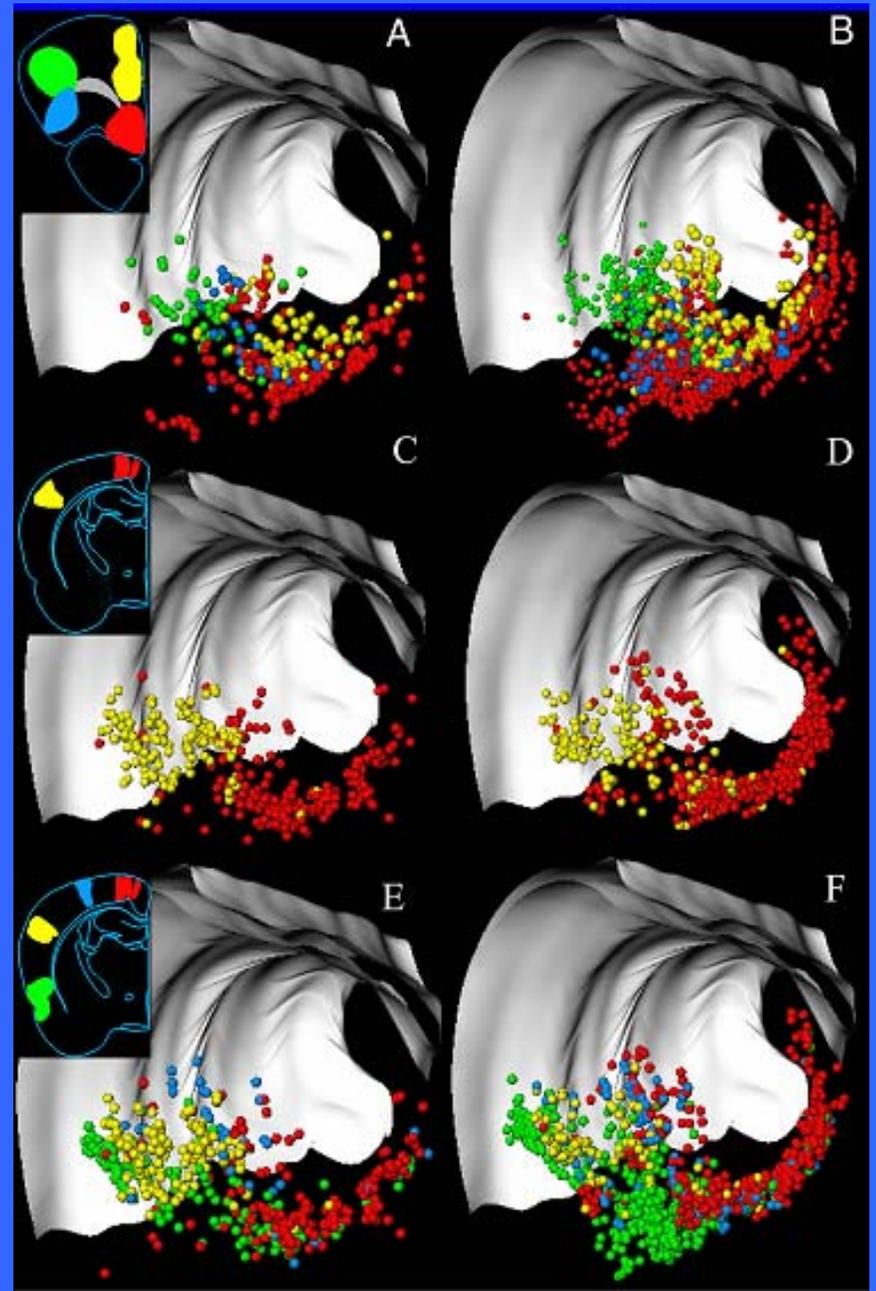
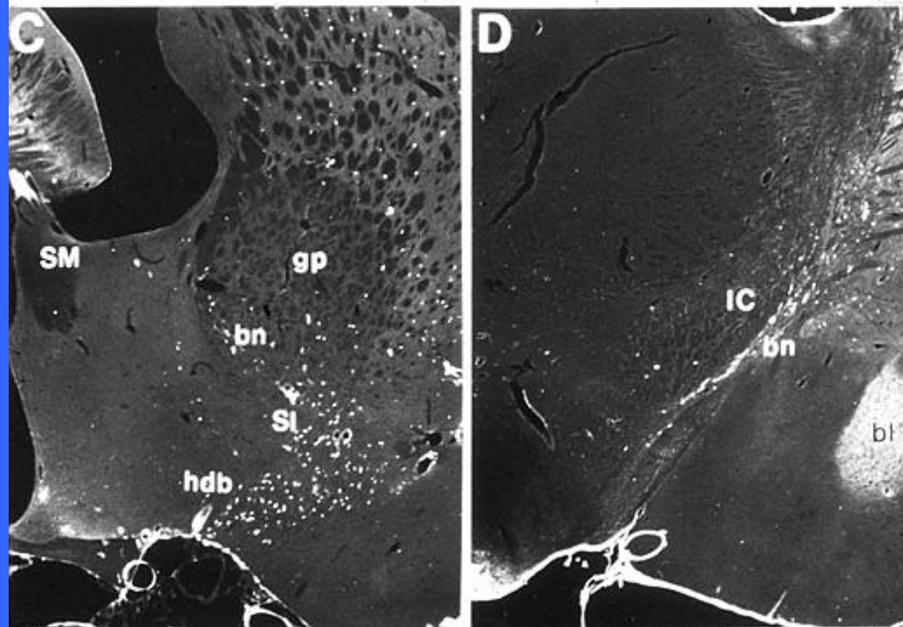
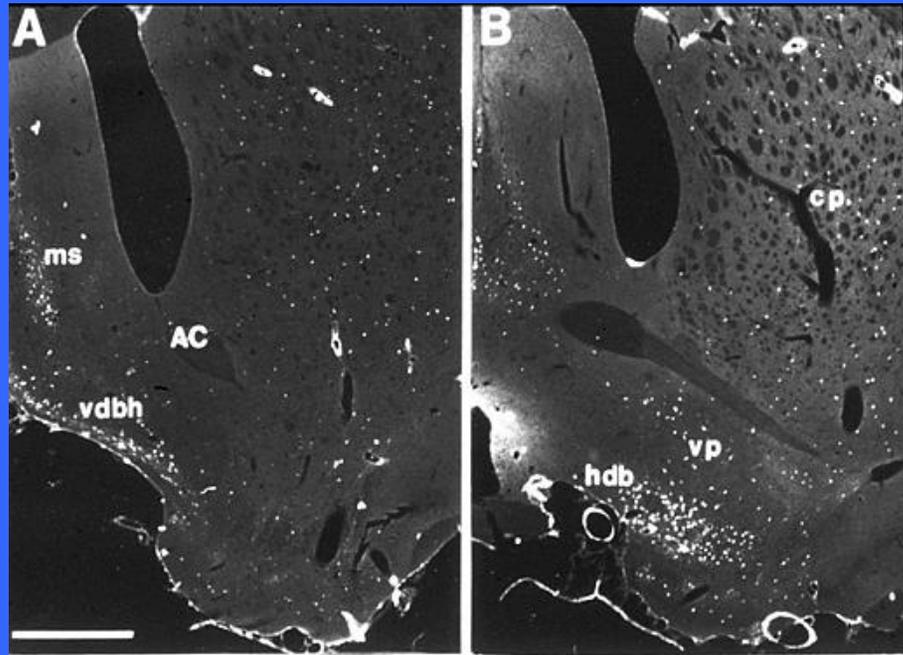
CHOLINERGIC INNERVATION IN VARIOUS CORTICAL AREAS

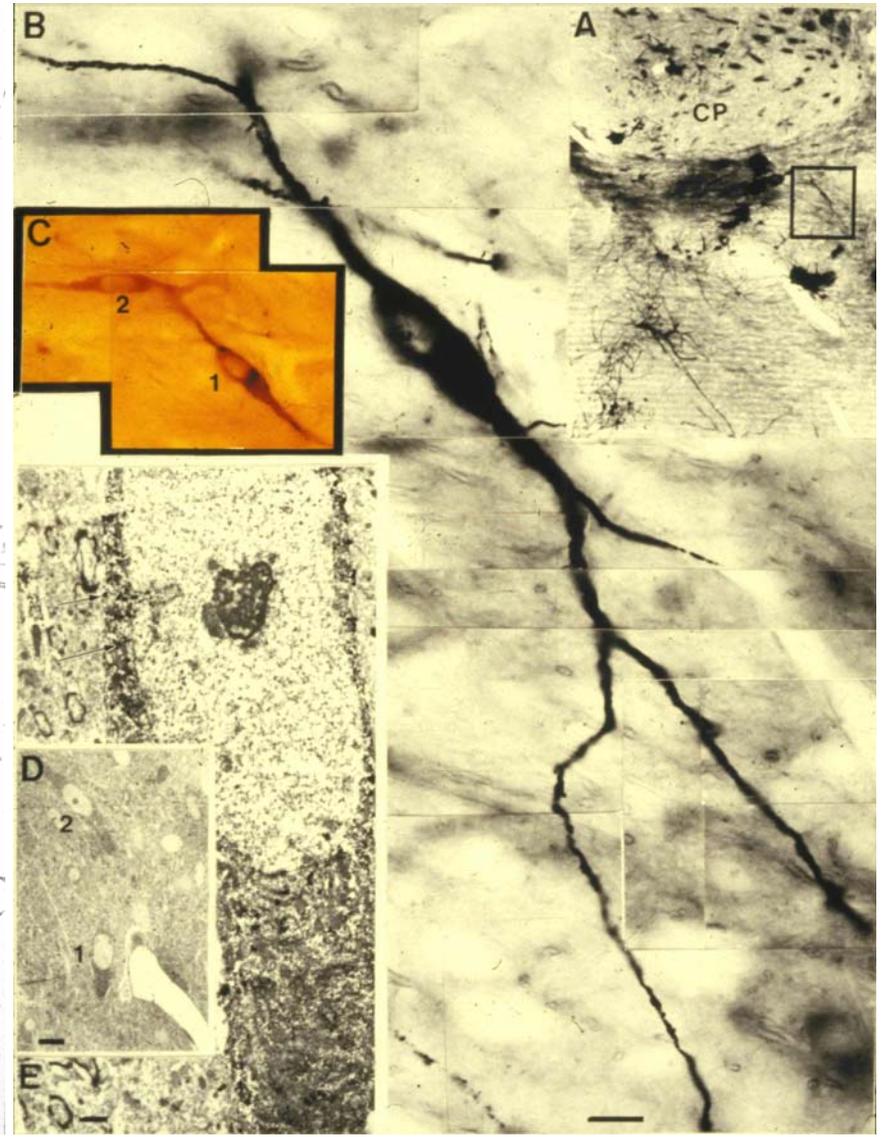
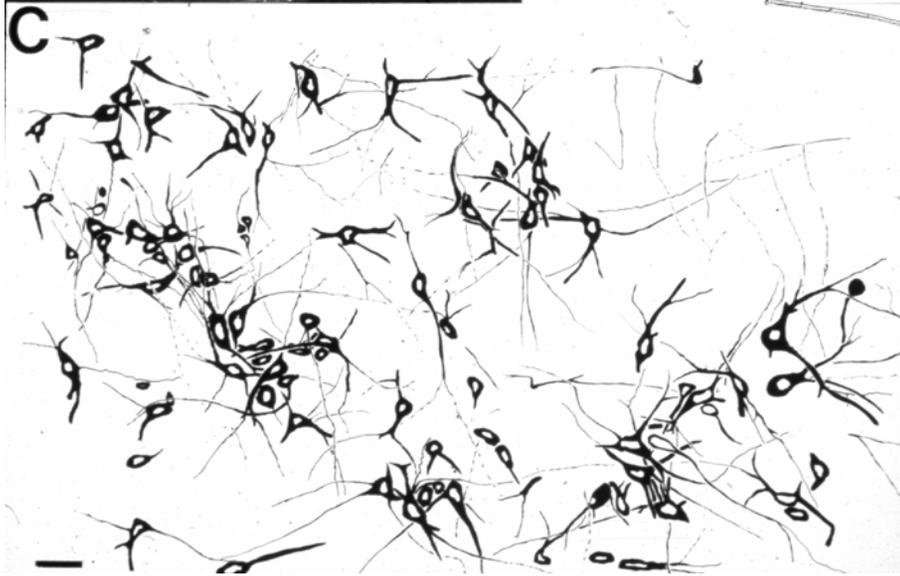
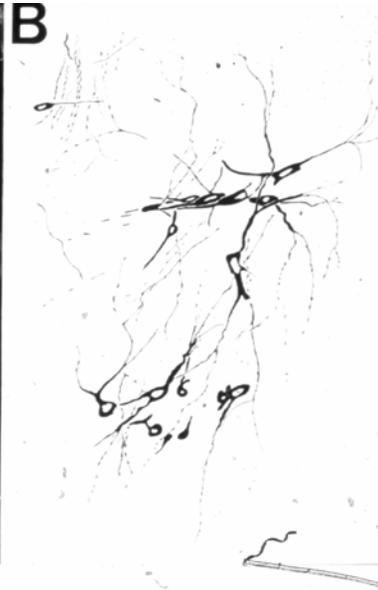
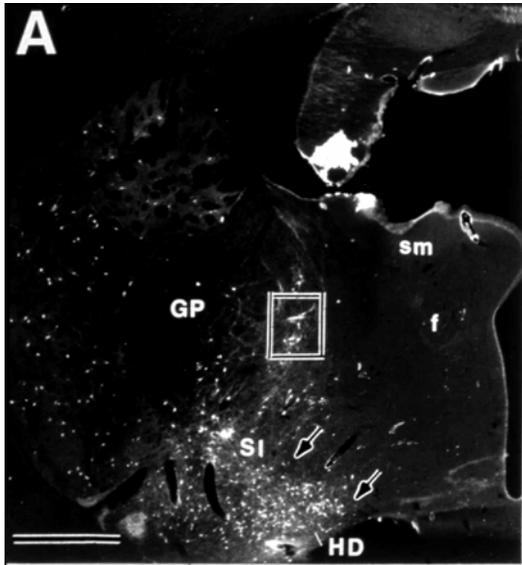


Cholinergic innervation in rat
(Lysakowski et al., 1988)

Comparing the ACh innervation of motor (4) and sensory regions of the rat, cat, monkey and human). From Avendano et al., 1996

CORTICOPETAL NEURONS IN THE B. FOREBRAIN OF RATS





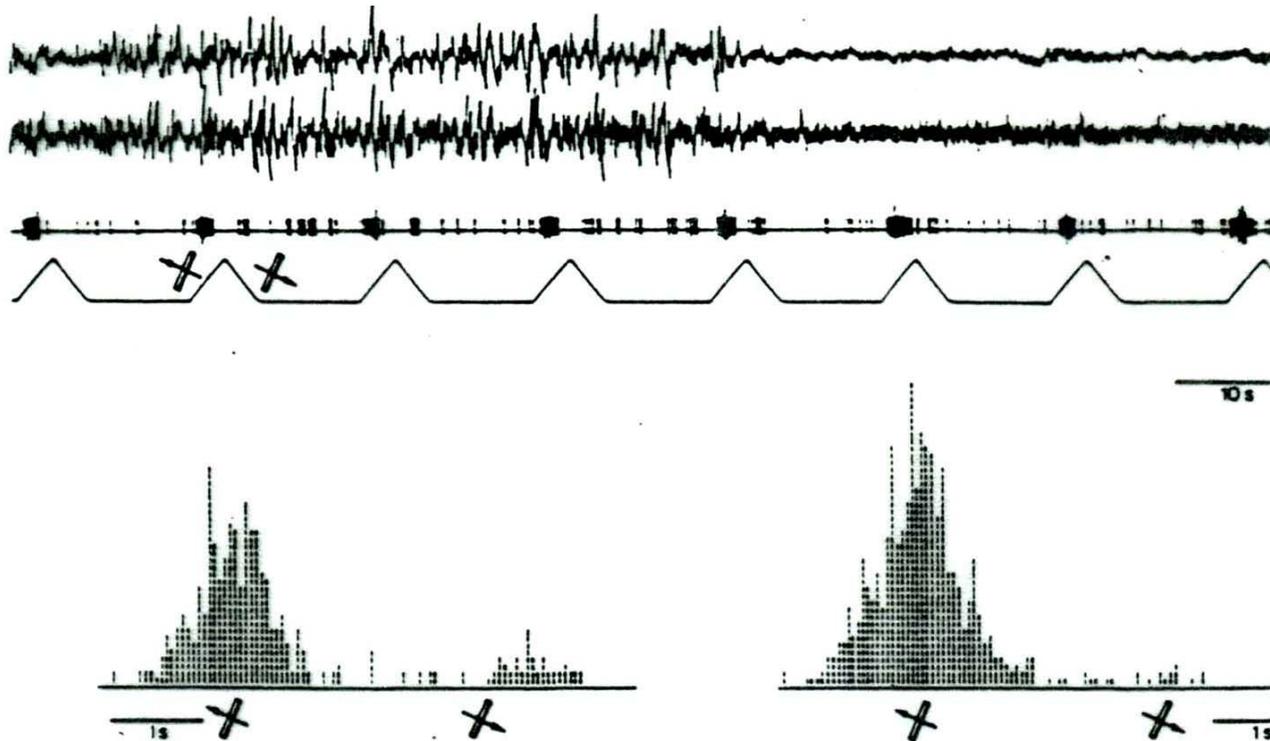
THE BASAL FOREBRAIN IN CORTICAL ACTIVATION AND SENSORY PROCESSING

- AROUSAL CONCEPTS
- FIRING PROPERTIES OF BF NEURONS
- DIFFUSE INPUT TO THE BF FROM ASC. MODULATORY SYSTEMS

- THE BF PROJECTION IS BIASED TOWARD ASSOCIATED CORTICAL AREAS
- FEEDBACK FROM THE PREFRONTAL CORTEX (PFC) TO THE BF
- DISTINCT OPERATION REQUIRES SPECIFIC ORGANIZATION
- WHERE THE BOTTOM UP MEETS WITH THE TOP DOWN?

- HYPOTHESIS: THE BF PARTICIPATE WITH THE PFC IN SELECTIVE AMPLIFICATION OF SENSORY PROCESSES

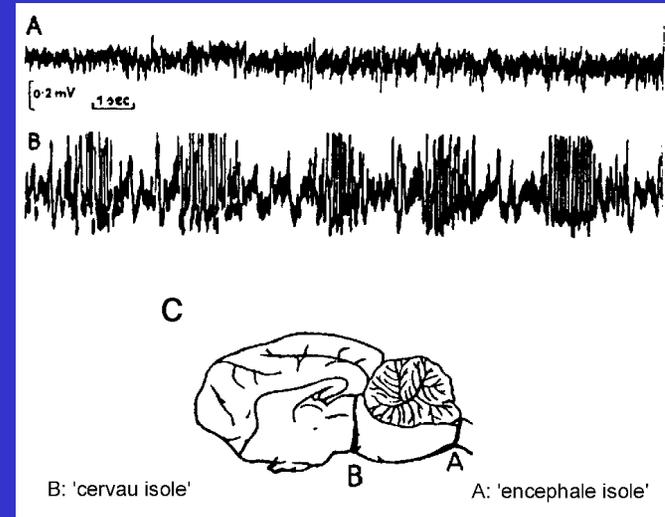
• **Arousal** is a complex state. It is characterized by: 1) switch from slow to fast oscillatory pattern in thalamo-cortical circuits; 2) increased discharge in neurons of the ‘diffuse’ corticothalamic systems. Arousal is a fundamental change of operation from the storage mode to information-processing mode, including perceptions influenced by external and internal sensory input; capacity of directing attention, accessing memory faithful to recent history; array of motor output; and emotions that are focused to percepts and thoughts.



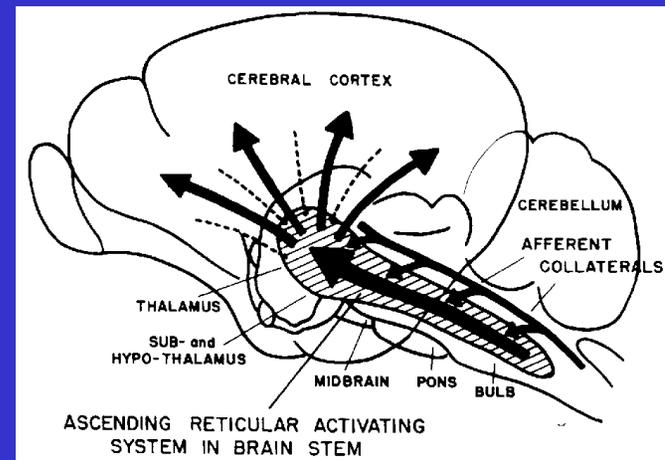
Effects of arousal (noise) from SWS on response selectivity of a cell in LII of the striate cortex. Arousal results in a moderate increase in the response to movement and a virtual elimination of the response to rightward movement (Livingston and Hubel, 1981).

DEVELOPMENT OF AROUSAL CONCEPTS

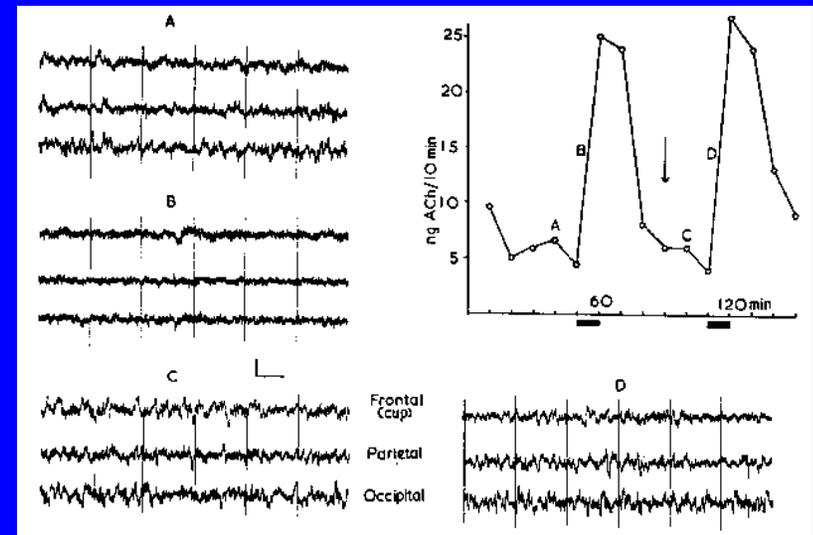
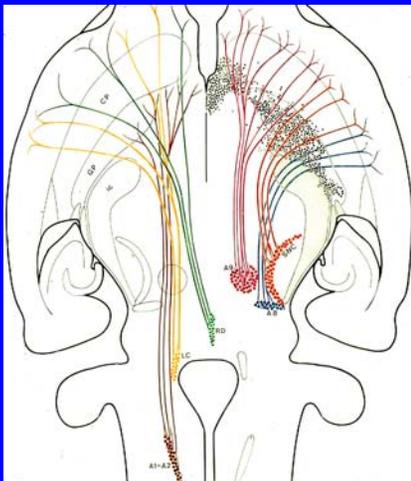
- **1930s:** wakefulness is maintained by afferent input to the brain and sleep ensues when that input is removed, as in the 'cerveau isole' cat, or falls below a certain critical level, as in normal sleeping (Bremer: deafferentation theory)



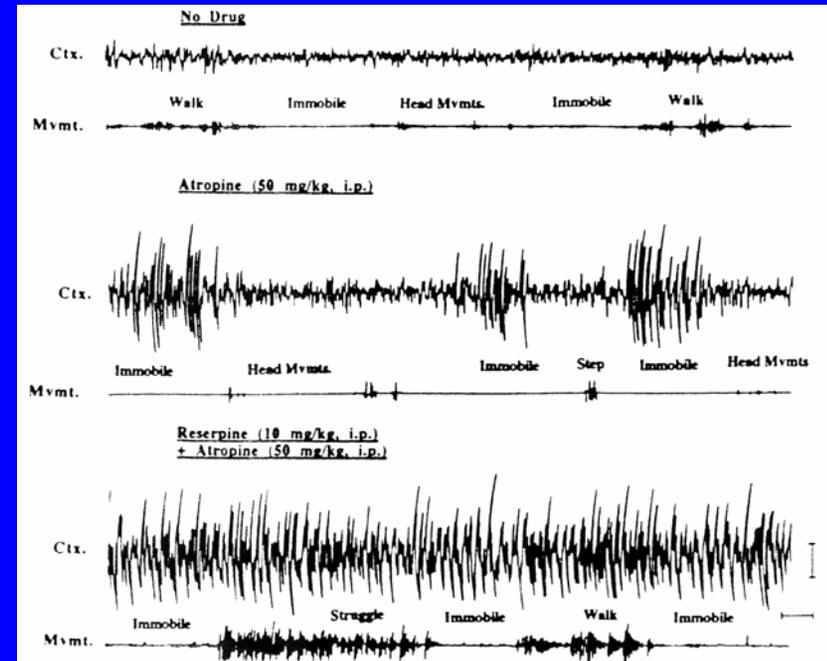
- **1950s:** the brain actively controls its own state by the ascending reticular activating system (Magoun-Moruzzi).



1965s- the concept of an undifferentiated reticular formation is gradually replaced by the description of transmitter-specific cell groups in the brainstem, the basal forebrain and the hypothalamus that send widely branching axons to the cerebral cortex and other parts of the brain. Role of acetylcholine and monoamines in cortical activation and sensory processing.



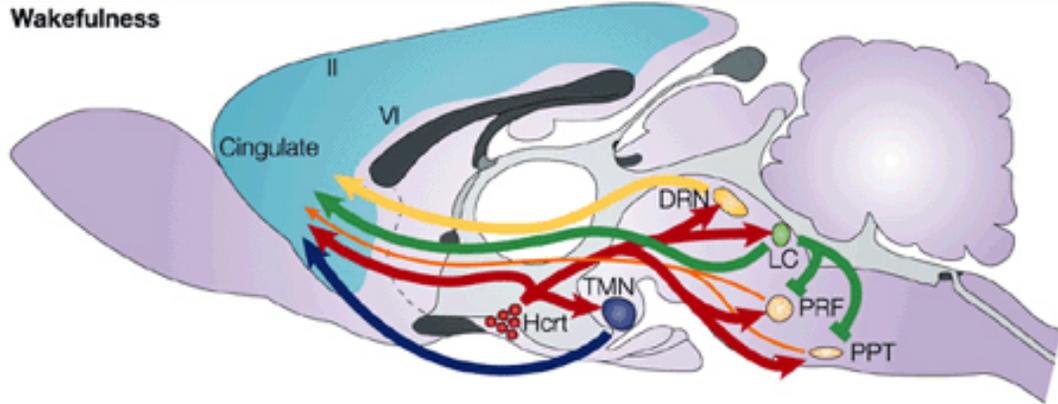
Kanai and Szerb, 1965



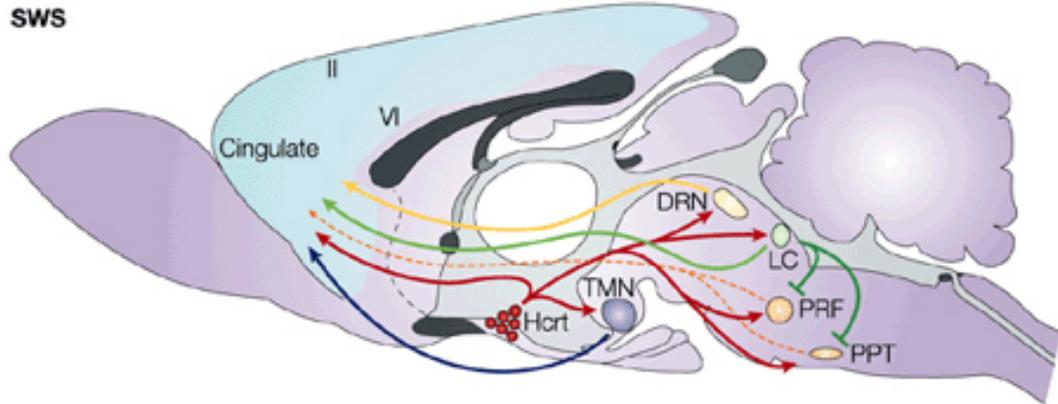
Vanderwolf and Dringenberg, 1998

DEVELOPMENTS OF AROUSAL CONCEPTS: Today

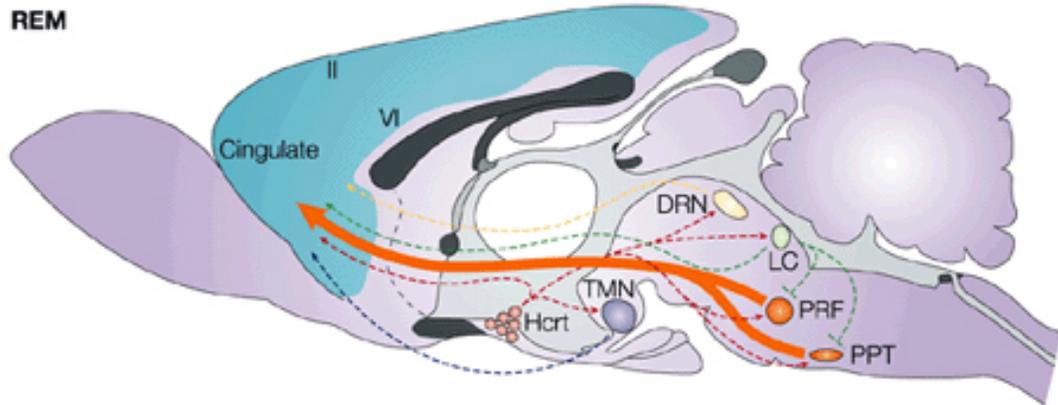
Wakefulness



SWS



REM



• **1990s**- various states can tentatively be linked to the accompanying systematic changes in balance between various neuromodulators.

DRN-5HT (yellow)

LC-NE (green)

PPT-Ach (orange)

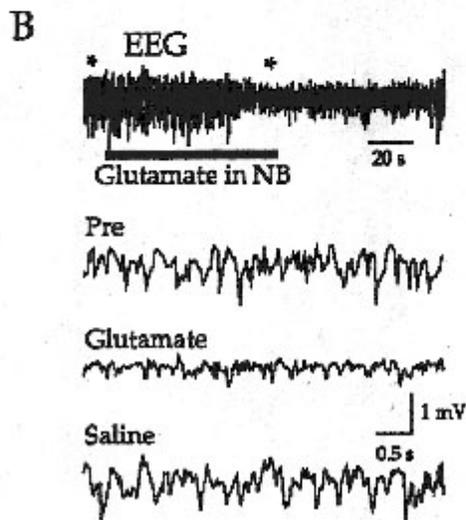
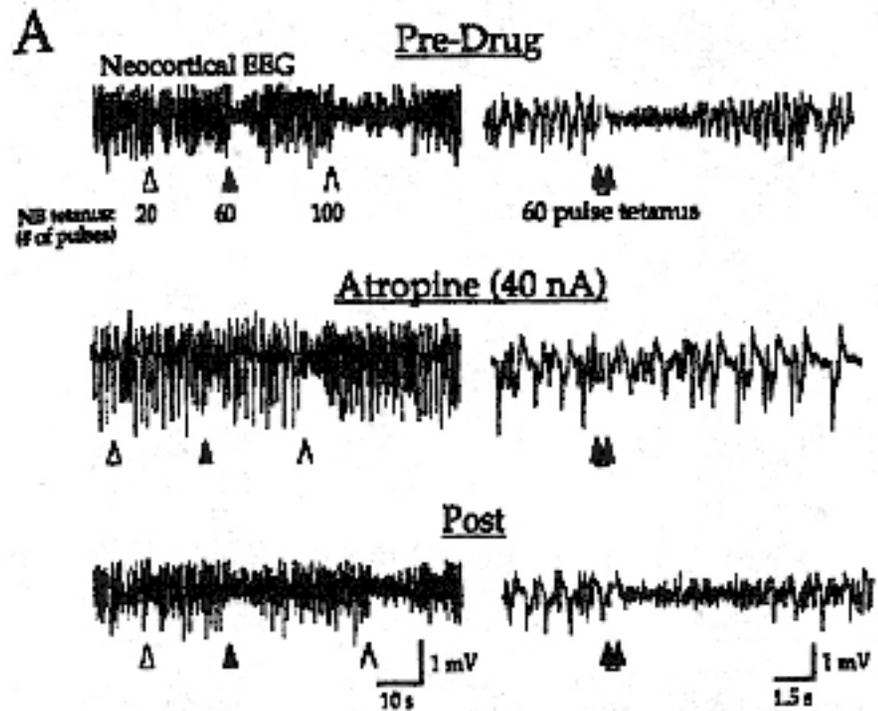
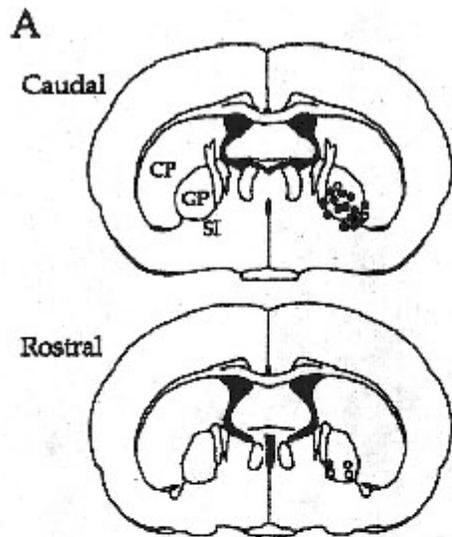
TMN-His

Hcrt/Orexin (red)

Sutcliffe and De Lecea, (2002)

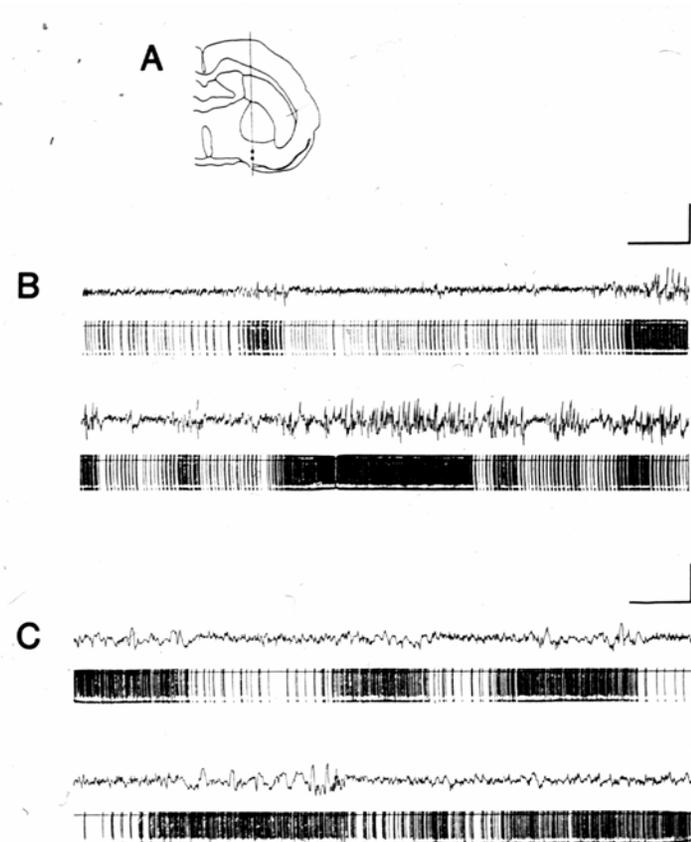
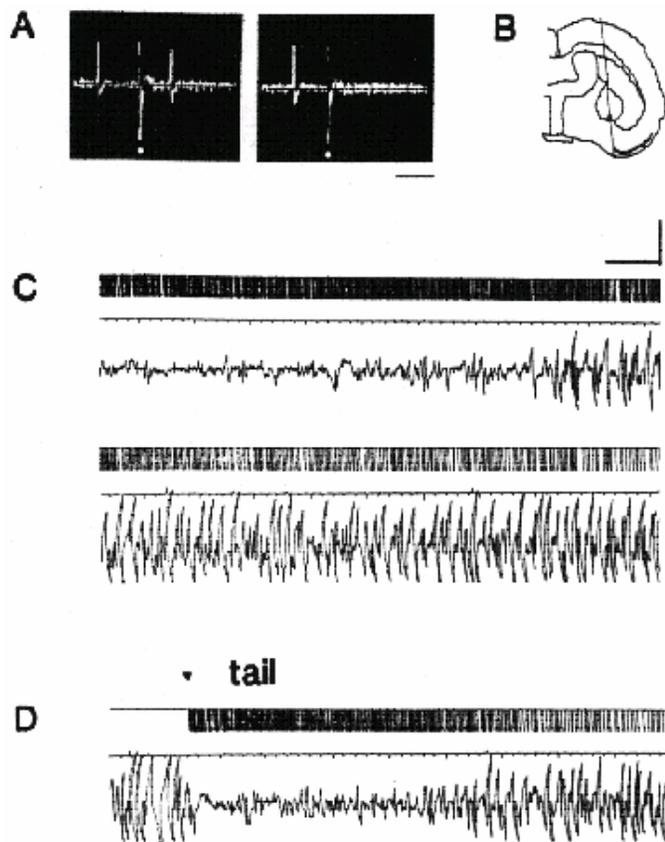
HOW ACTIVITY IN BASAL
FOREBRAIN NEURONS RELATE TO
CORTICAL ACTIVATION?

BF stimulation, EEG, Ach action in vitro

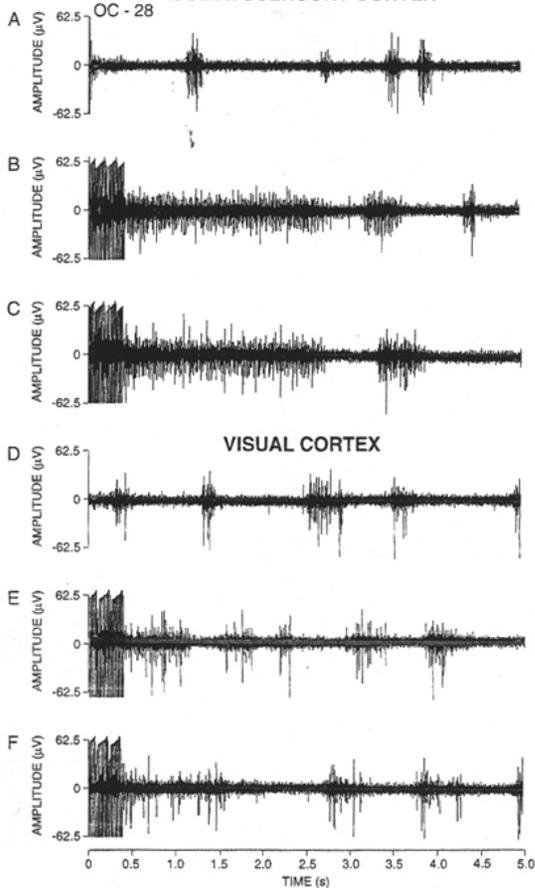


Metherate and Ashe, 1992

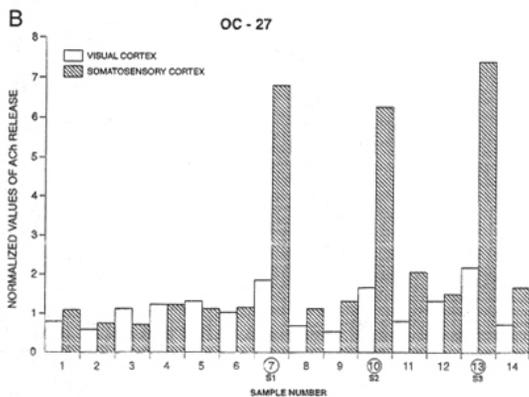
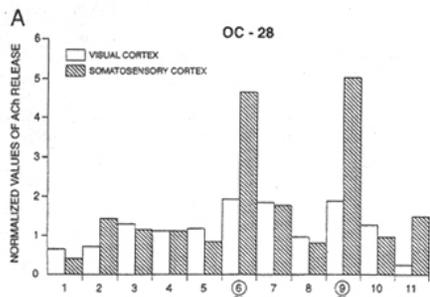
BF neurons fire in correlation to various EEG epochs.



SOMATOSENSORY CORTEX

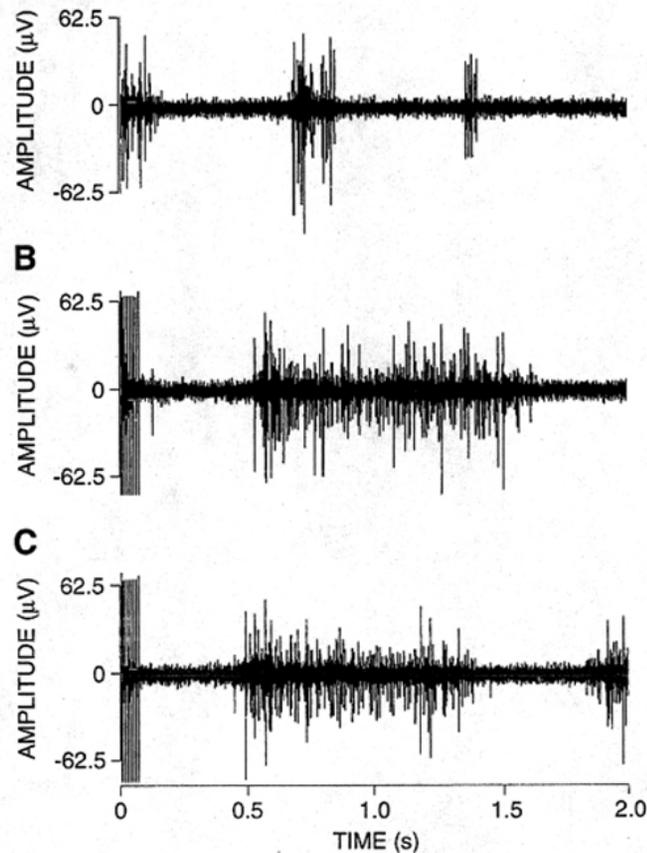


An example of the effects of BF stimulation on the cerebral cortex elicited from a region of the BF that produced increased ACh release from somatosensory cortex. **A:** Spontaneous activity. **B:** Effect of a 400-ms train of pulses at 100 Hz in the BF. **C:** A second example of the response in the somatosensory cortex. **D:** Spontaneous activity in the visual cortex. **E:** The effect of BF stimulation in the visual cortex from the same site that produced the record in (B). **F:** A second example of the response to BF stimulation in the visual cortex. The release of ACh from the visual cortex was much less than from the somatosensory cortex.



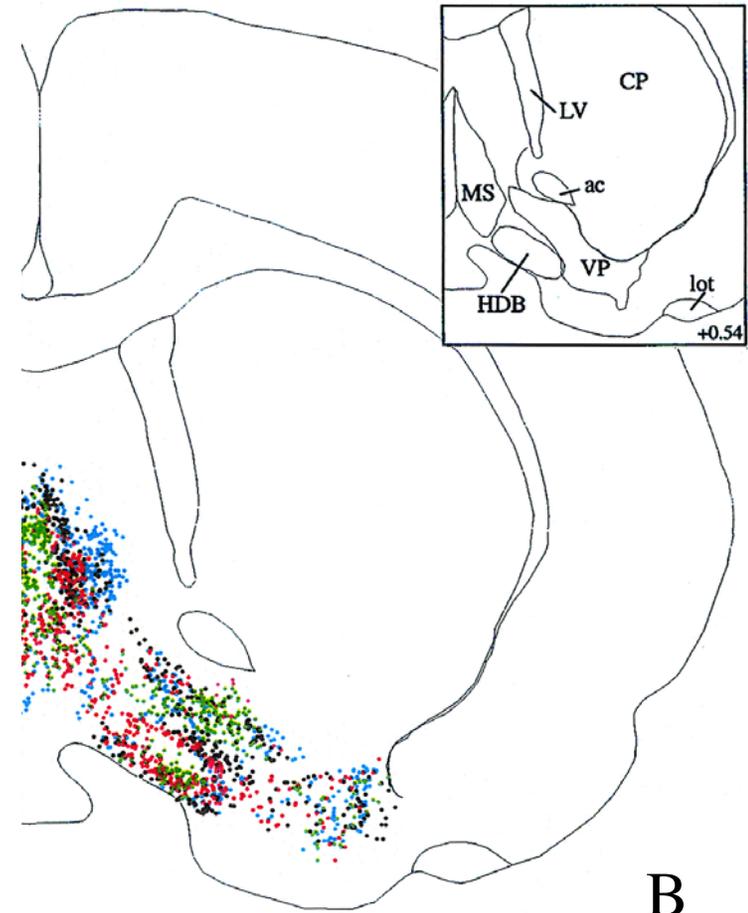
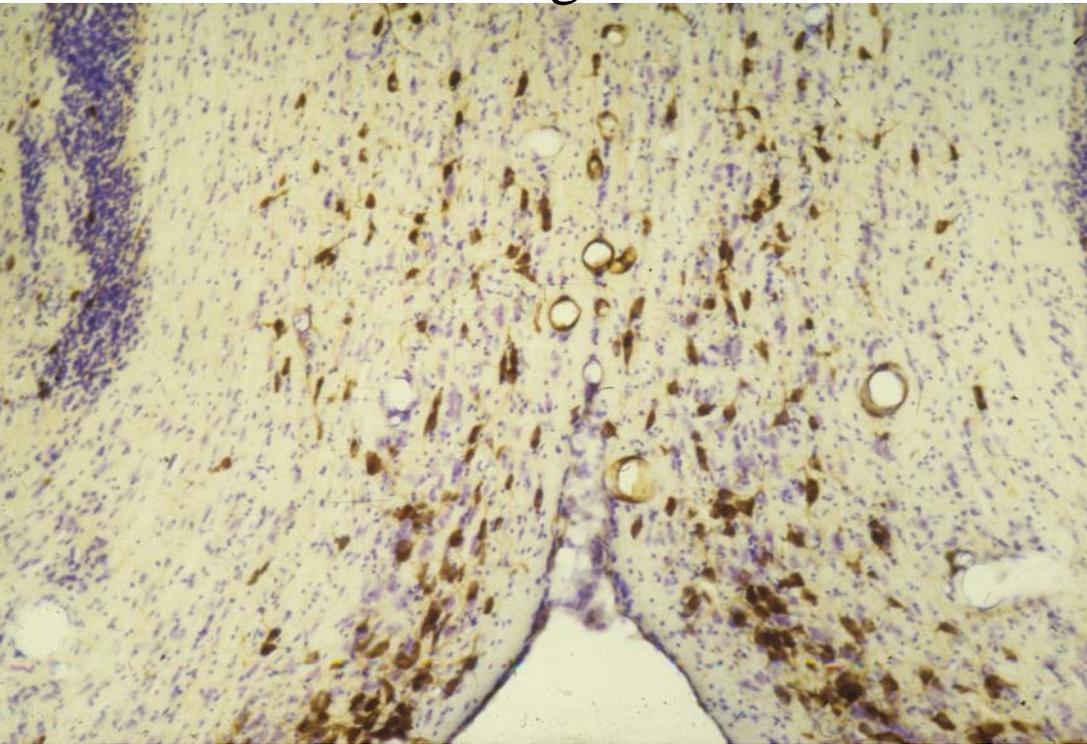
ACh release from sites in the visual and the somatosensory cortex following electrophysiological recordings of the neuronal activity provoked by BF stimulation. **A:** Eleven pairs of 30-minute samples obtained simultaneously from the somatosensory and the visual cortex. The first five pairs provided the baseline levels for both cortical regions. During collection of the sixth pair of samples the BF was stimulated every 4 seconds with a train of 40 pulses at 100 Hz. ACh release increased more in the somatosensory cortex than in the visual cortex. ACh levels returned to normal over the next hour, and the stimulation was repeated during the ninth sampling period. **B:** A second experiment showing the same pattern of differential ACh release following BF stimulation. In this experiment the BF stimulation was performed three times using a train of 50 pulses every 4 seconds for the duration of the 30-minute sampling period (sample pairs 7, 10, and 13). A slight, but significant, increase in ACh occurred in the visual cortex, whereas a 600–700% increase occurred in the somatosensory cortex.

SE - 27 SOMATOSENSORY CORTEX



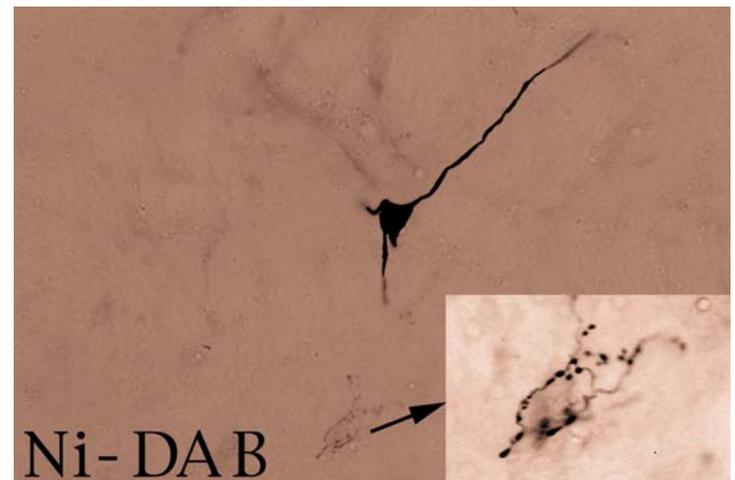
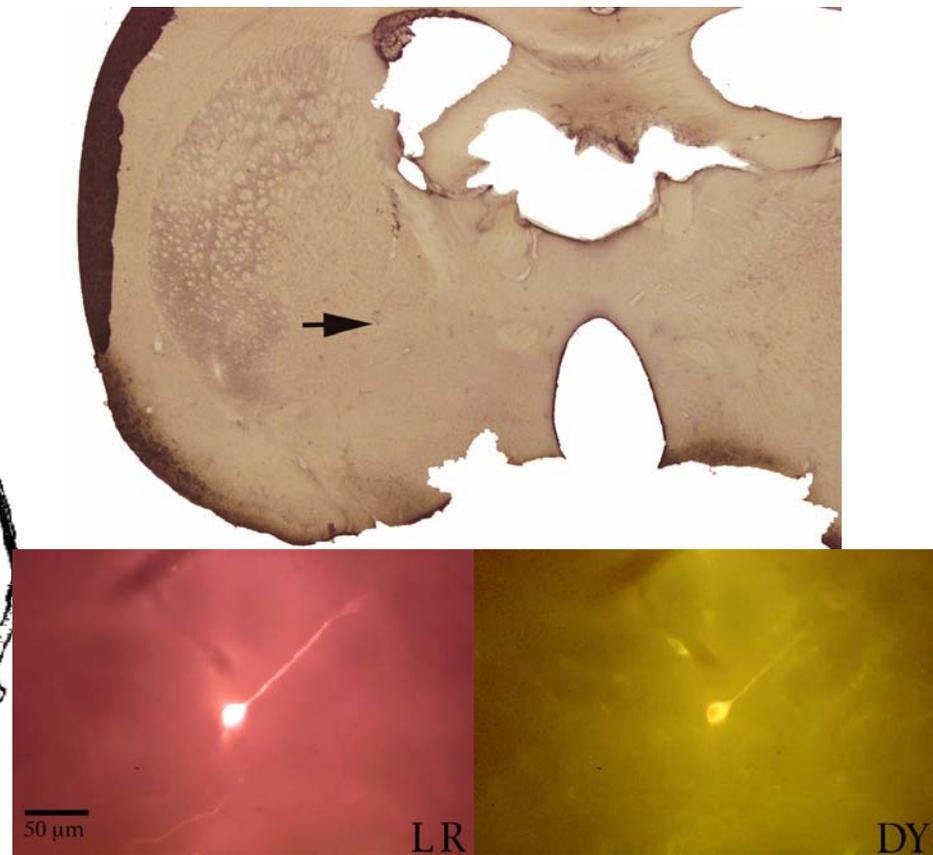
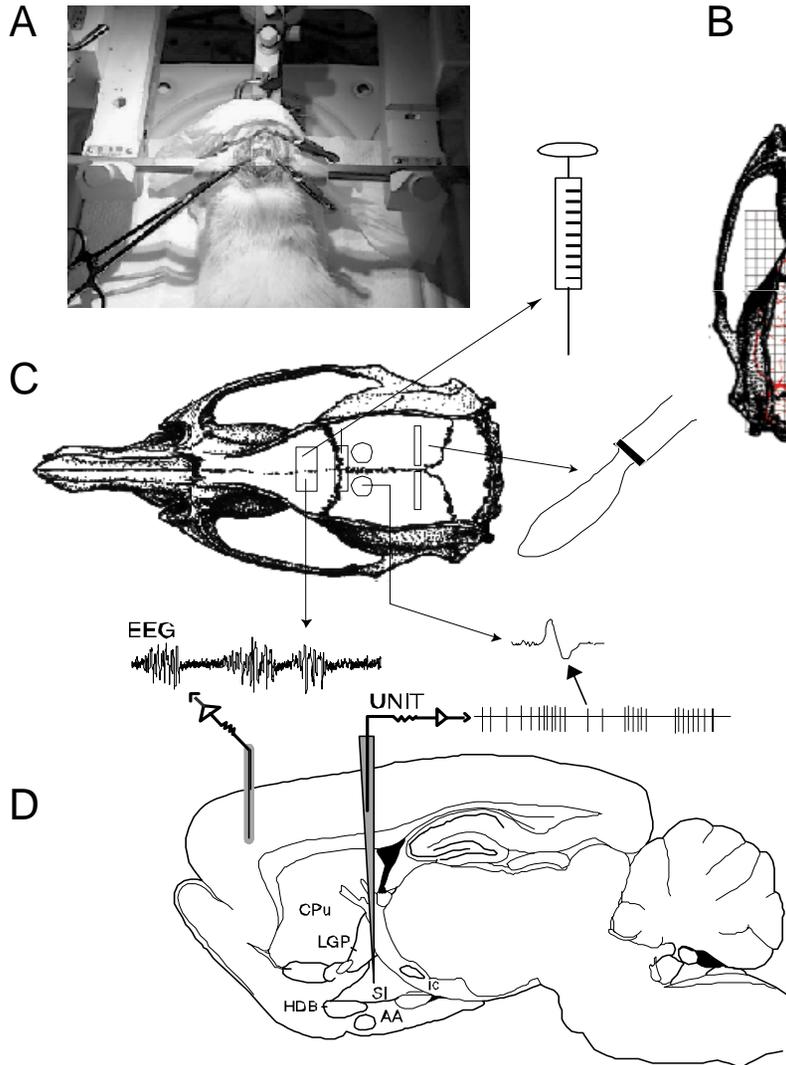
Multiunit recordings from the hindlimb somatosensory cortex showing how BF stimulation could provoke a long suppression of spontaneous activity followed by a large and long increase in neural activity. The several large neurons in this recording were spontaneously active. **A:** A 2-second record of spontaneously active neurons discharging in bursts. **B:** When a 100-ms BF stimulus was administered, all neural activity stopped for about 450 ms after the end of the stimulus; then an intense discharge occurred, lasting for approximately 1.1 second. **C:** A second example of the inhibitory pause produced by BF stimulation.

Basal forebrain corticopetal cholinergic neurons are intermingled with non-cholinergic neurons

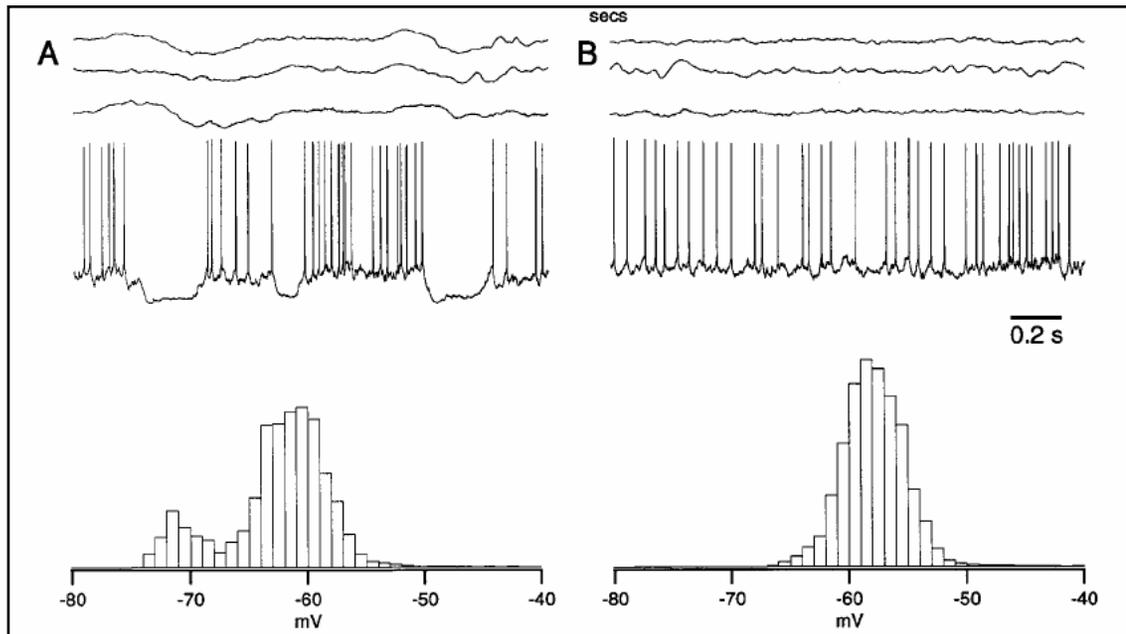
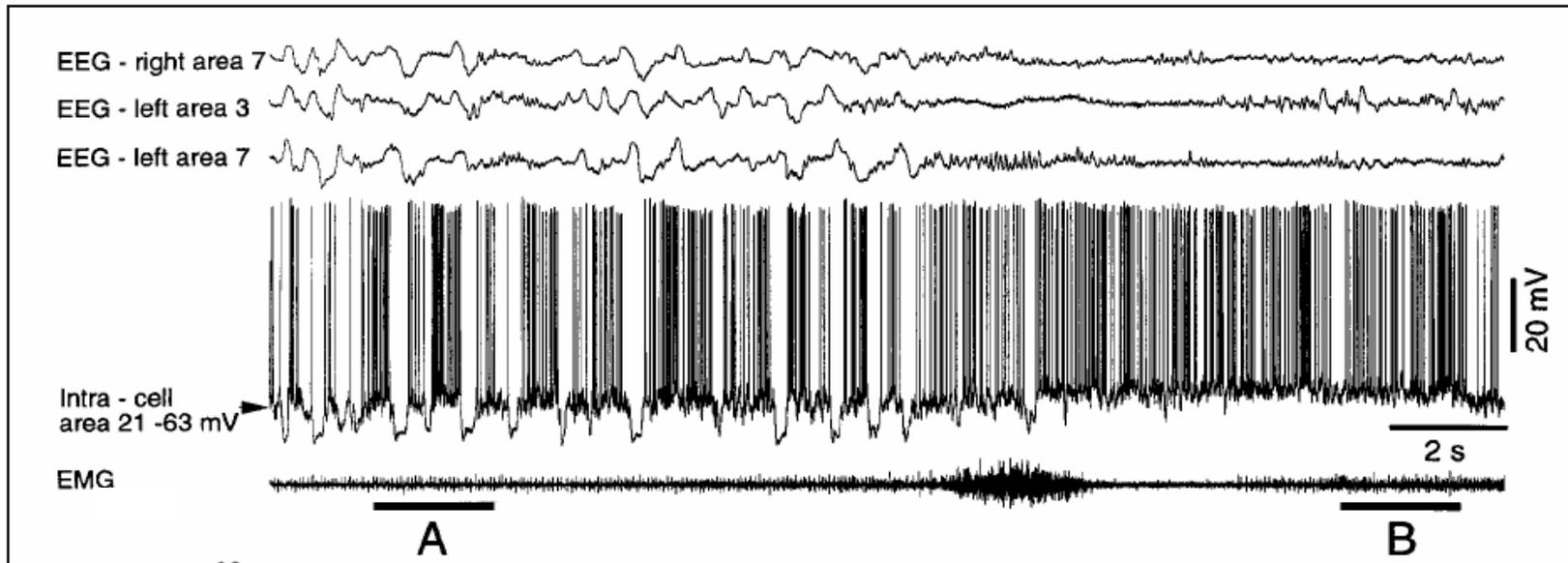


A: Choline acetyltransferase (ChAT) and Nissl-staining. B: cholinergic neurons (red) are intermingled with Parvalbumin (green), Calretinin (black) and calbindin (blue) neurons

Juxtacellular filling, transmitter or target identification of the recorded neurons, EEG monitoring. The methods



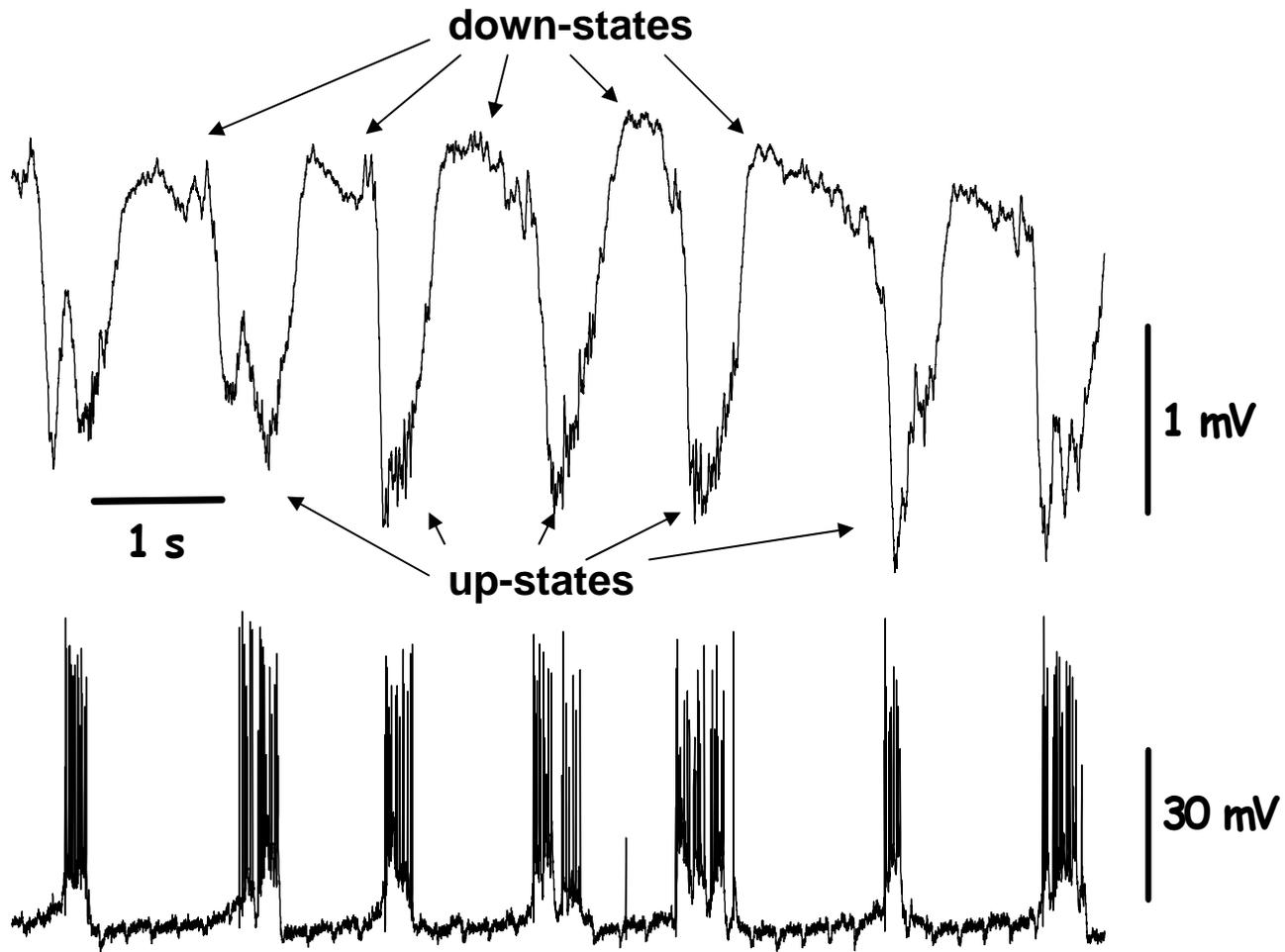
CORTICAL UP AND DOWN STATES



unanesthetized cat

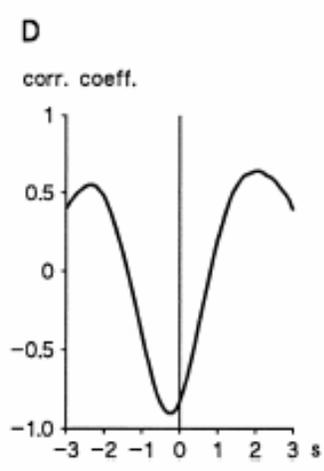
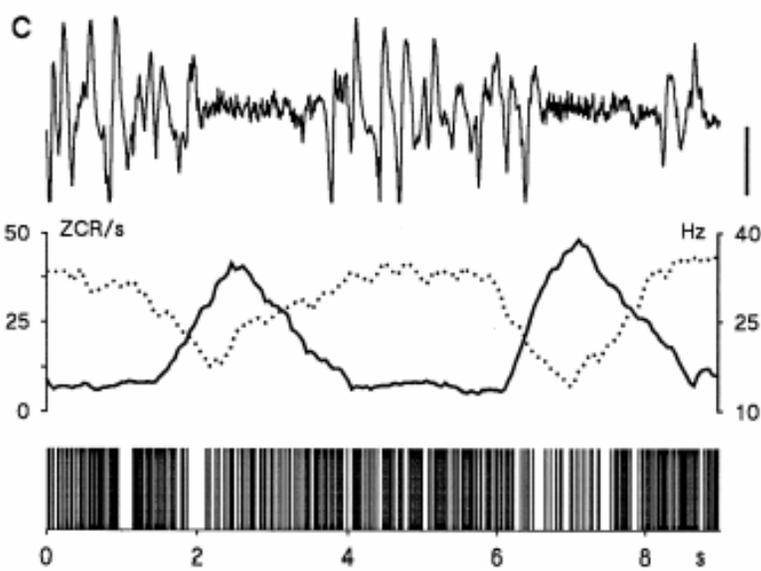
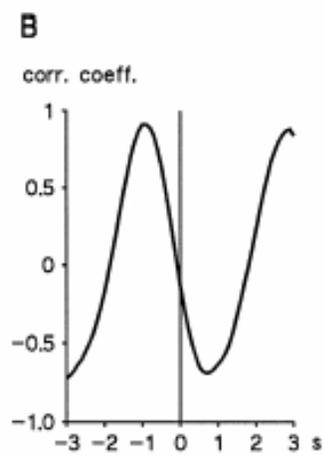
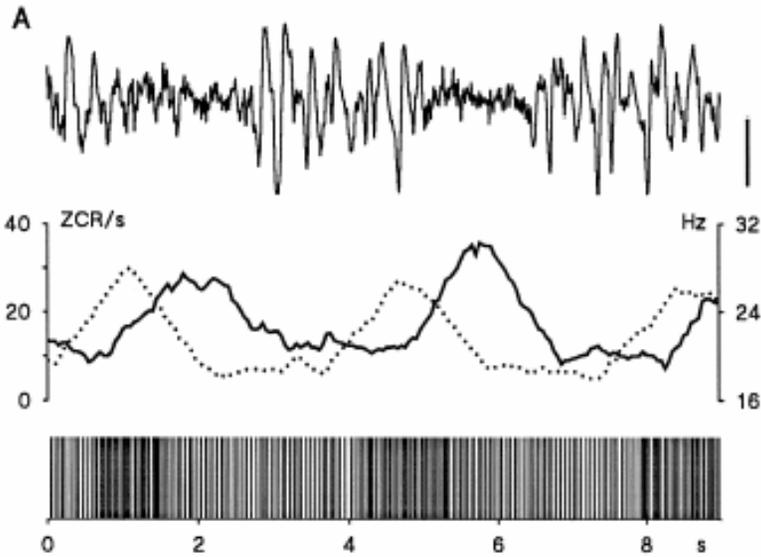
Transitions from SWS to wakefulness. Brief periods of high-frequency firing during SWS are interspersed with silent periods corresponding to the hyperpolarizing phase of the slow oscillation. EEG is characterized by bipolar deflections corresponding to bimodal distribution of membranepotential.

Fig. 9 from Steriade et al., (2001)



intracellular recording from a pyramidal cell of a urethane-anesthetized rat and the simultaneous EEG recording with transcortical electrodes from the other hemisphere

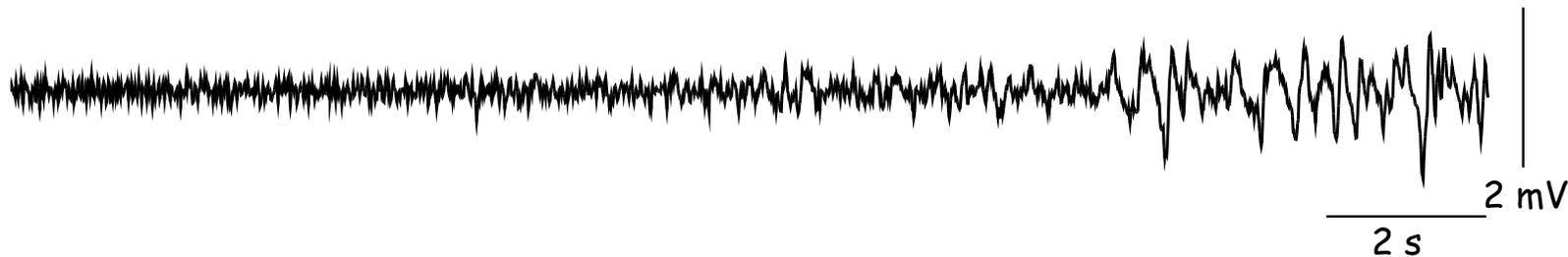
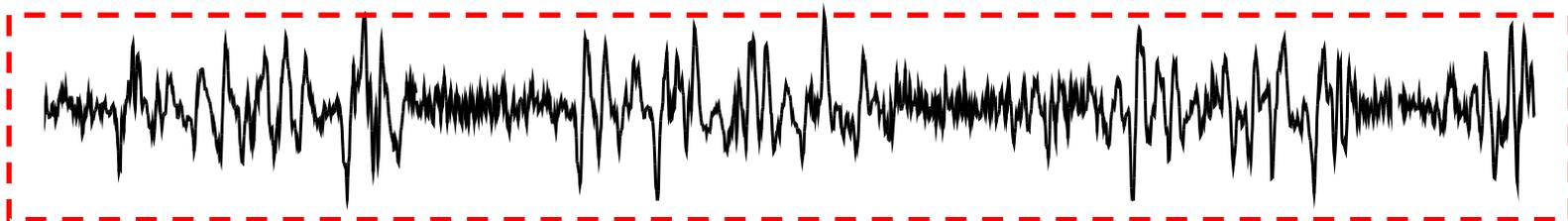
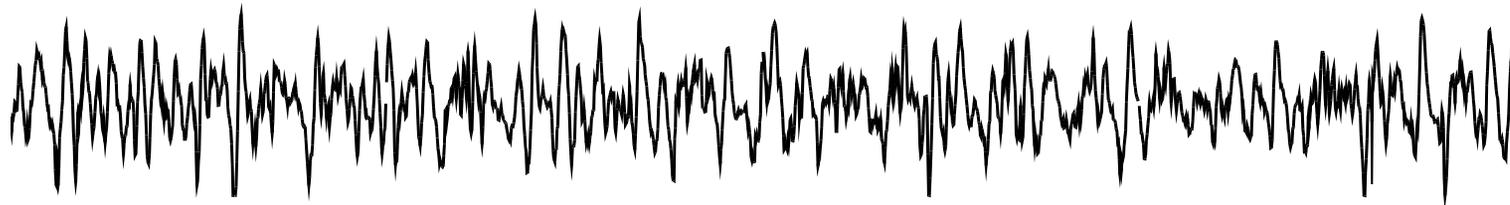
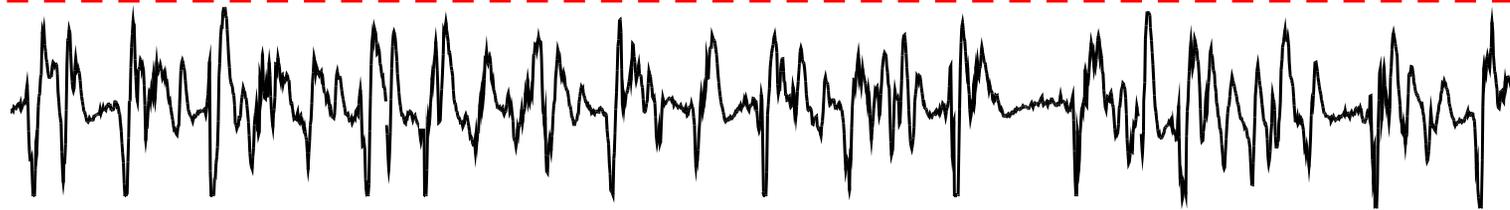
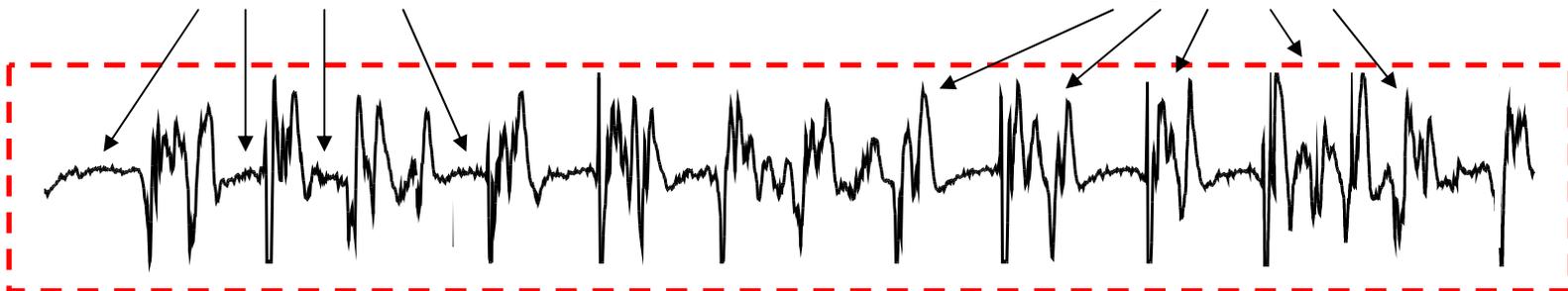
during the **up-states**, the cell is firing intensively and there are higher frequency waves on the EEG
during **down-states**, EEG is a thin line only and the intracellular recording is noisy and with ECG artefacts (Detari, unpublished observation)



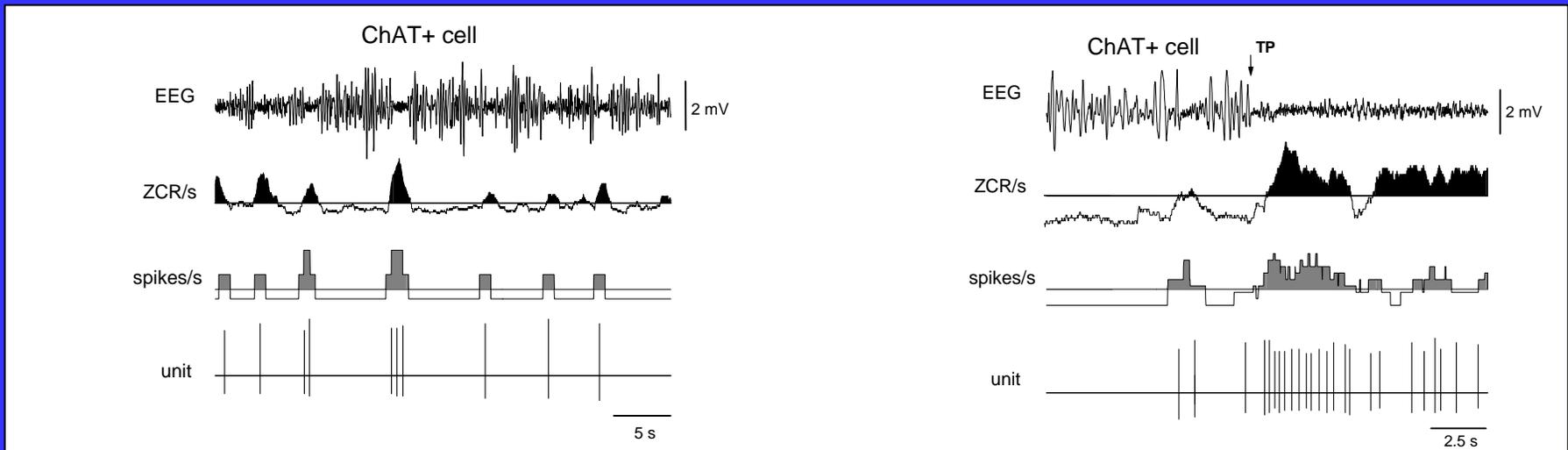
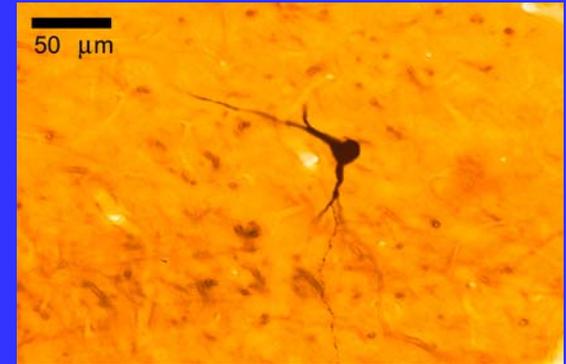
Timing of changes in EEG and neuronal firing rates for an F- (A) and an S-cell C . Top trace: EEG positivity at the deep electrode is upward, calibration 1 mV ; middle traces: frequency of zero-crossings ZCRrs, solid line in EEG and neuronal firing rate dotted line ; bottom trace: computer-generated ticks marking the occurrence of neuronal spikes. The strong positive (A) and negative (C) correlations between ZCR and firing rate can be seen in the correlation coefficients (B and D), peak of 0.91 and y 0.90 at -900 and -200 ms, respectively calculated by shifting the curves in 100 ms steps between -3 and 3 s. (From Detari et al., 1997)

down-states

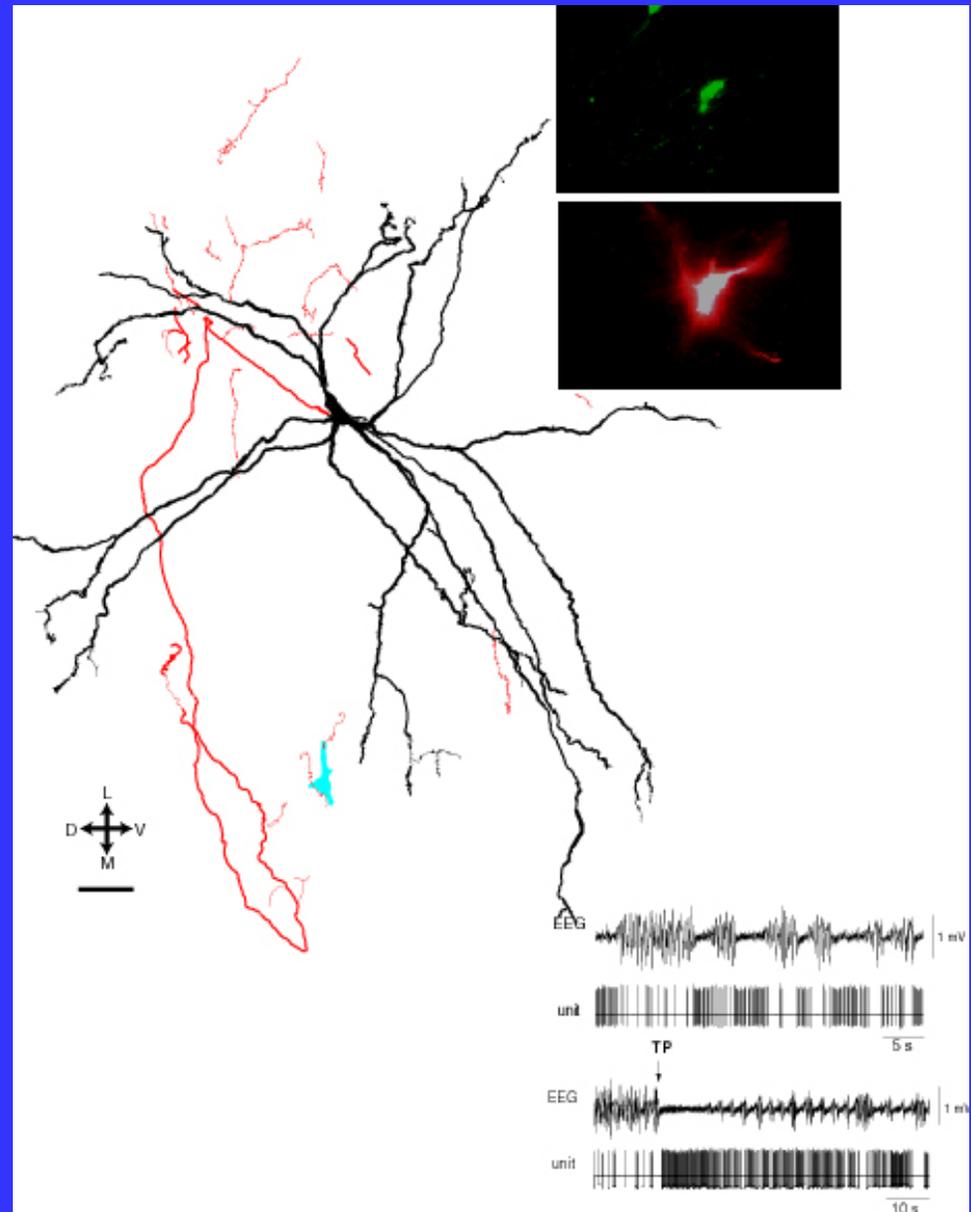
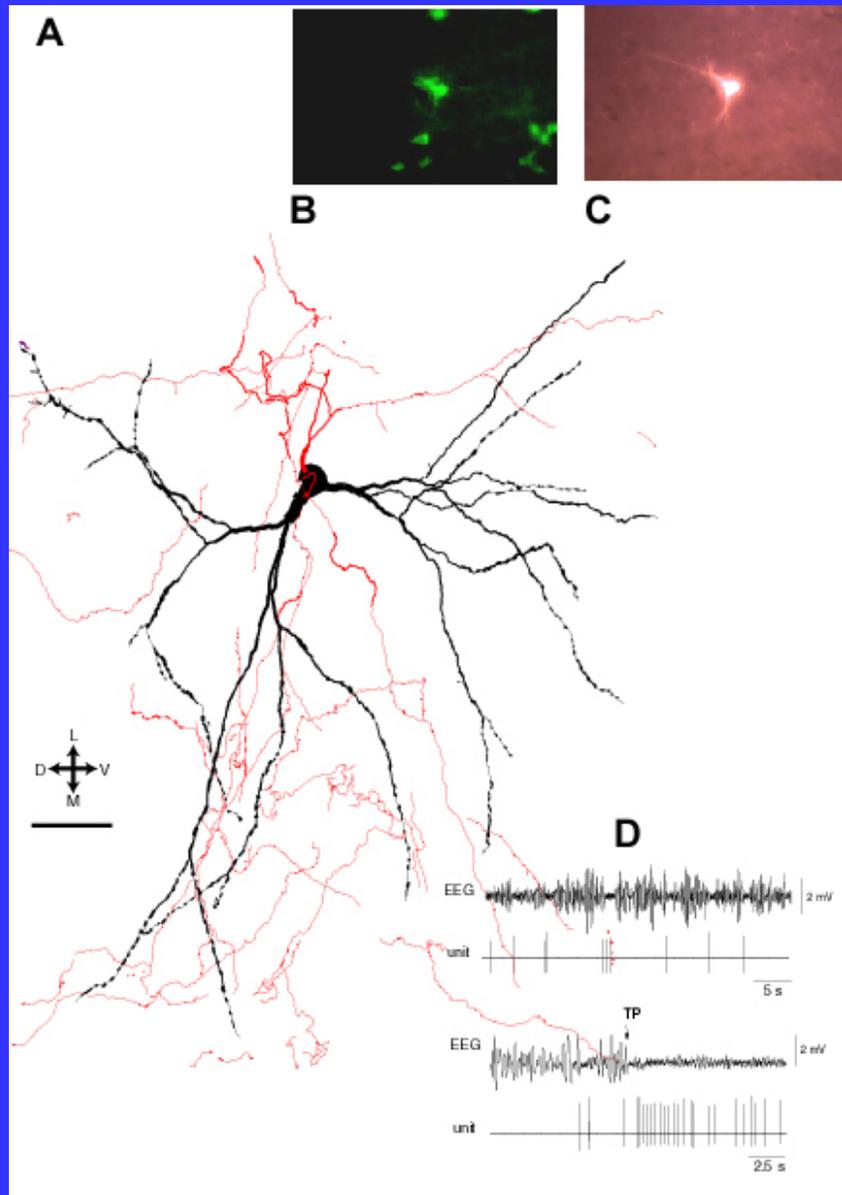
up-states



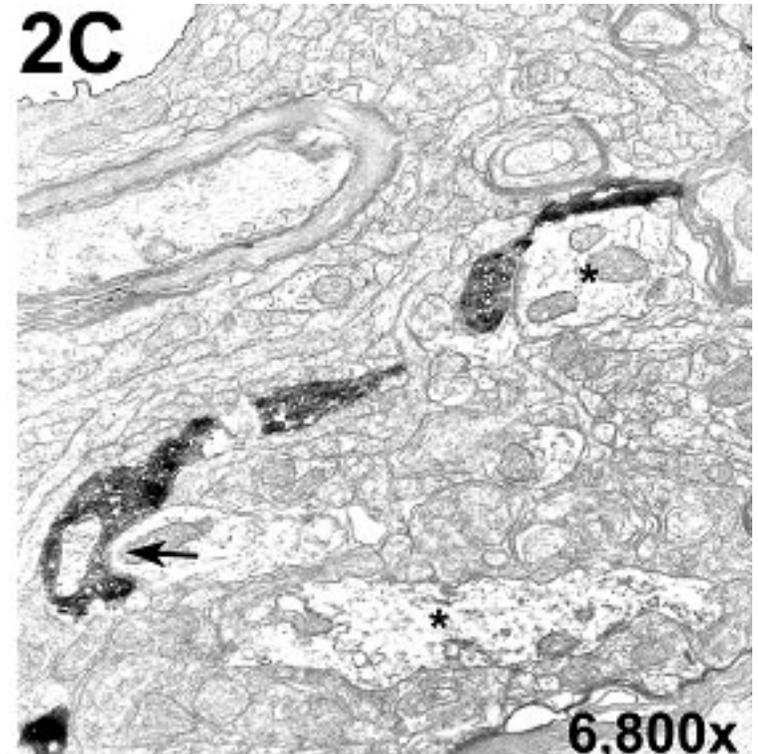
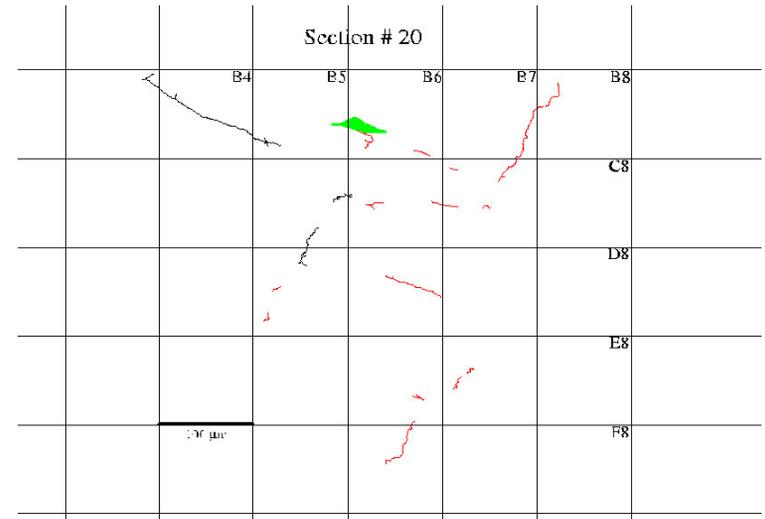
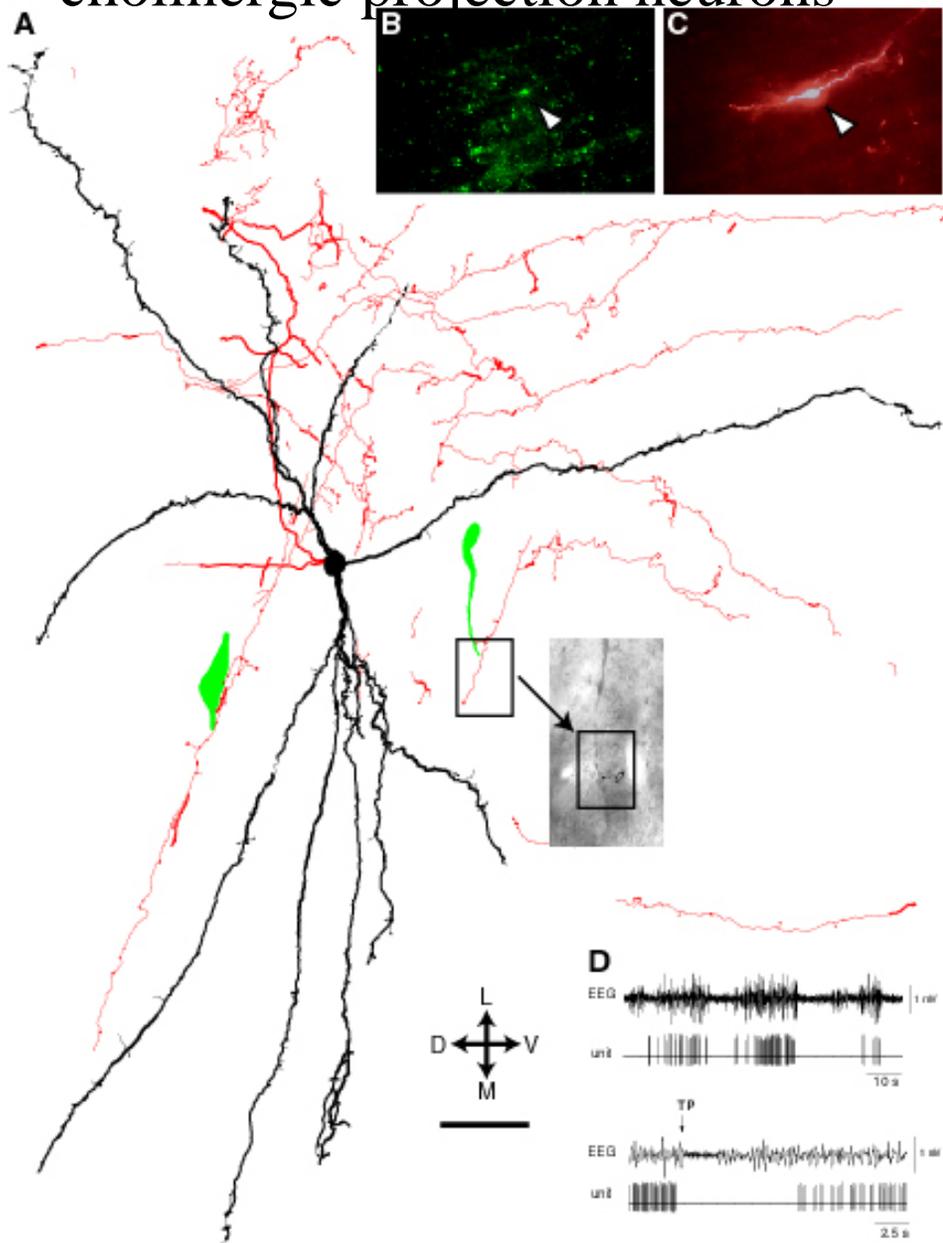
Identified cholinergic cell



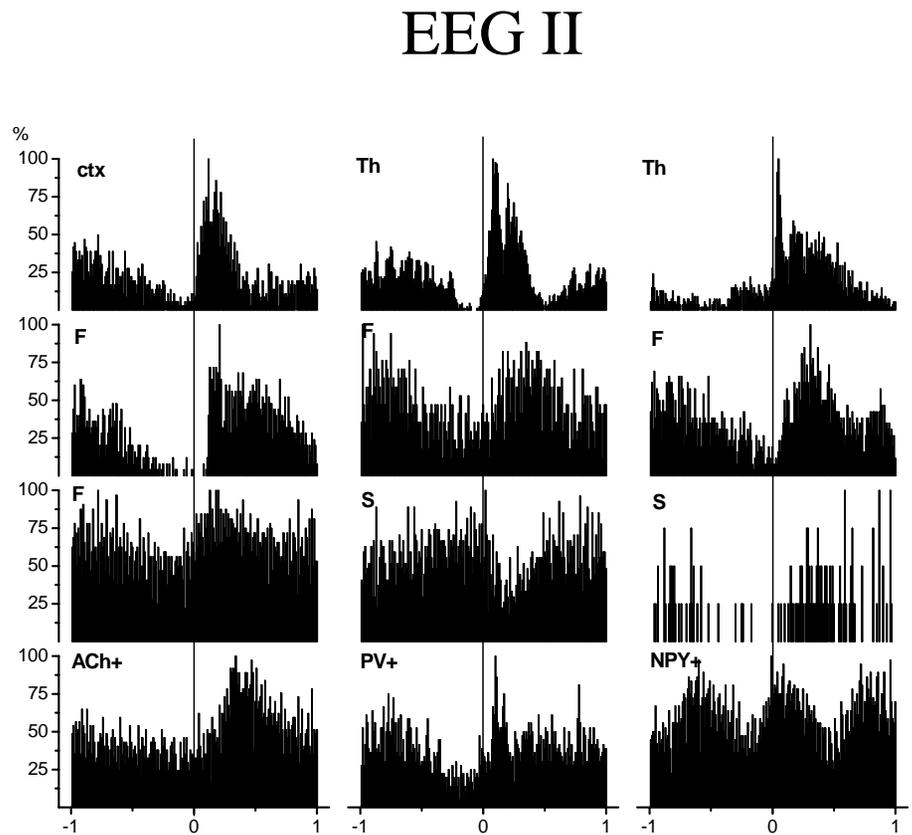
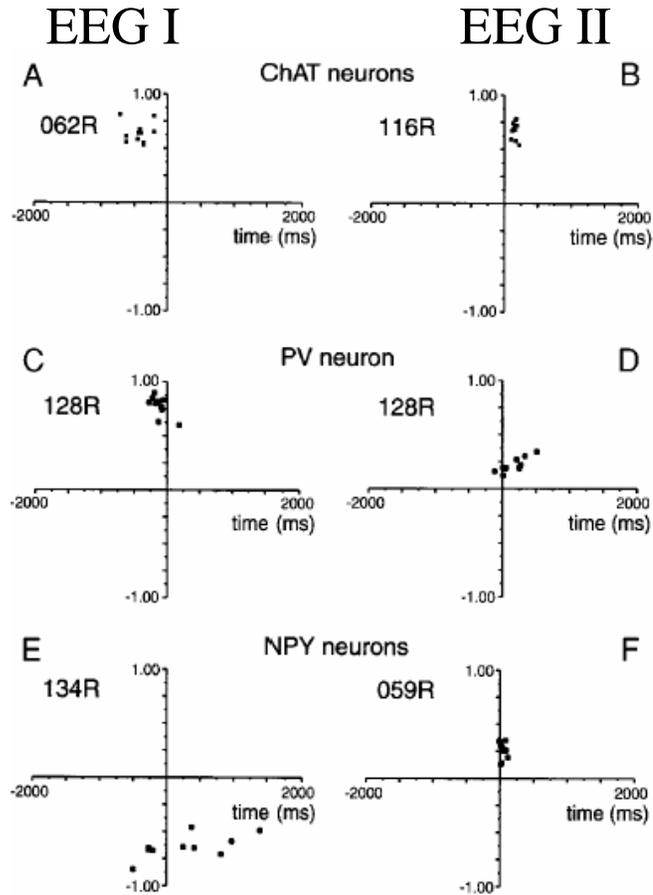
Identified Cholinergic and Parvalbumin (GABA) projection neurons



Identified NPY-containing local interneuron and its input to cholinergic projection neurons



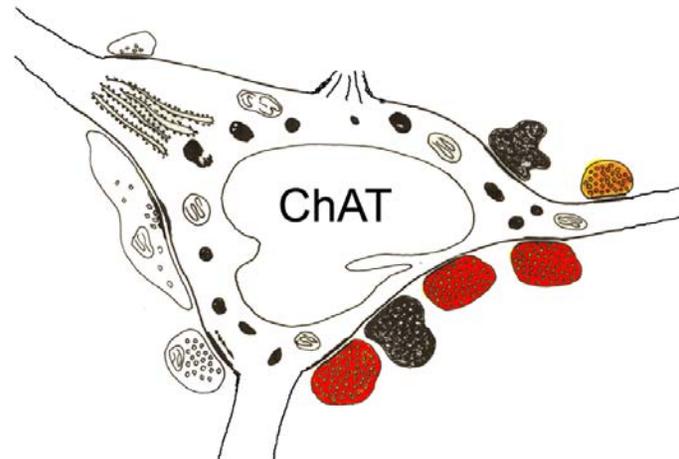
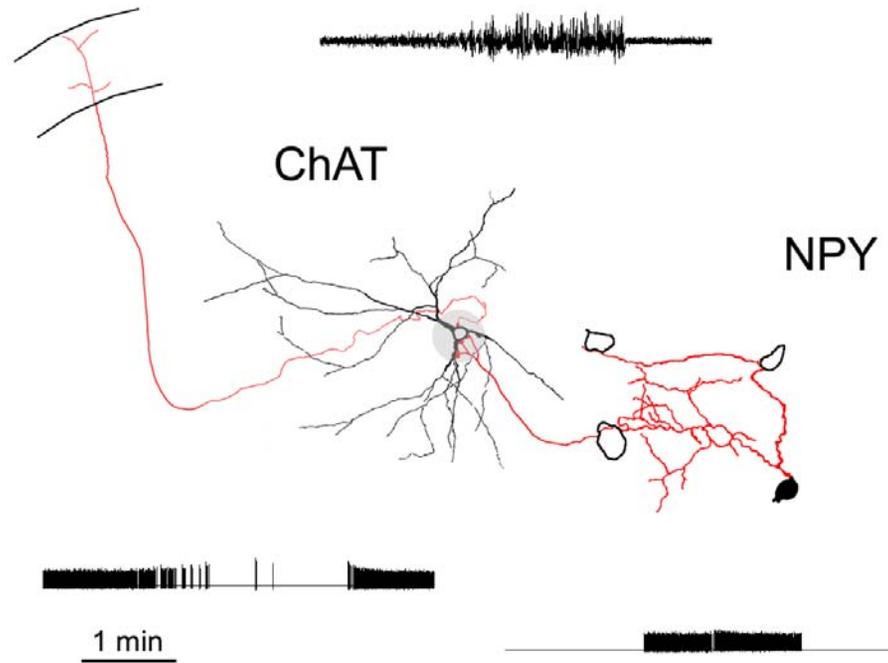
TIMING OF CHANGES IN EEG AND BF UNIT FIRING RATE



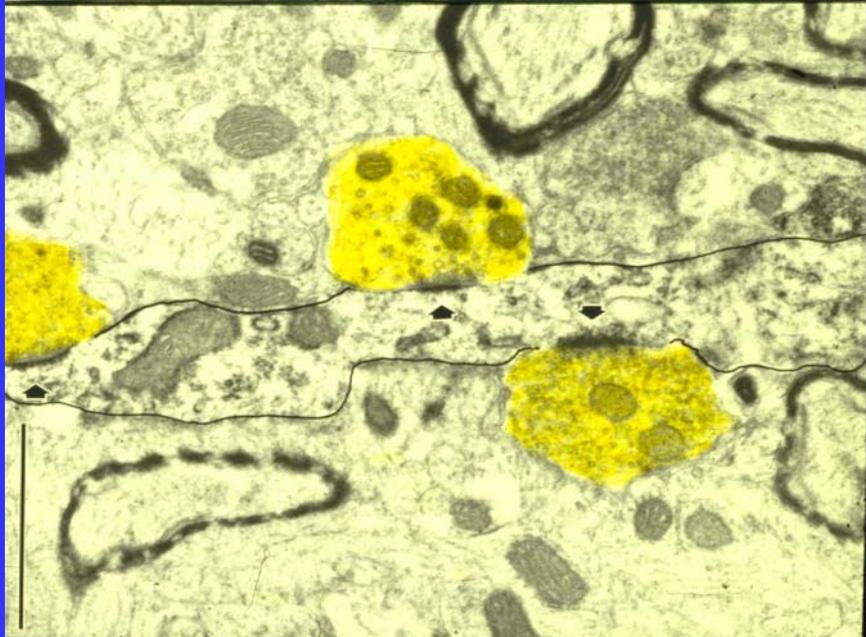
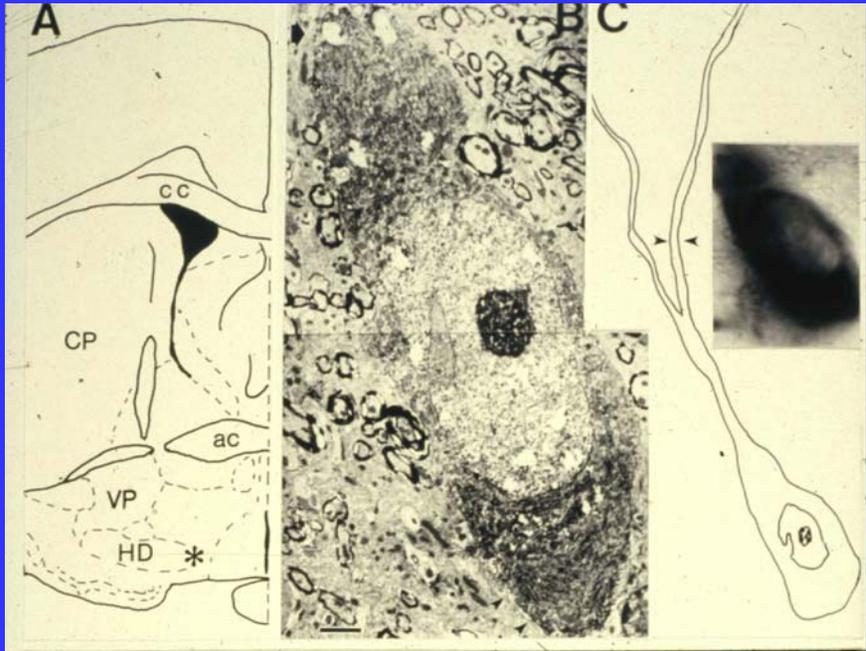
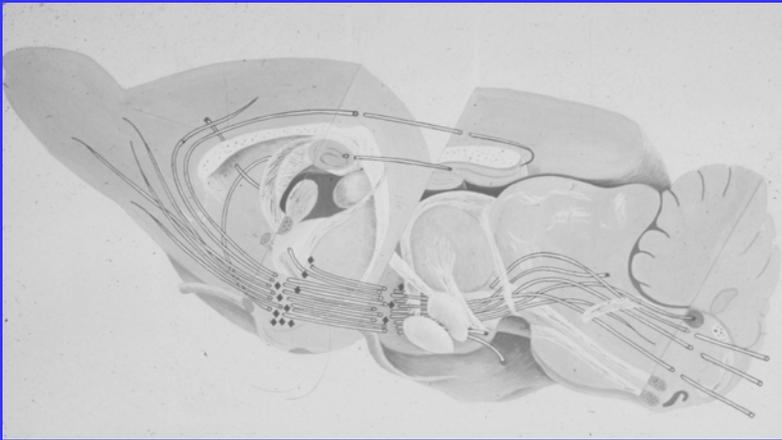
A: change in unit firing precedes the change in EEG pattern (median: **-405 ms**). **B:** changes in unit firing follow the EEG changes (median **159 ms**). **C:** the correlation indicates that the increased firing precedes (median **-153 ms**) the change in EEG- I (C) while it lags (median **210 ms**) the EEG- II (D). **E** because the points are in both the left and the right side of the lower quadrants, the correlation indicates that the unit activity during EEG pattern I can either precede or lag (median 87 ms) the changes in EEG activity but that in both cases the firing rate is increased during the EEG slow wave activity. **F:** during EEG- II, the NPY cell firing occurs lagging the EEG changes (median **40 ms**). Duque et al., 2000

Histograms of neuronal activity around the fast transitions (vertical line; ± 1 s) between the down-, and up-states marked in the cortical EEG. Different number of transitions were averaged, then normalized for the maximal value to construct the histograms. Immediate excitation was obvious in thalamic (Th) and cortical cells (ctx), while the response was usually delayed in both fast-wave-active (F-cell), and slow-wave-active (S-cell) neurons in the BF. Activation in the PV+ and NPY+ neurons (last row) had much sorter latency then the excitation in the ACh+ cell.

BF CIRCUITRY: A PRELIMINARY IDEA

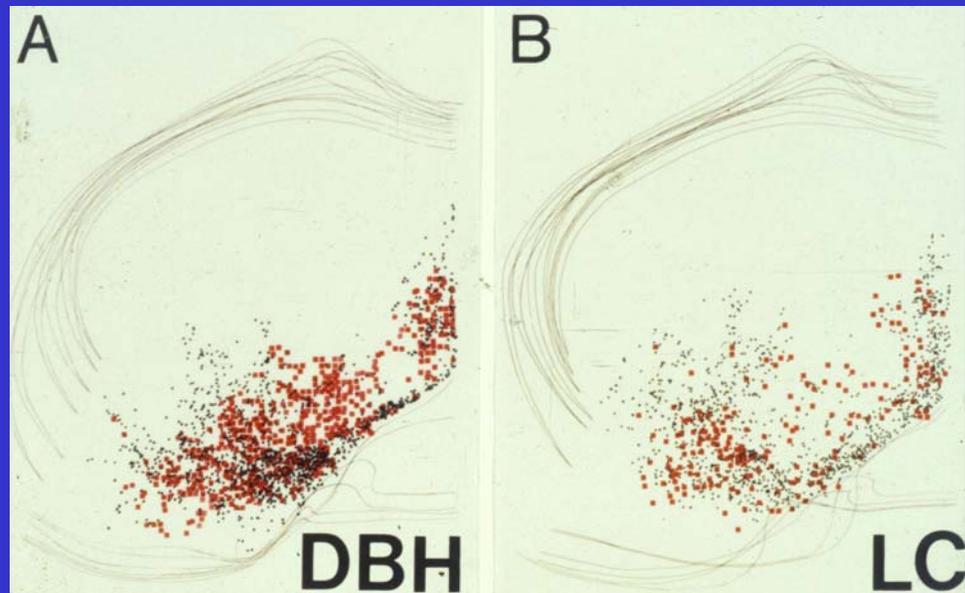
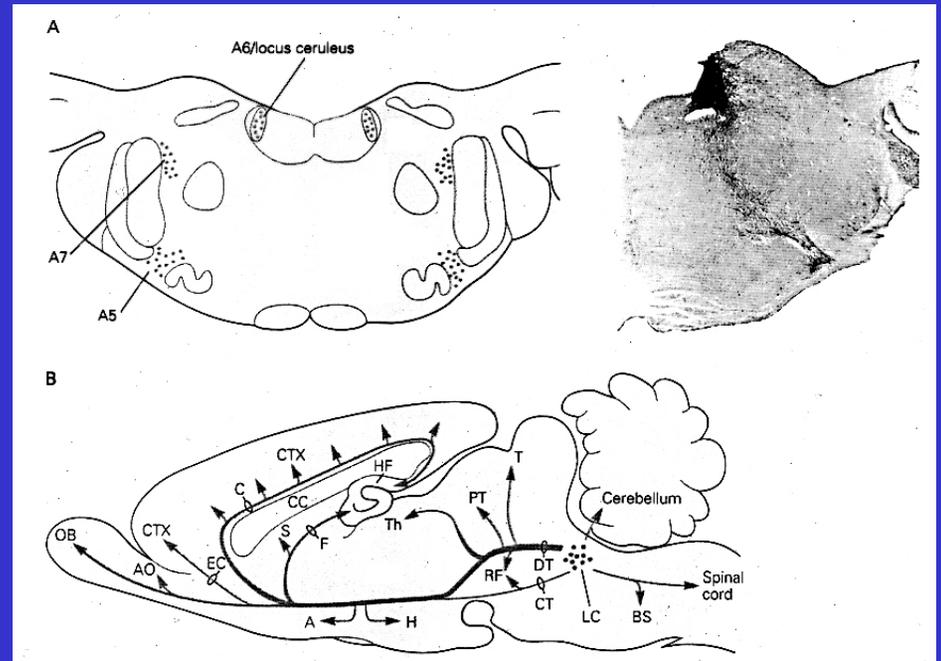
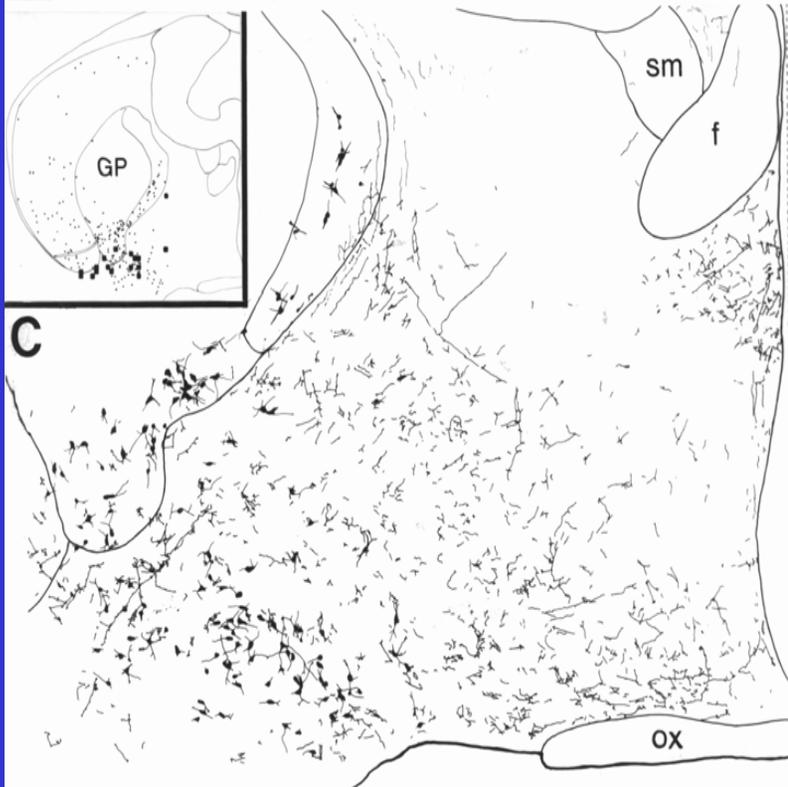
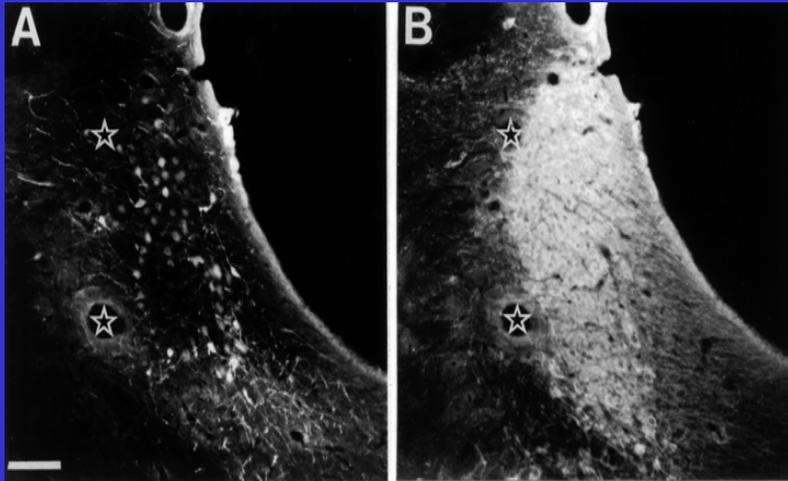


DO CHOLINERGIC OR OTHER BF NEURONS RECEIVE INPUT FROM ASCENDING MODULATORY SYSTEMS?

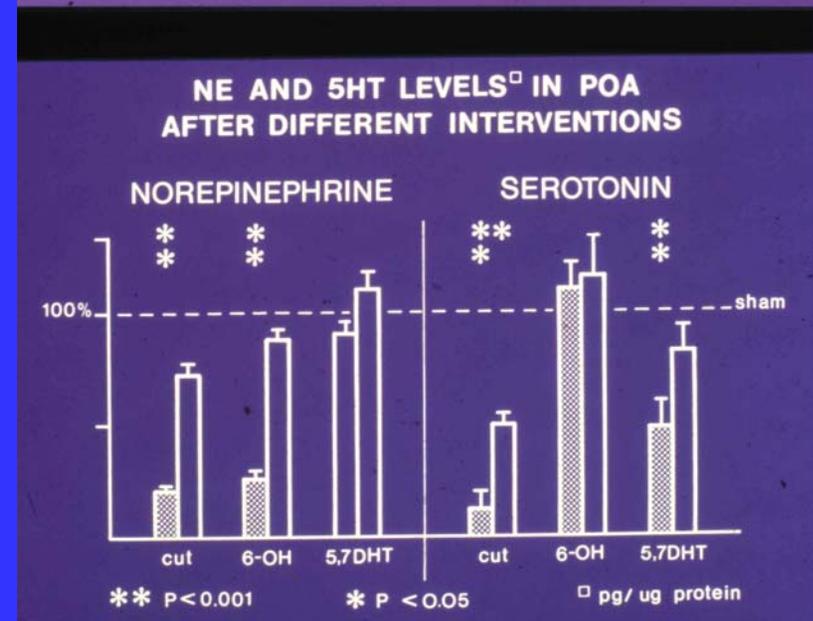
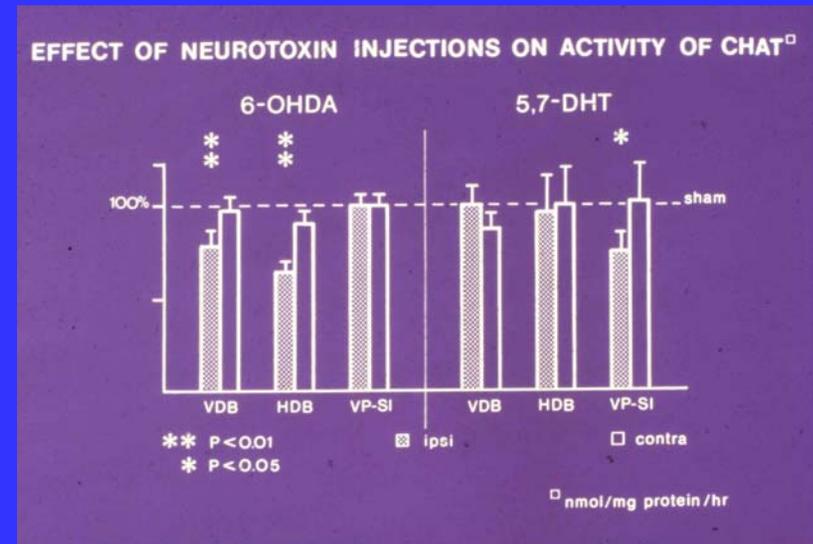
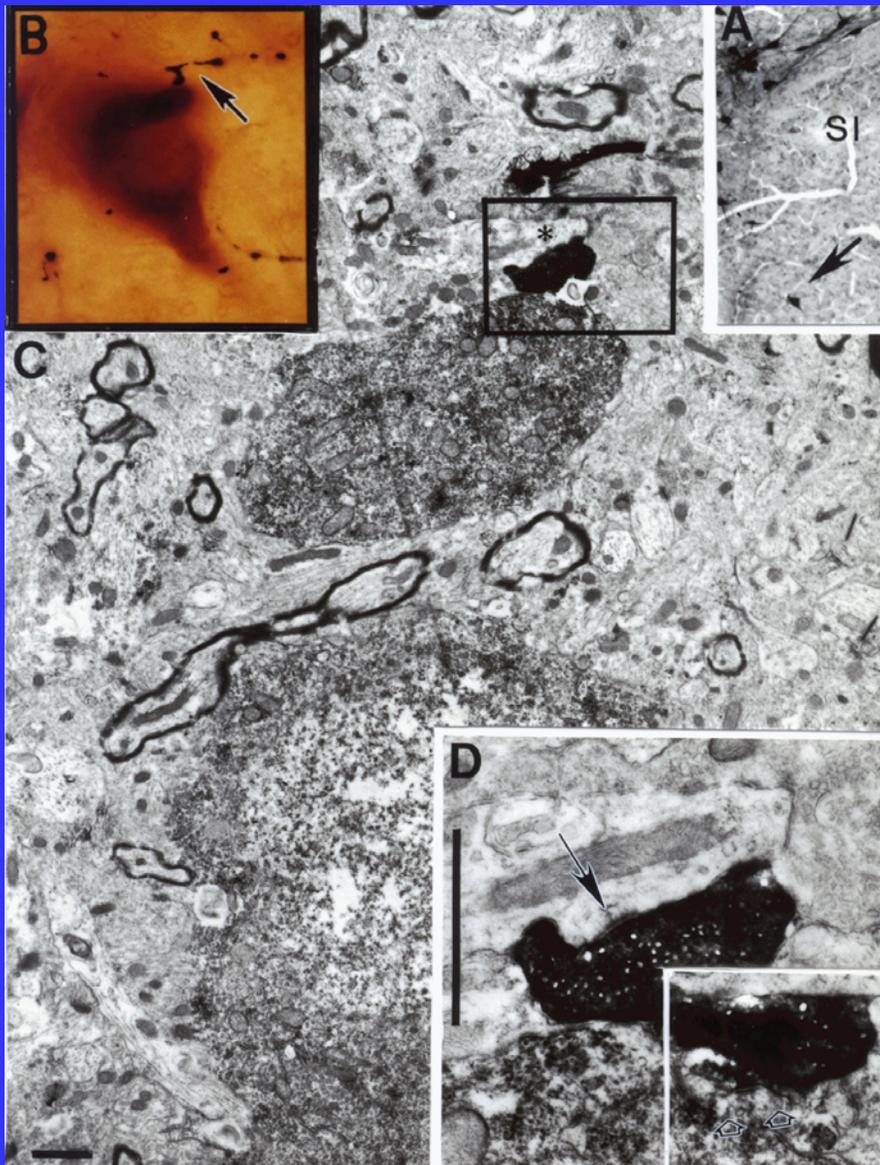


THE NEED FOR EM STUDIES

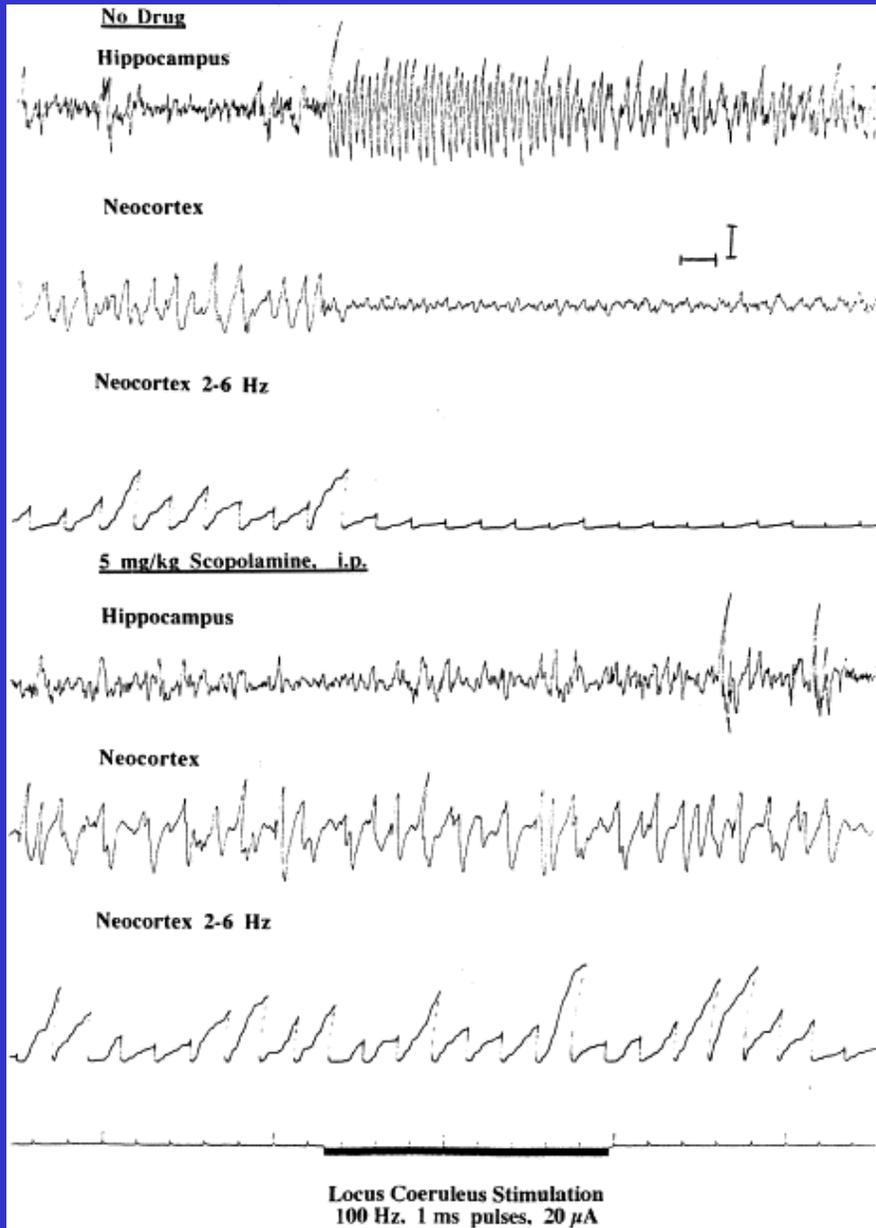
Diffuse input to BF: LC axons



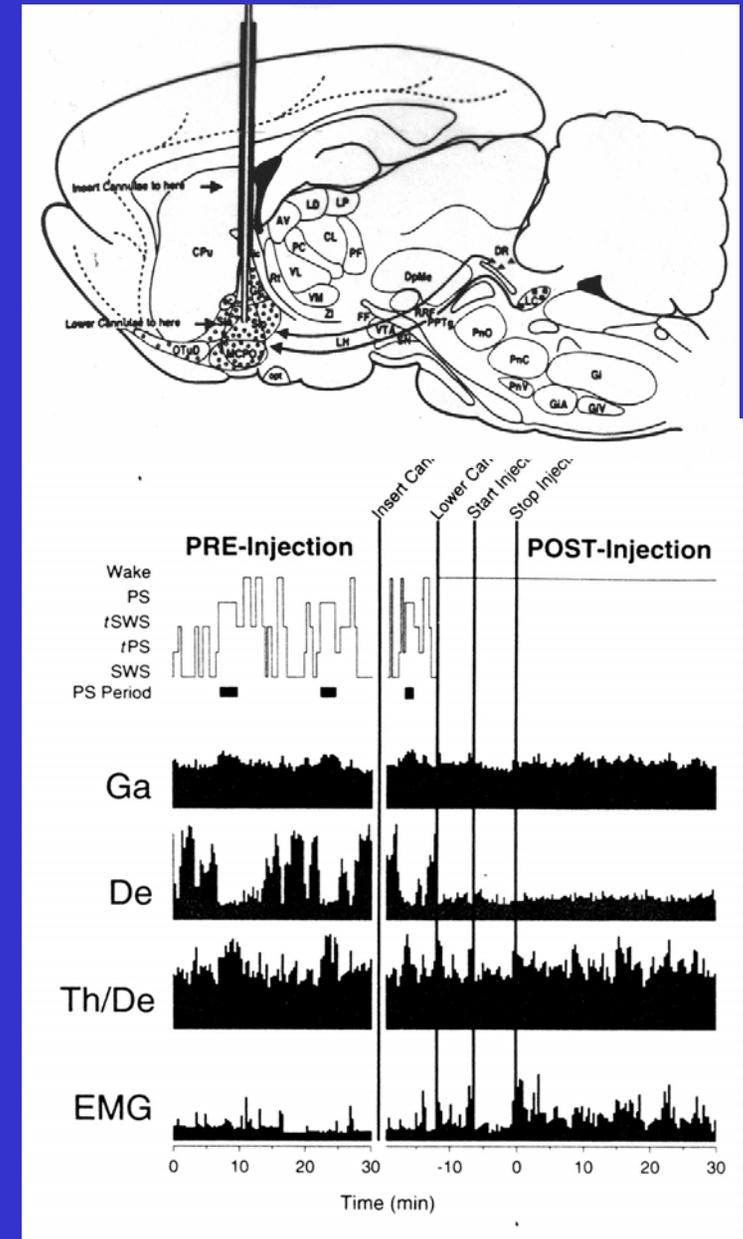
LC axons synapse on cholinergic and non-cholinergic neurons. Effect of 6-OHDA on forebrain ChAT level



LC can affect cortical EEG via the BF

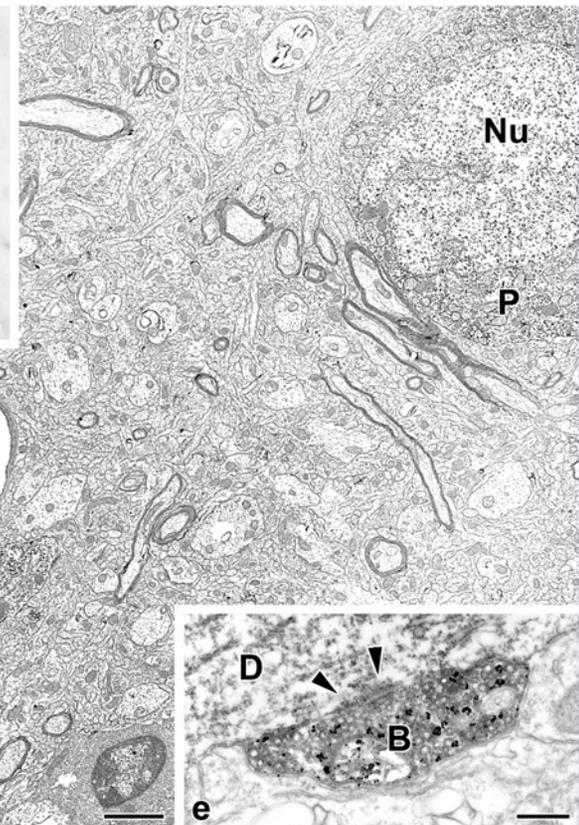
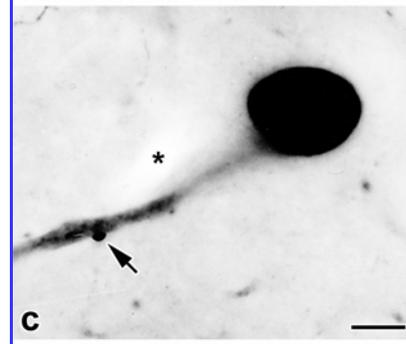
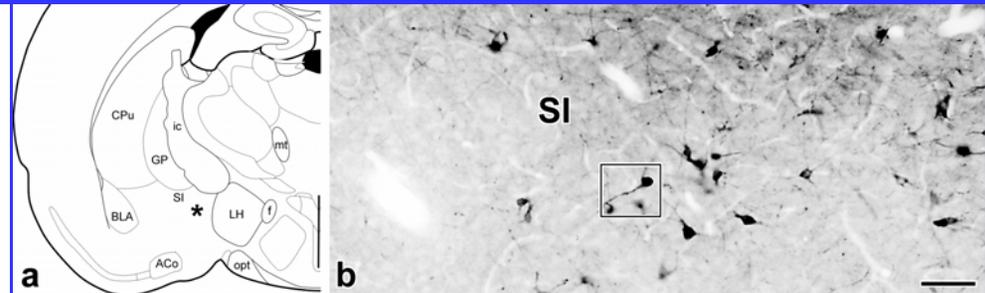
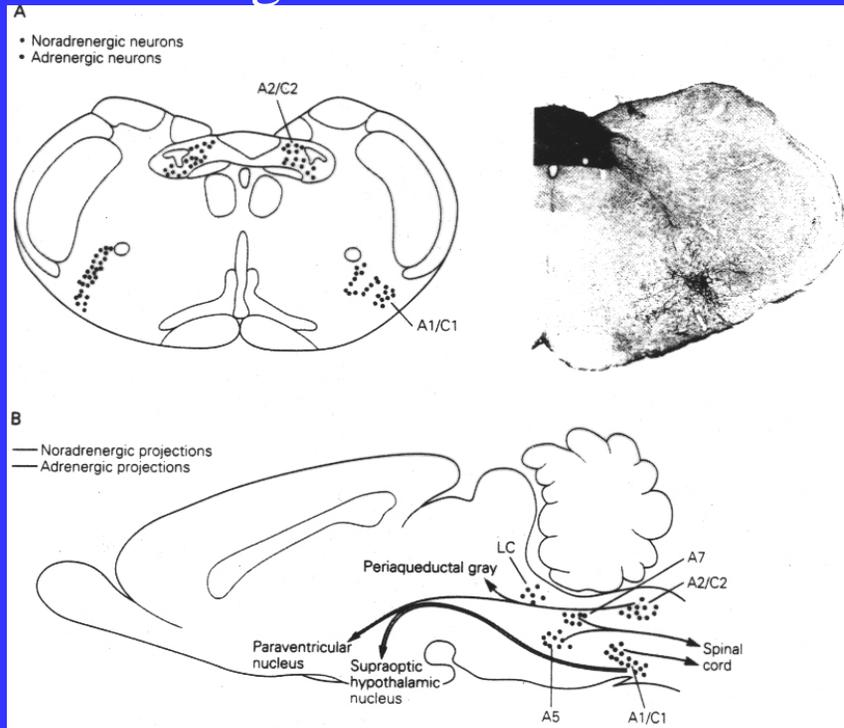


Dringenberg and Vanderwolf, 1998



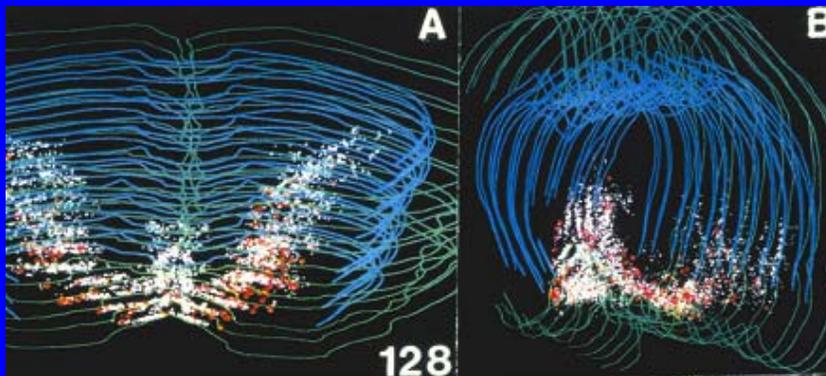
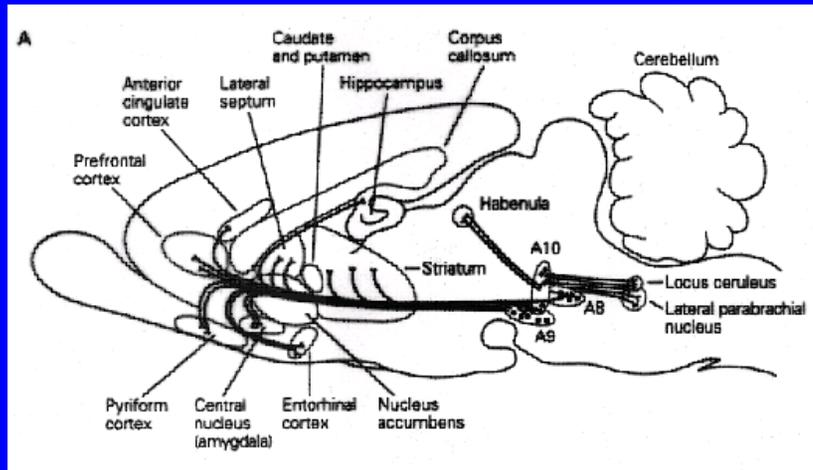
Cape and Jones, 1998

Diffuse input to cholinergic neurons: PNMT axons synapse with cholinergic neurons

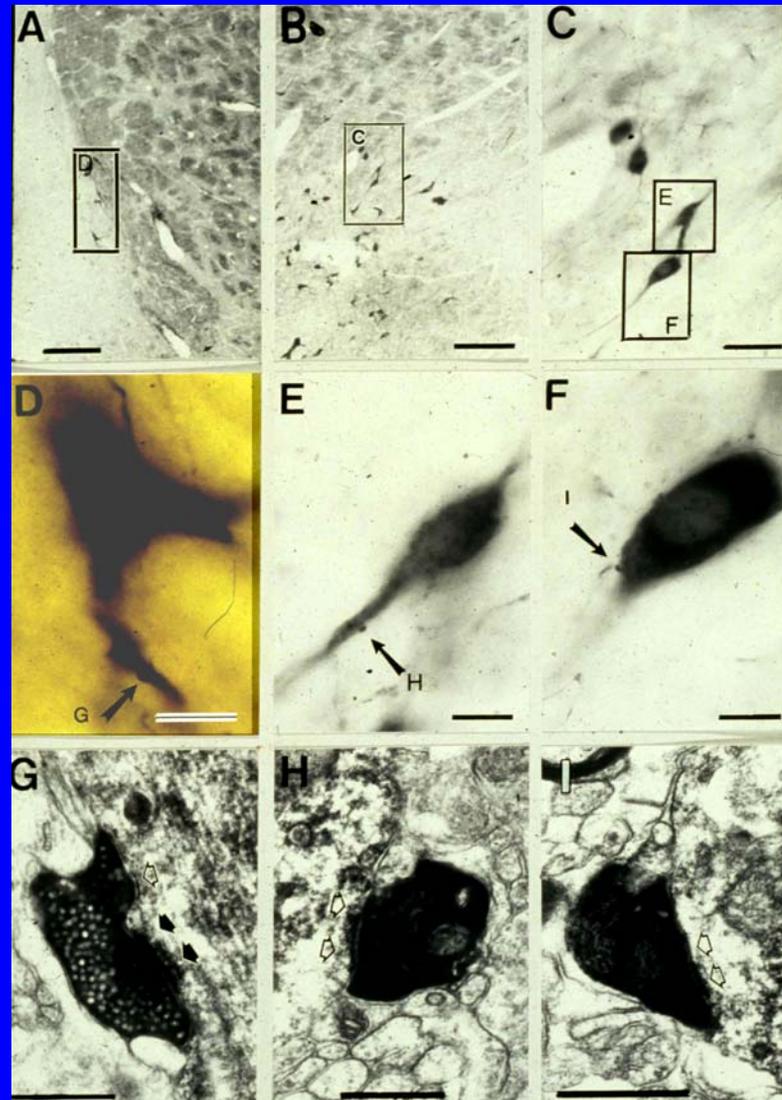


Hajszan and Zaborszky, 2002

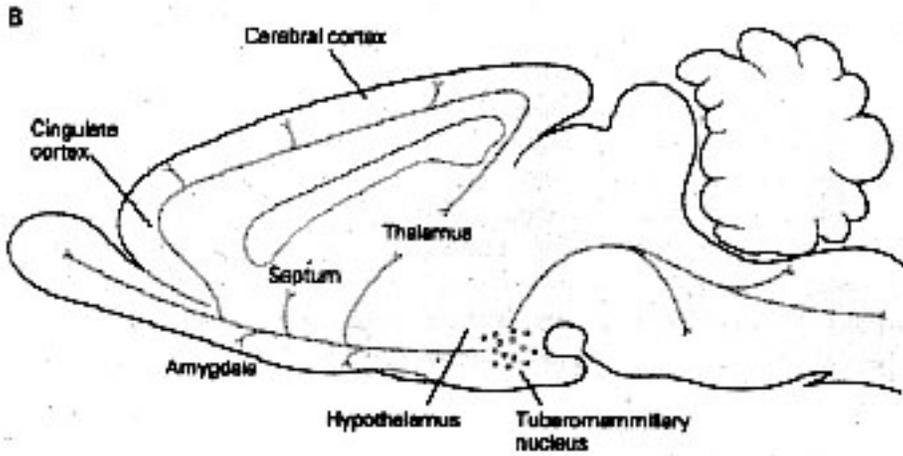
CHOLINERGIC NEURONS RECEIVE SYNAPSES FROM THE MIDBRAIN DOPAMINERGIC AREA



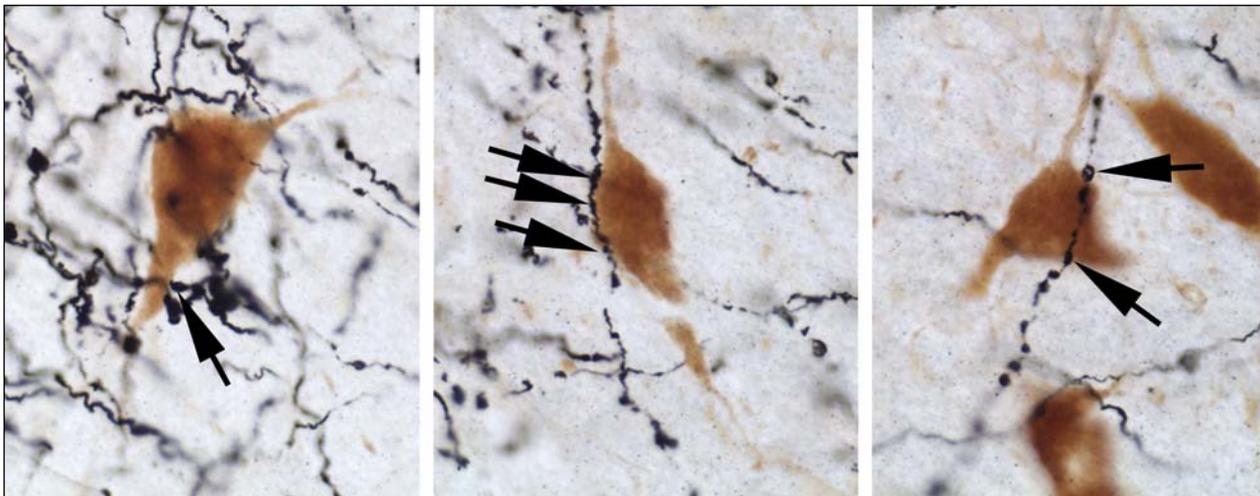
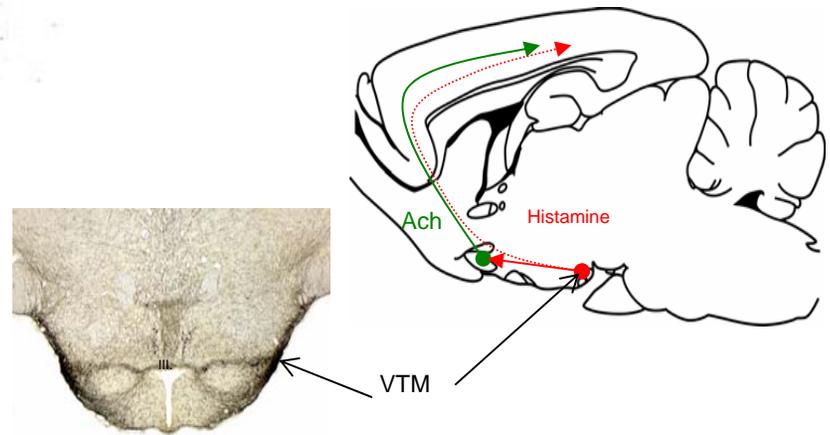
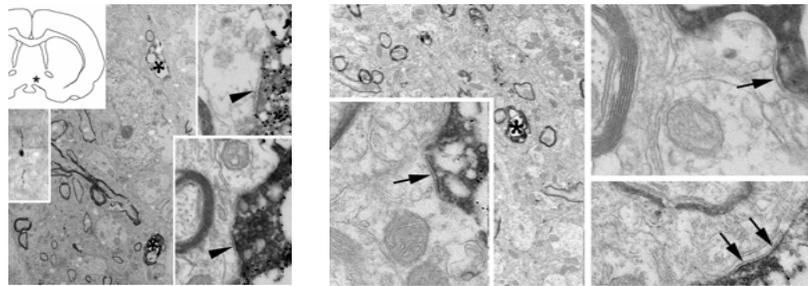
Ventral midbrain axons contact CH (as well as PV) cells in the BF.



Gaykema and Zaborszky, 1996

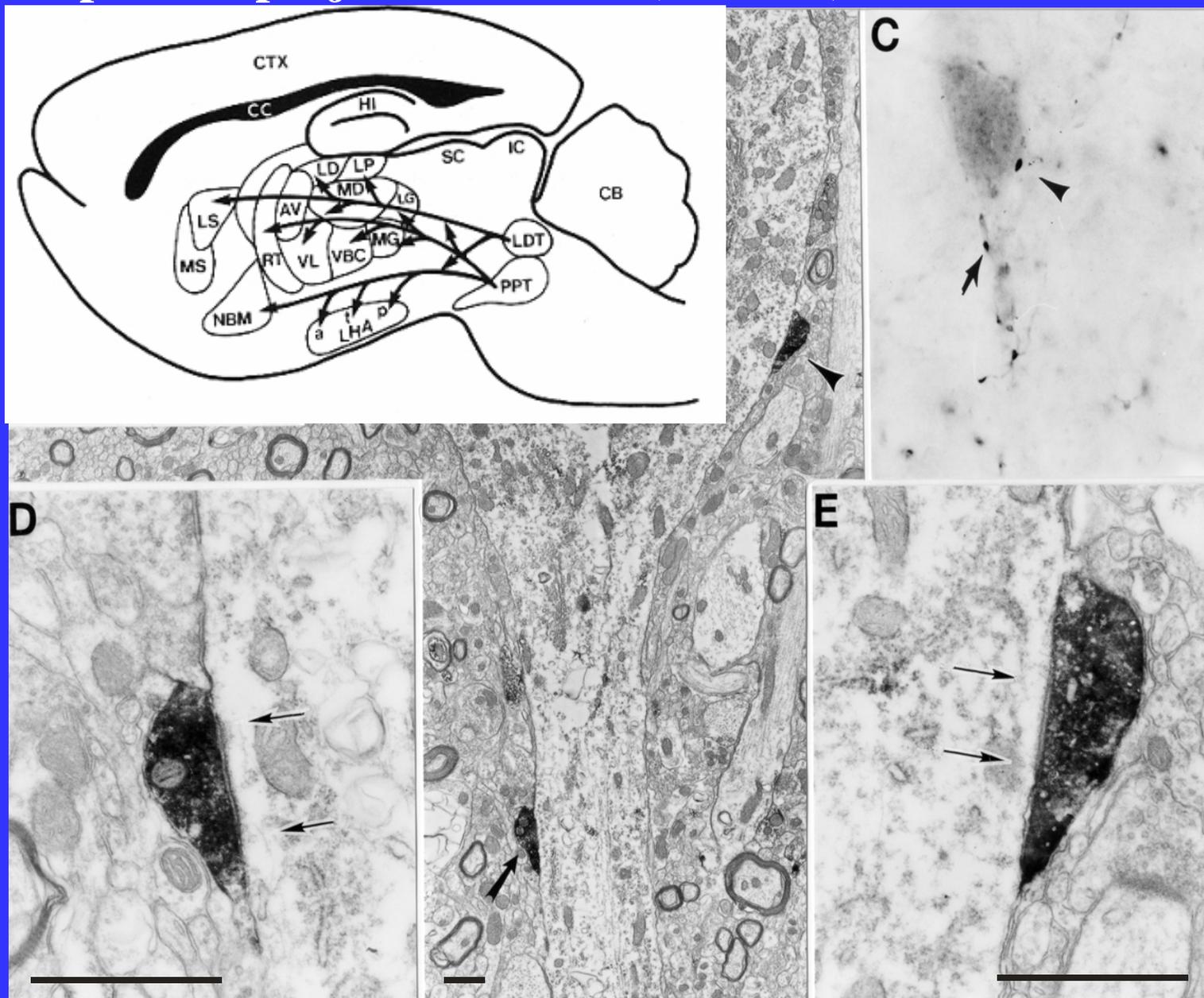


Diffuse input to the BF:
histaminergic axons in
close proximity to
ChAT neurons

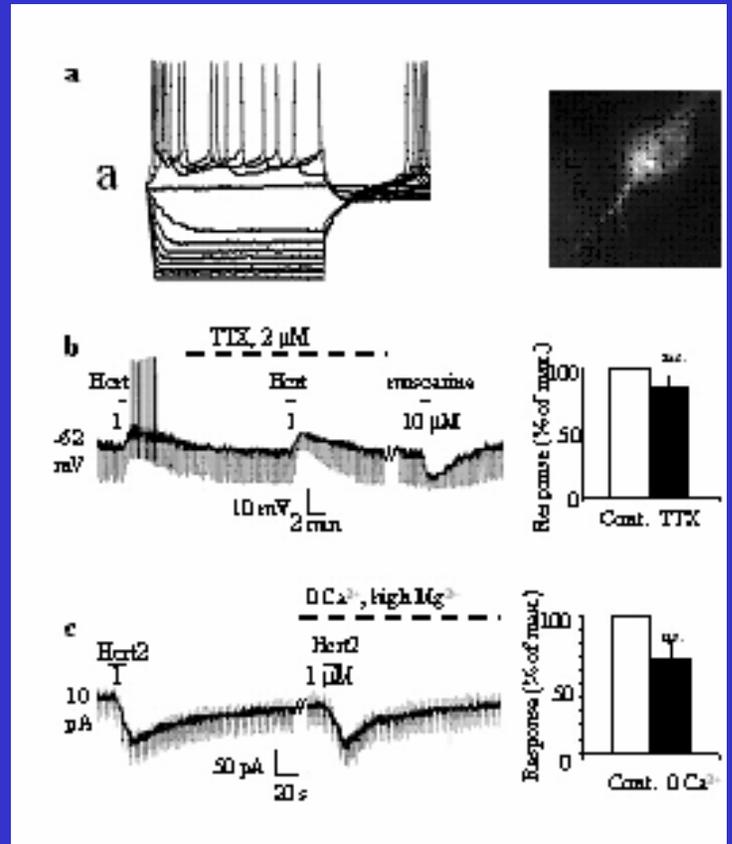
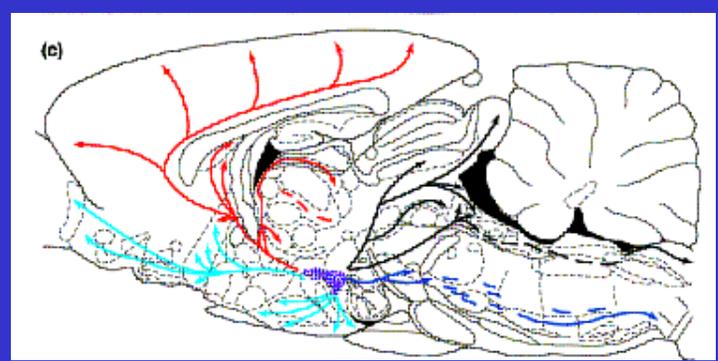
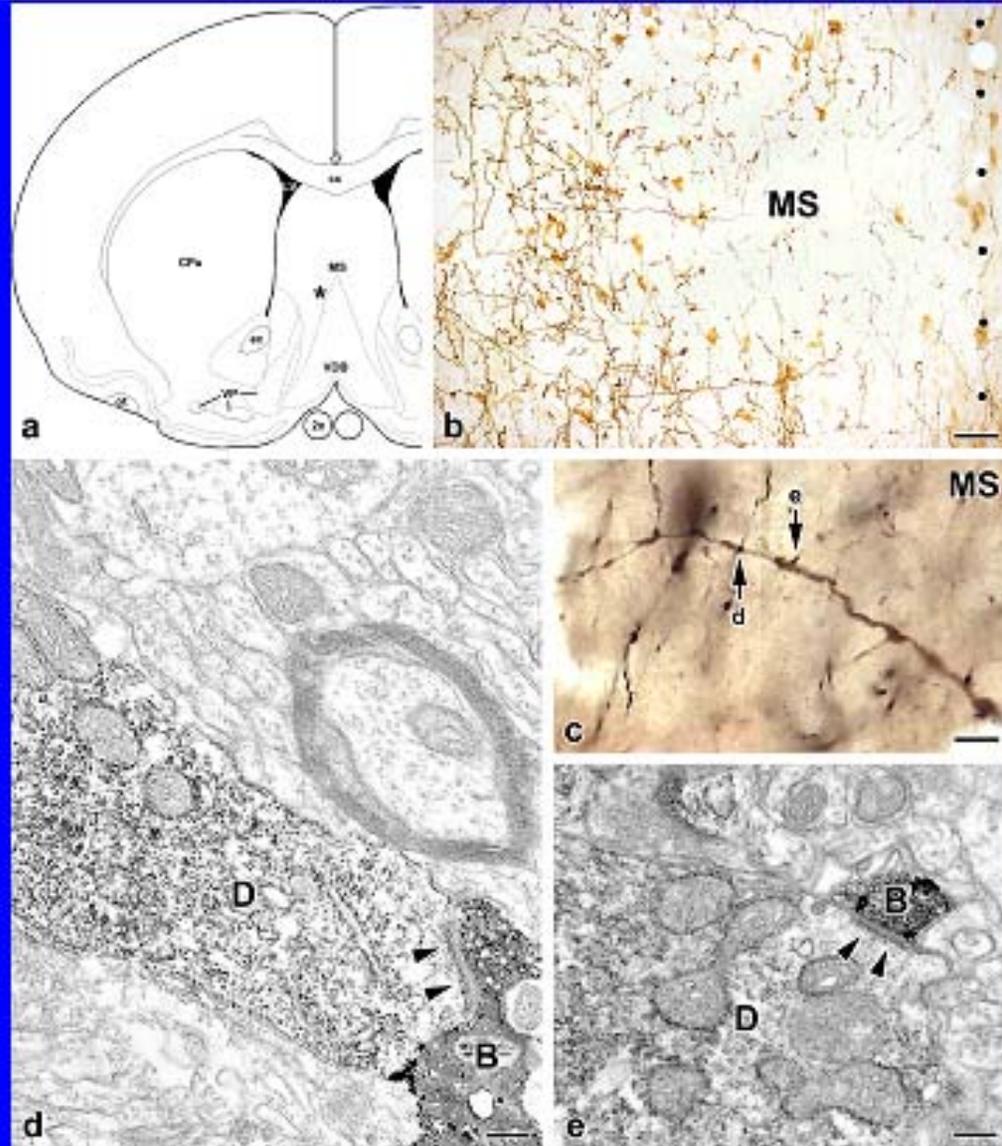


Turi et al.: SFN
2004

Mesopontine projections to PV (and CH) neurons in the BF

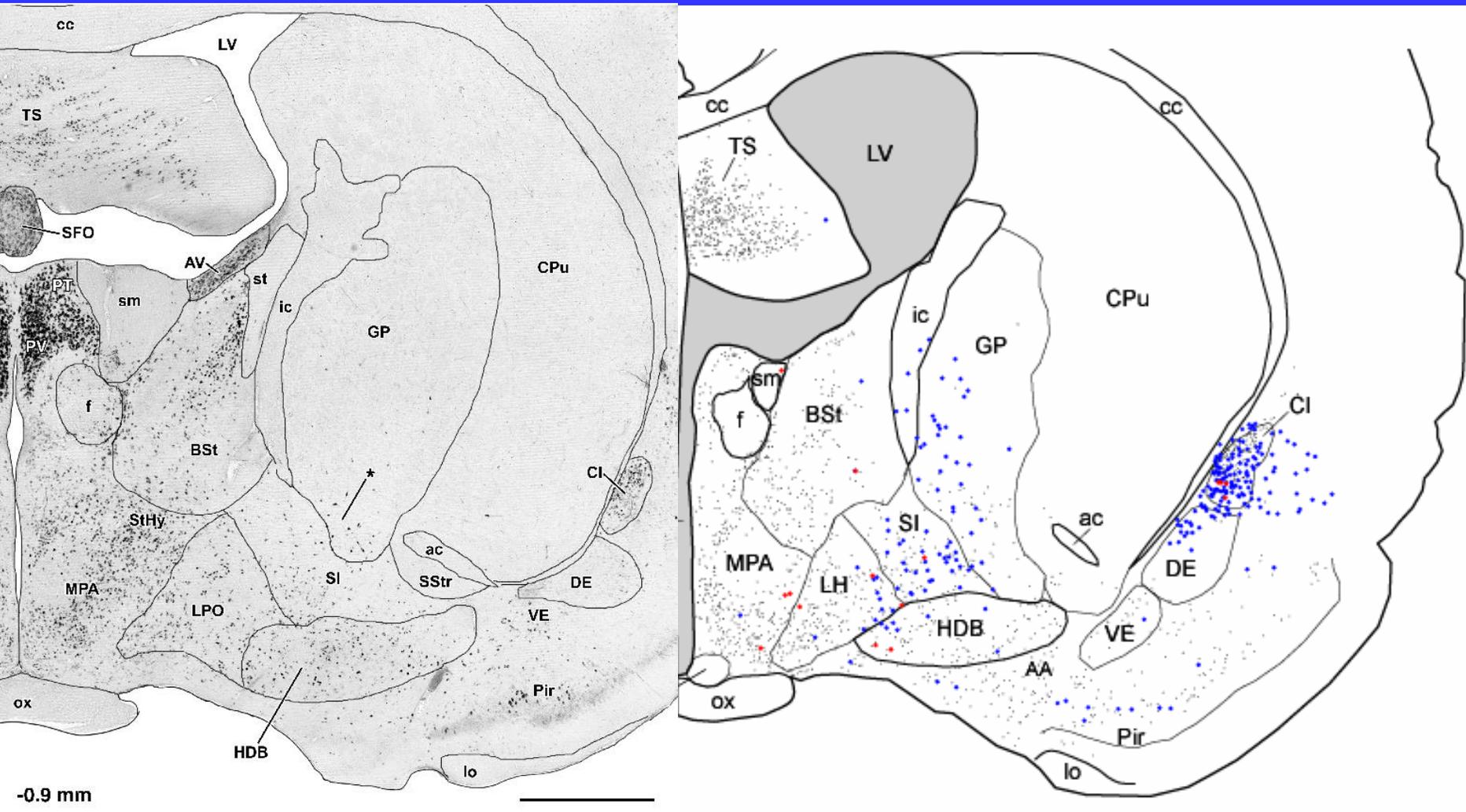


Diffuse input to BF: hypocretin axons



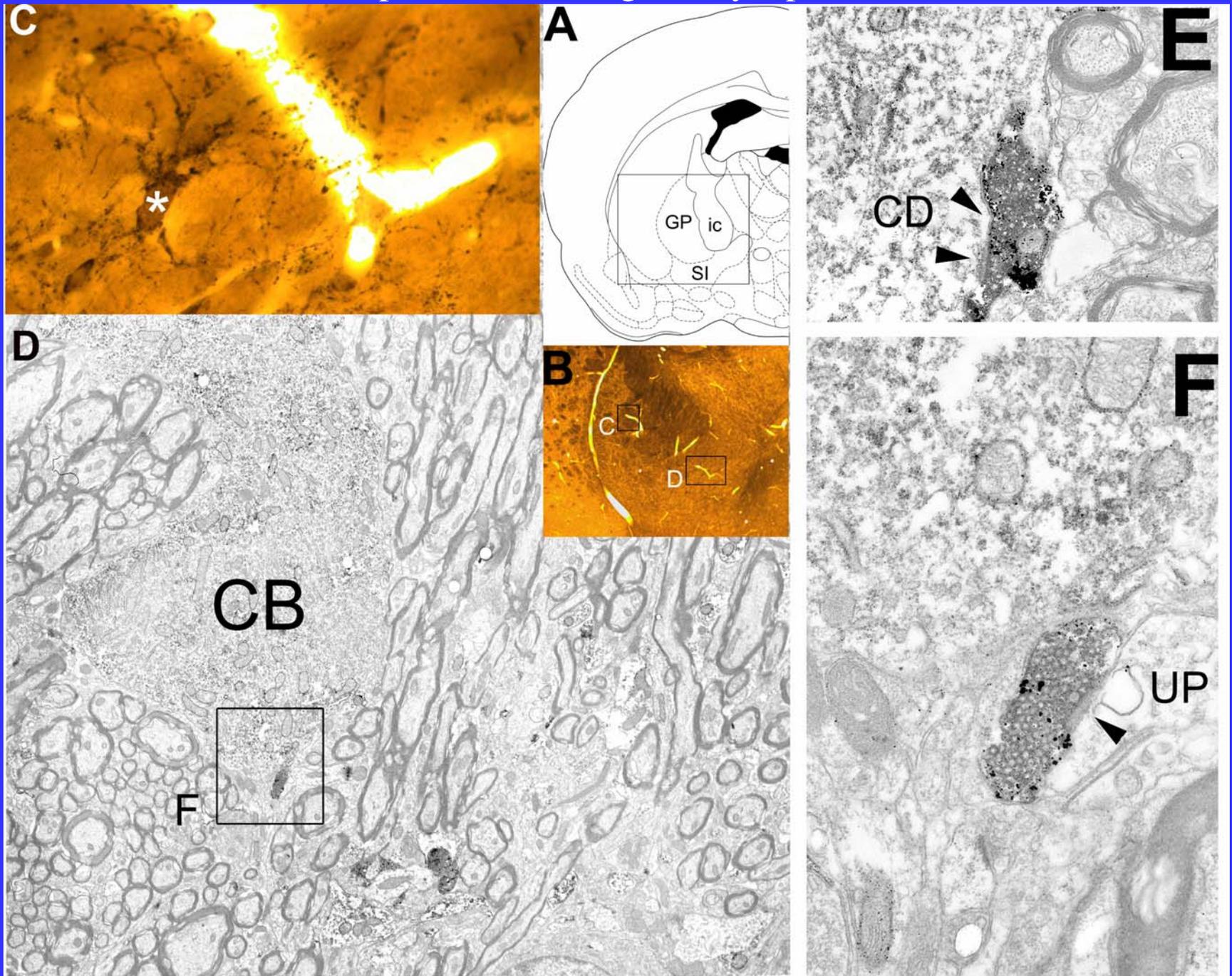
a: whole-cell current clamp; **b:** current clamp showing the depolarizing effect of Hcrt. **c:** voltage clamp (-65 mV) in which Hcrt2 induced an inward current that persisted in zero Ca²⁺, high Mg²⁺ ACSF.

Glutamatergic (Vglut2) projections to the prefrontal cortex

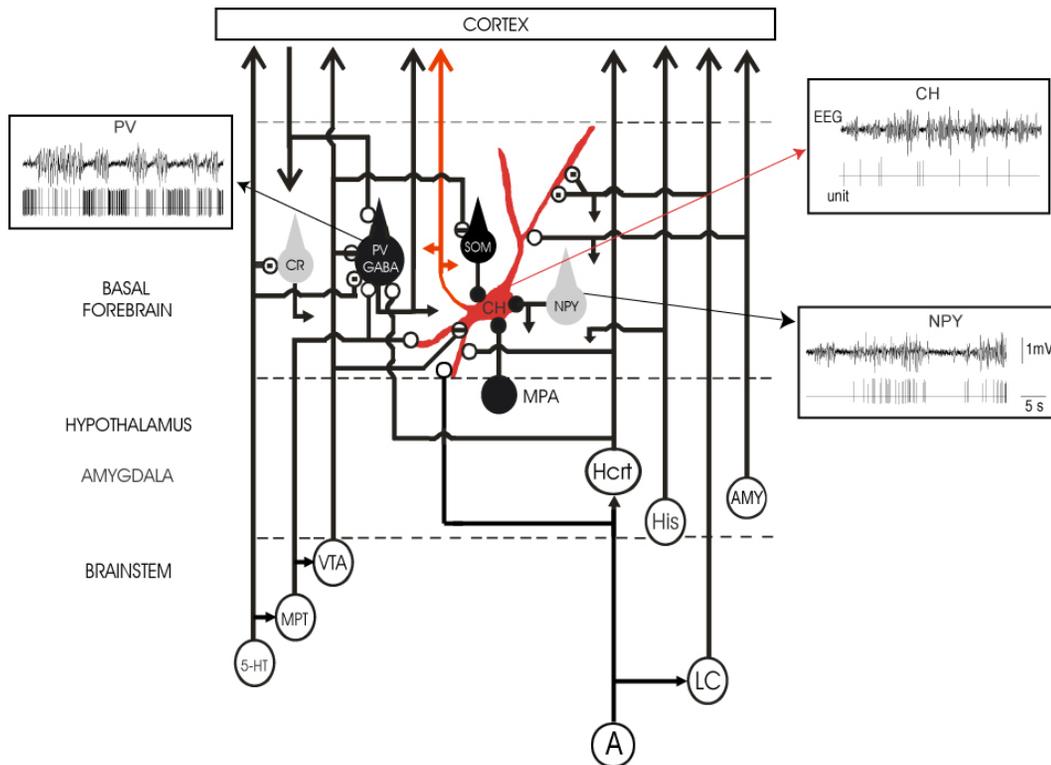
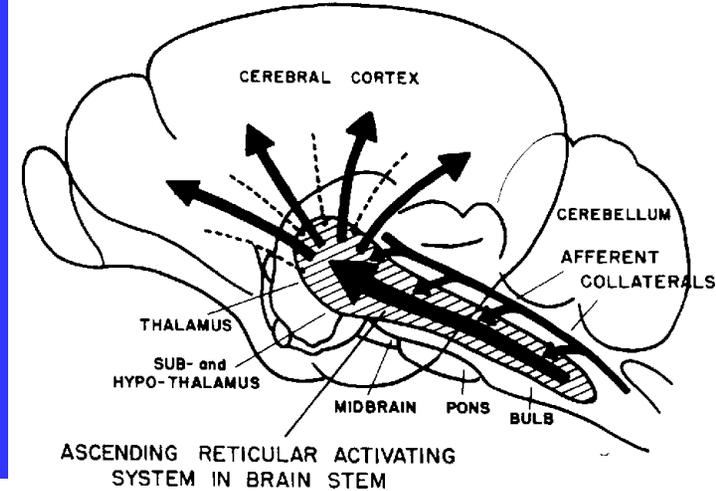


Hur and Zaborszky: J. Comp. Neurol, 2005

Diffuse input to BF (?): Vglut 2 synapses on CH dendrites



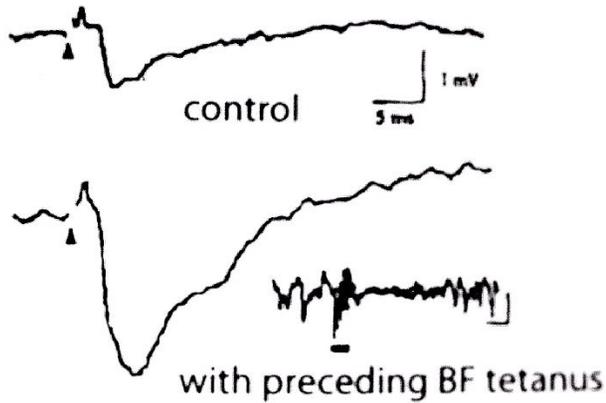
Moruzzi and Magoun, 1951



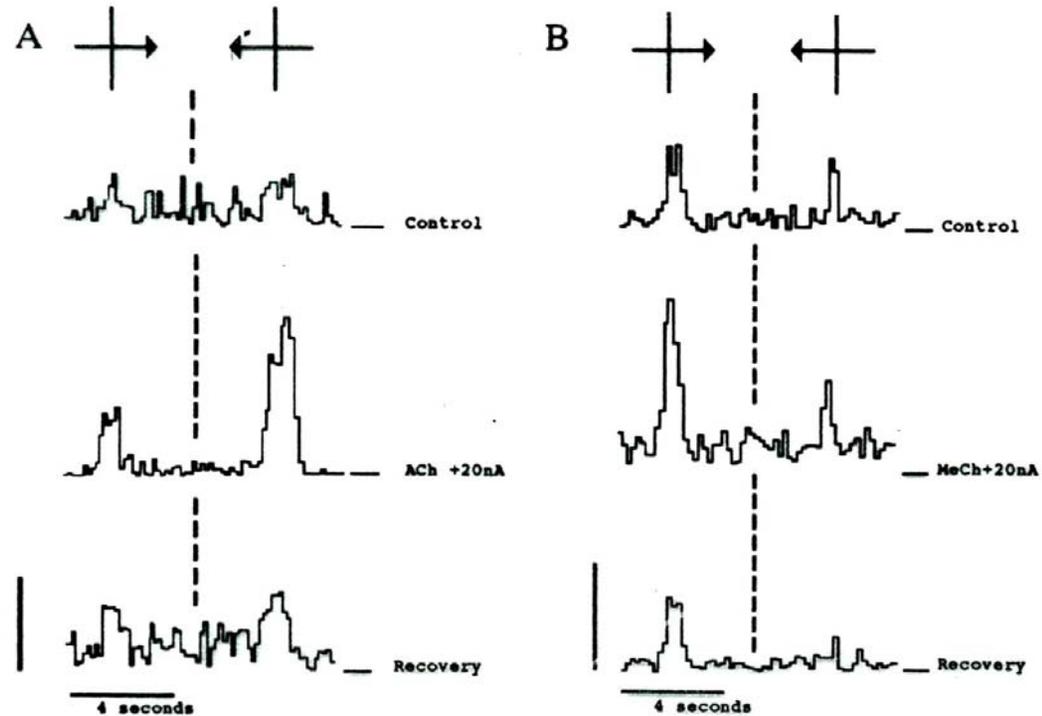
The brainstem can activate neocortical neurons via various neurons in the basal forebrain

Zaborszky and Duque, 2003

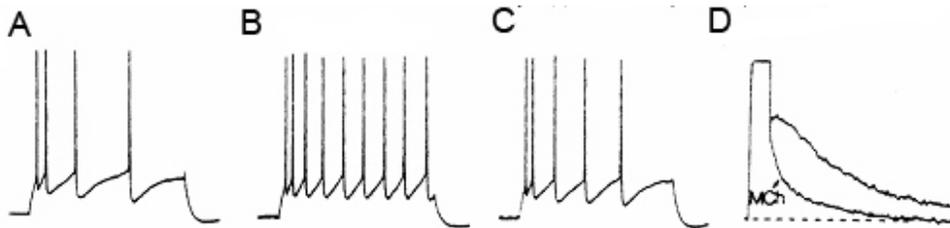
BF stimulation or cortical ACh application enhances sensory evoked responses: Is this part of an arousal reaction or a more selective process (attention)?



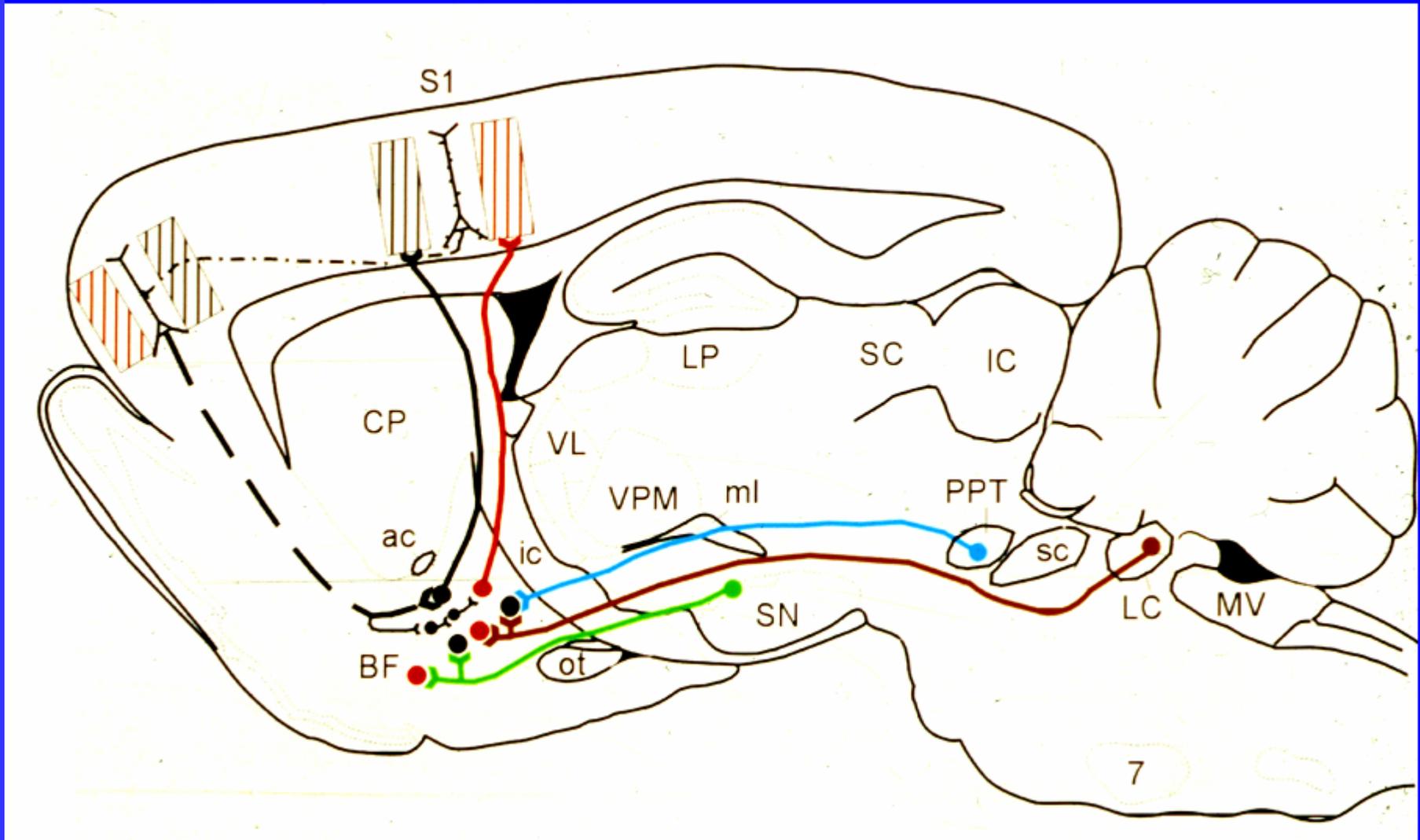
Auditory cortical response to medial geniculate stimulus is modified by preceding (30 msec) basal forebrain stimulation. (Metherate and Ashe, 1991)



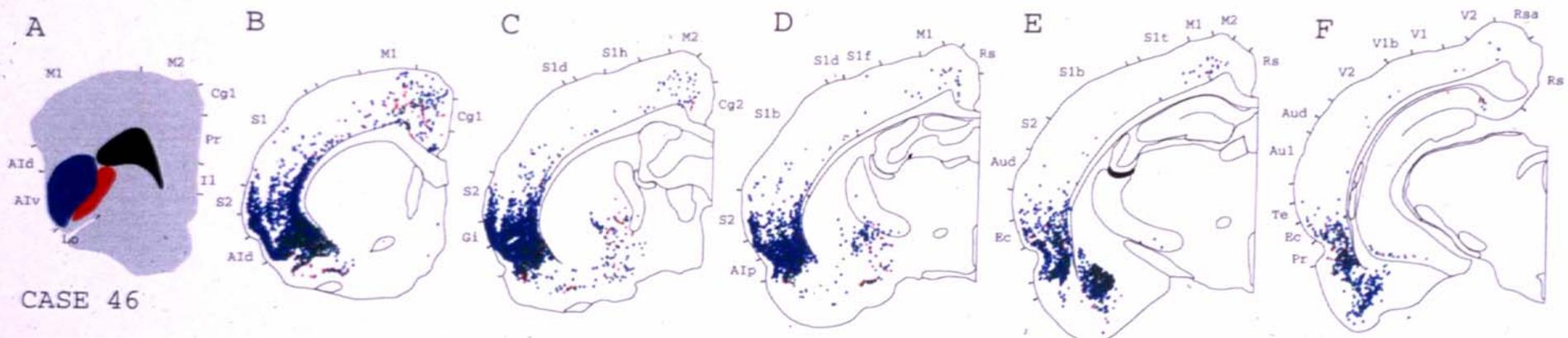
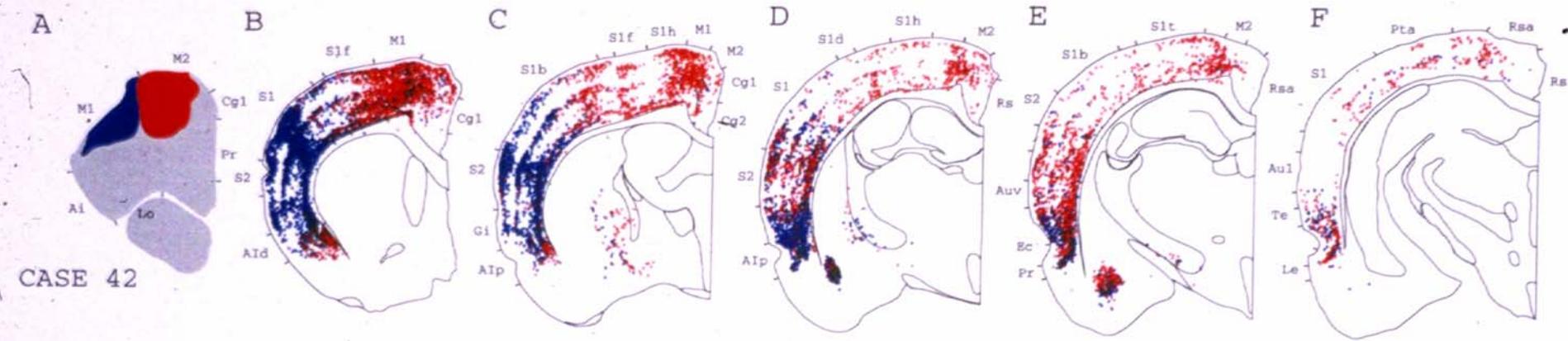
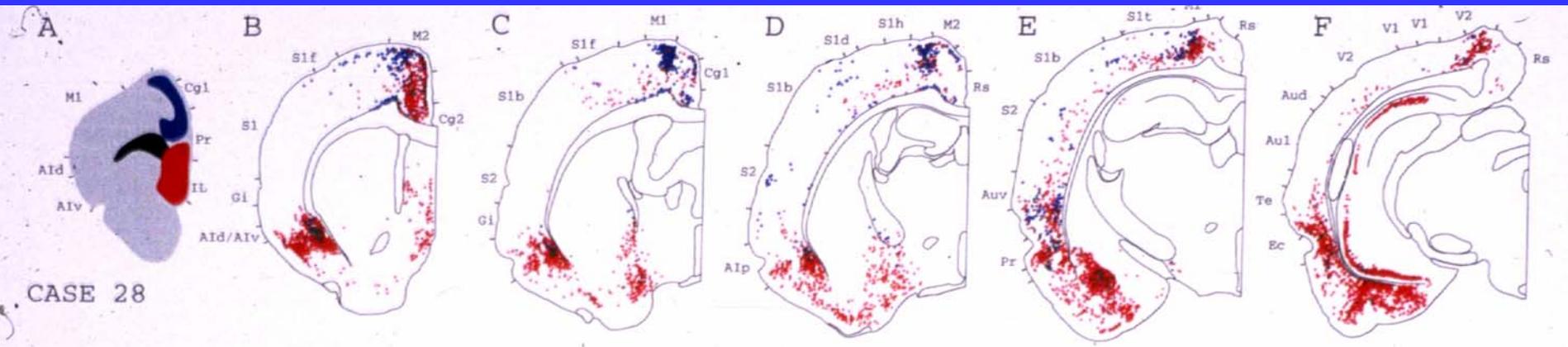
Effect of cholinergic facilitation on the direction selectivity of a LIV simple cell (A) and a LVI complex cell (B). PSTH showing the response of the cells to an optimally oriented bar of light moving forwards and backwards over the receptive field. (Silito, 1993)



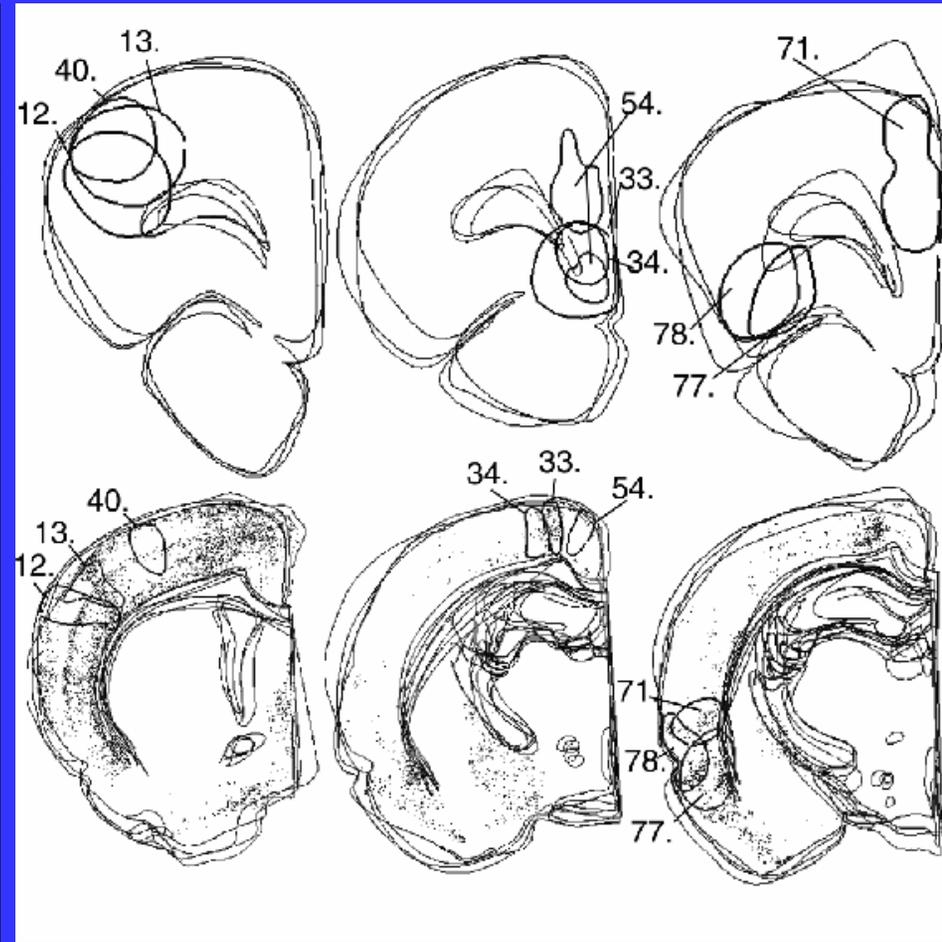
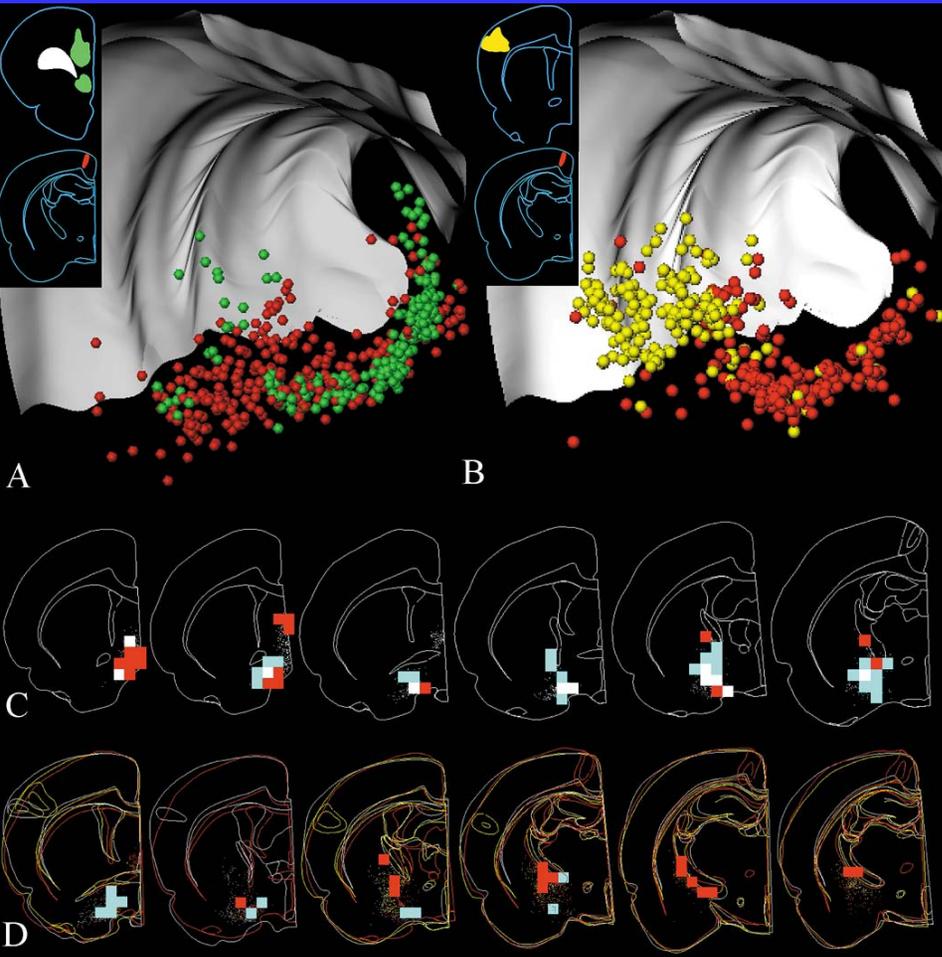
HOW THE BASAL FOREBRRAIN ORGANIZATION SUPPORTS A ROLE IN SPECIFIC SENSORY PROCESSING (ATTENTION)?



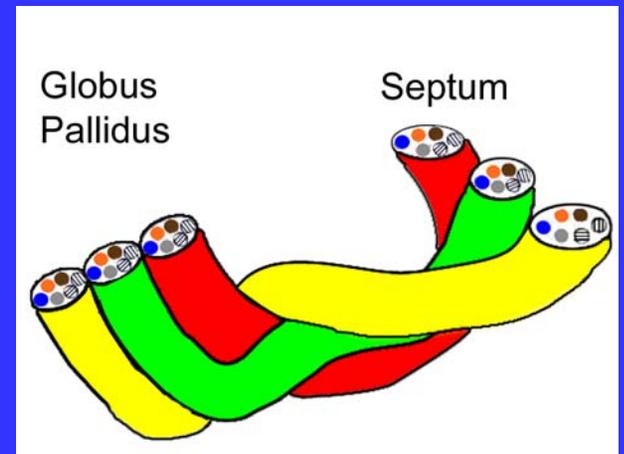
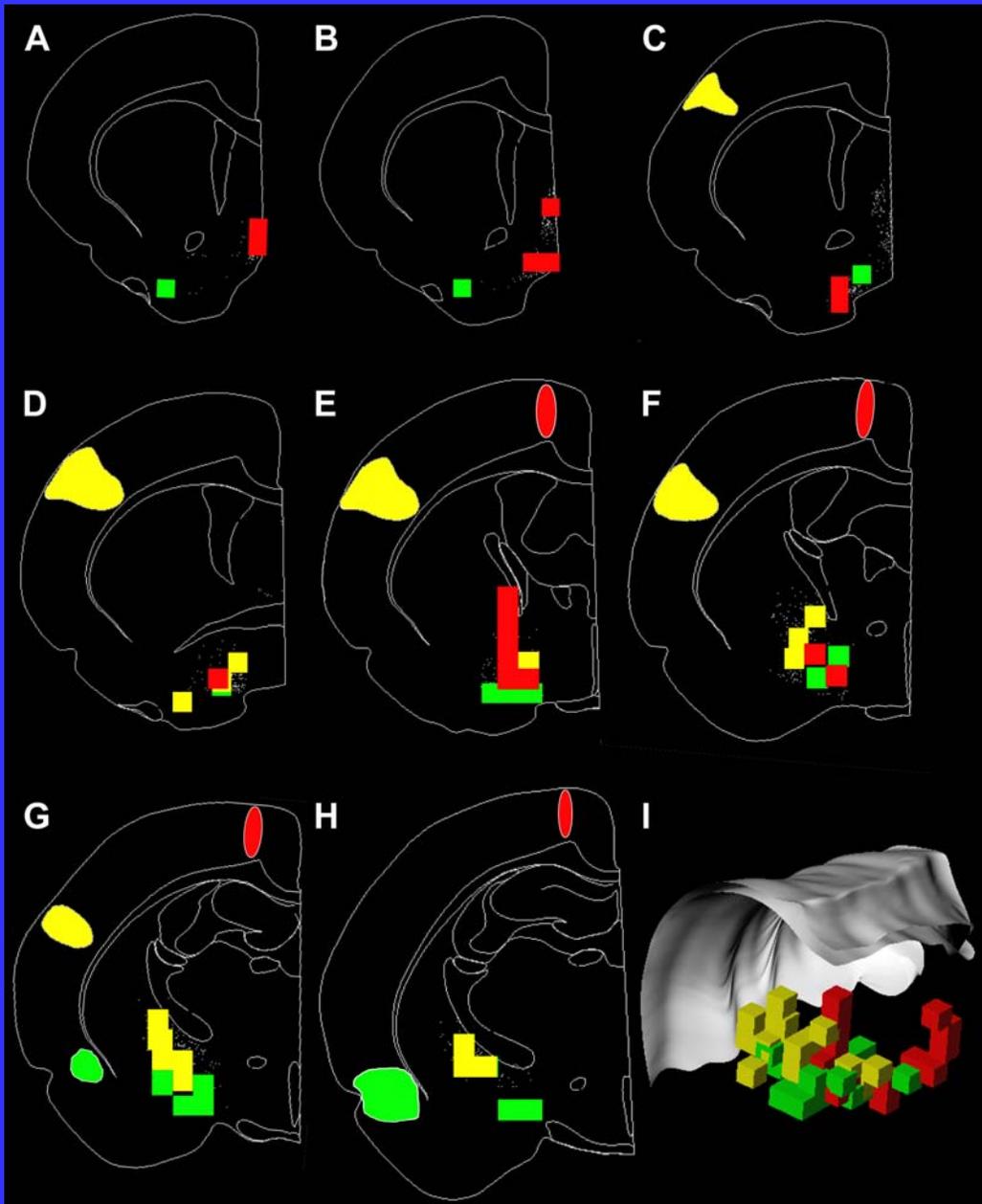
Hierarchical cortical processing



Cortical regions that are interconnected receive input from overlapping pool of neurons in the BF. In contrast, cortical regions that are not interconnected receive their input from segregated pool of BF neurons

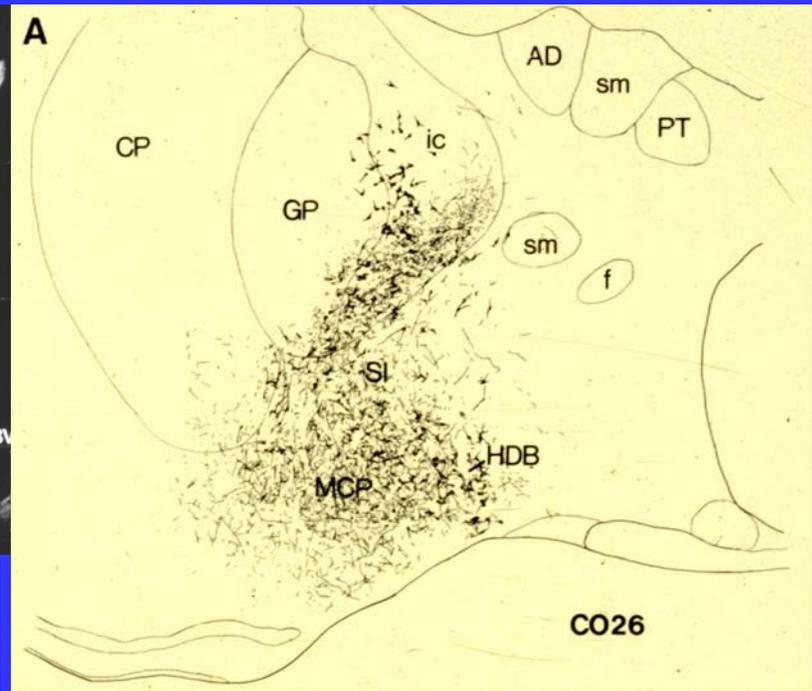
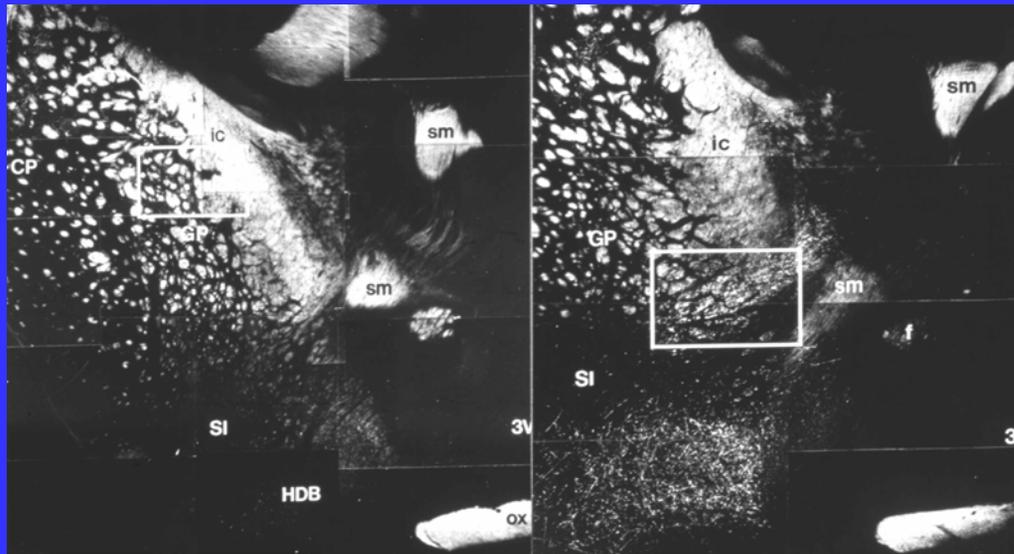


PROJECTIONS FROM BF
NEURONS TO DISTINCT
CORTICAL 'MODULES'
ARE SEGREGATED IN THE
BASAL FOREBRAIN

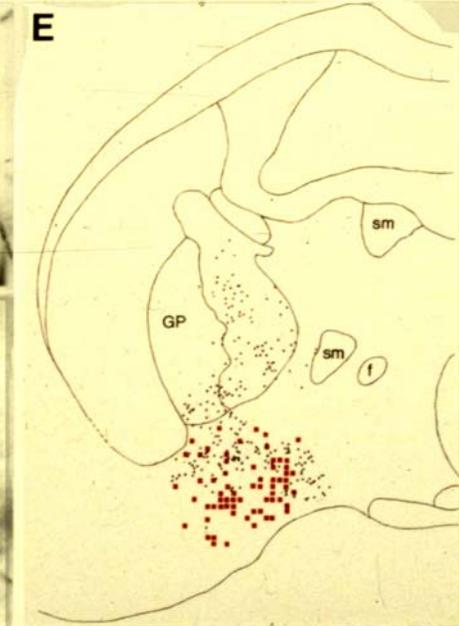
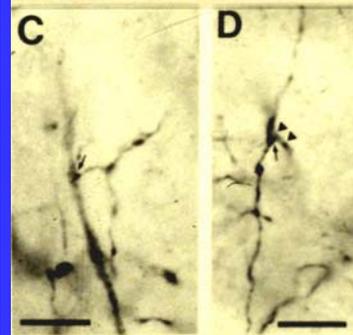
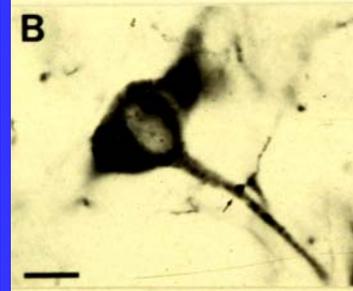
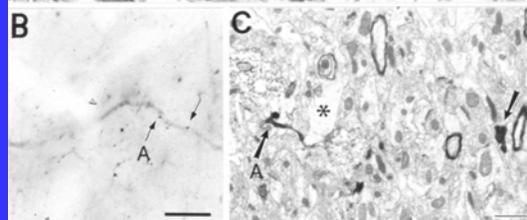
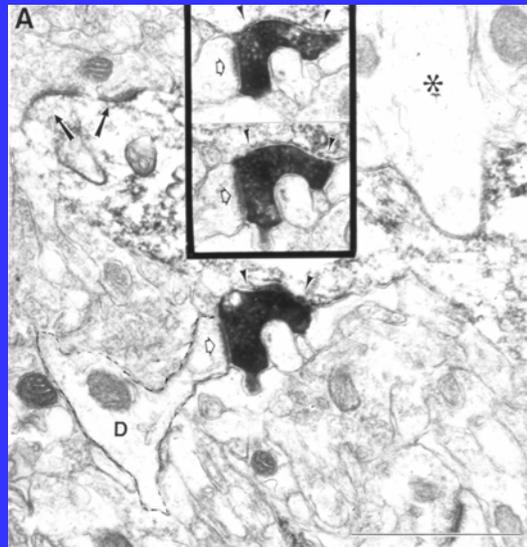


Zaborszky, 2002

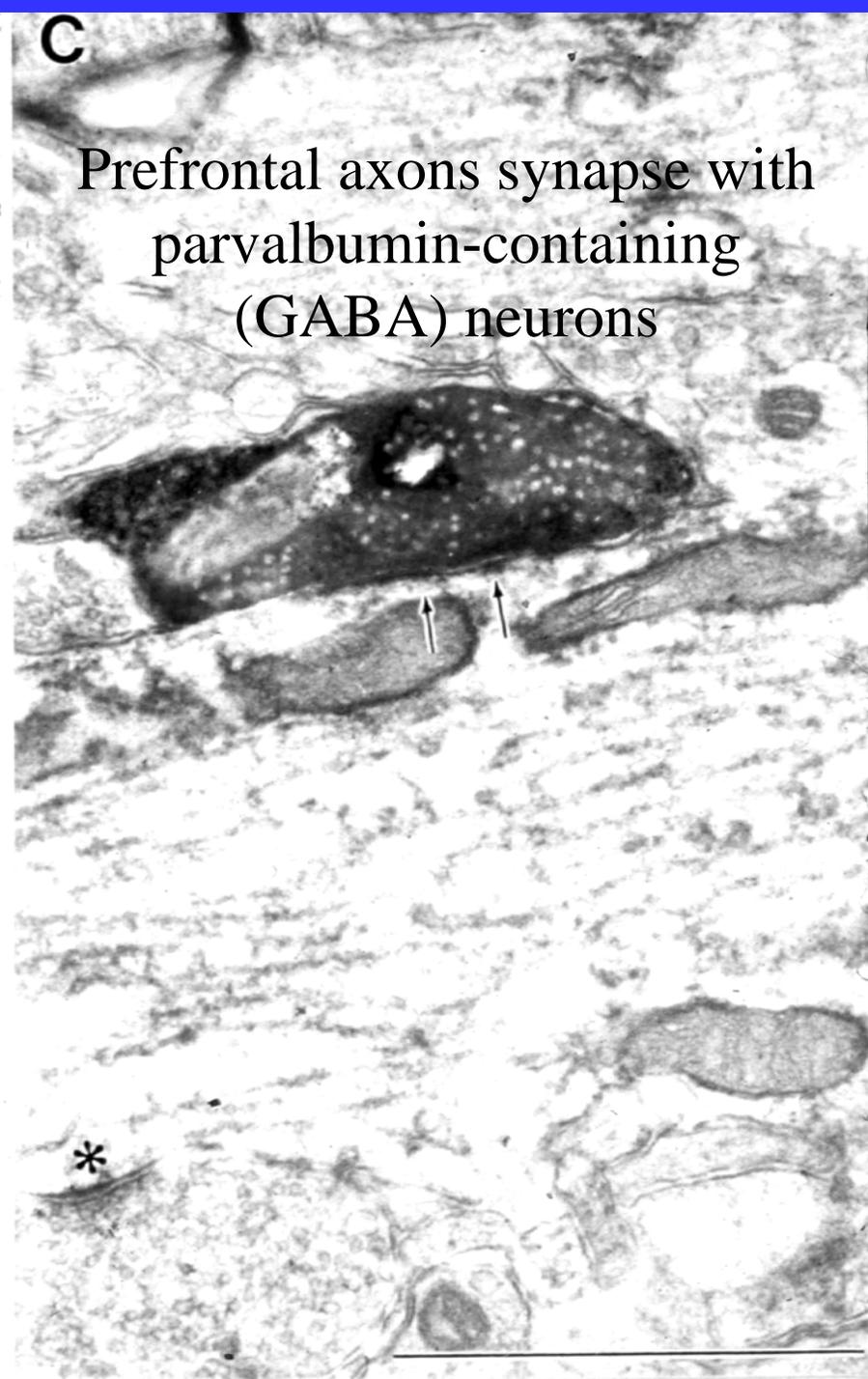
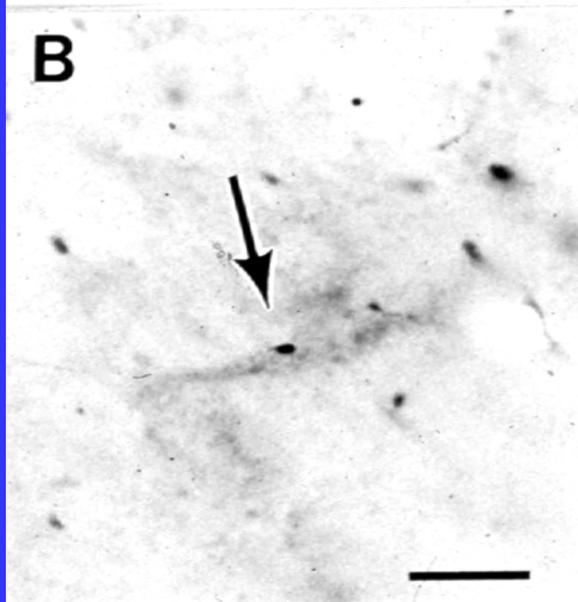
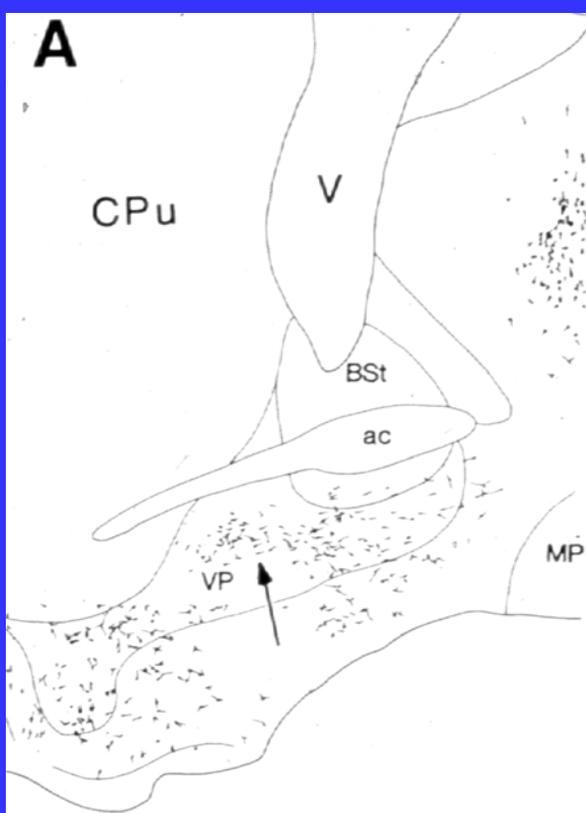
CORTICAL FEEDBACK TO BF ORIGINATES IN THE PREFRONTAL CORTEX



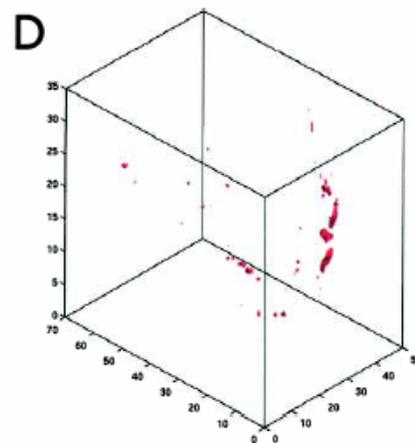
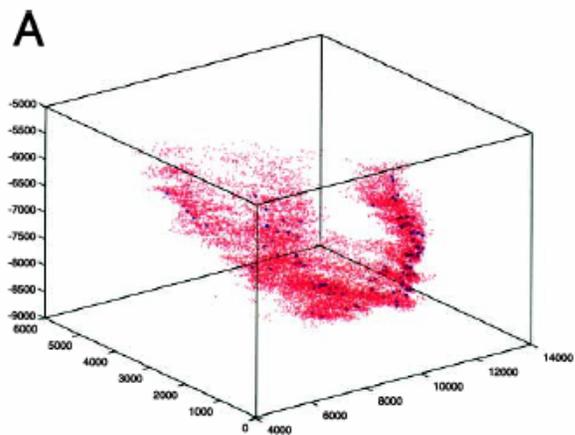
Synapses are on spine heads of non-cholinergic neurons



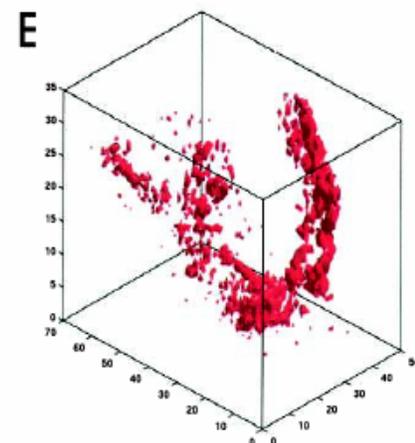
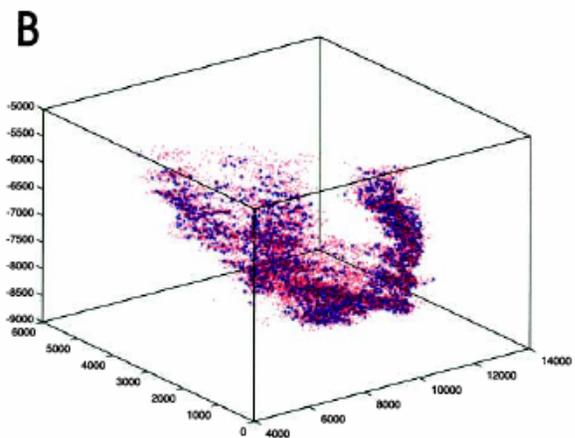
Zaborszky et al., 1997



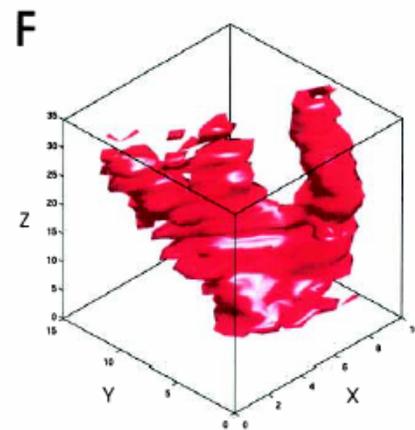
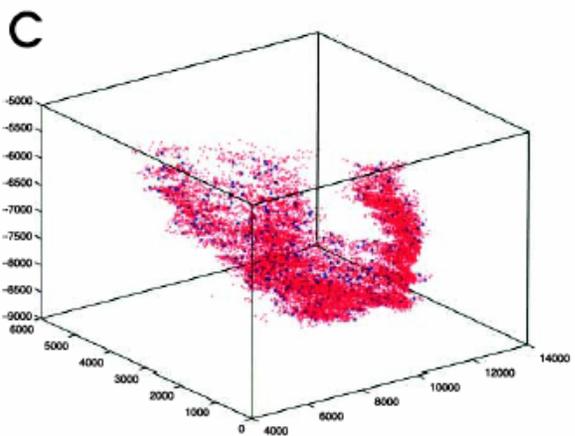
$d > 8/100 \times 100$

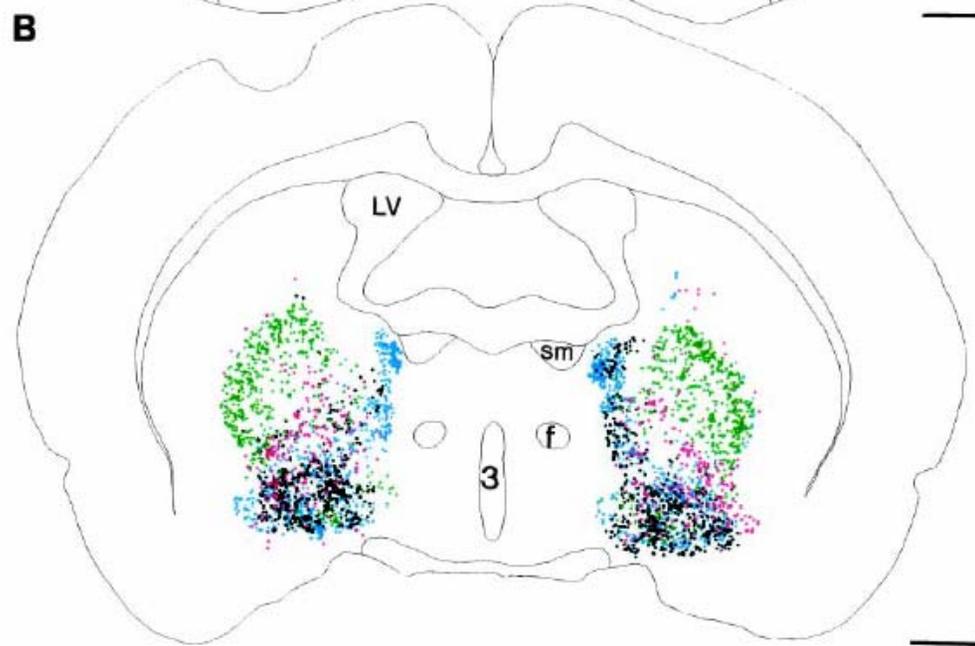
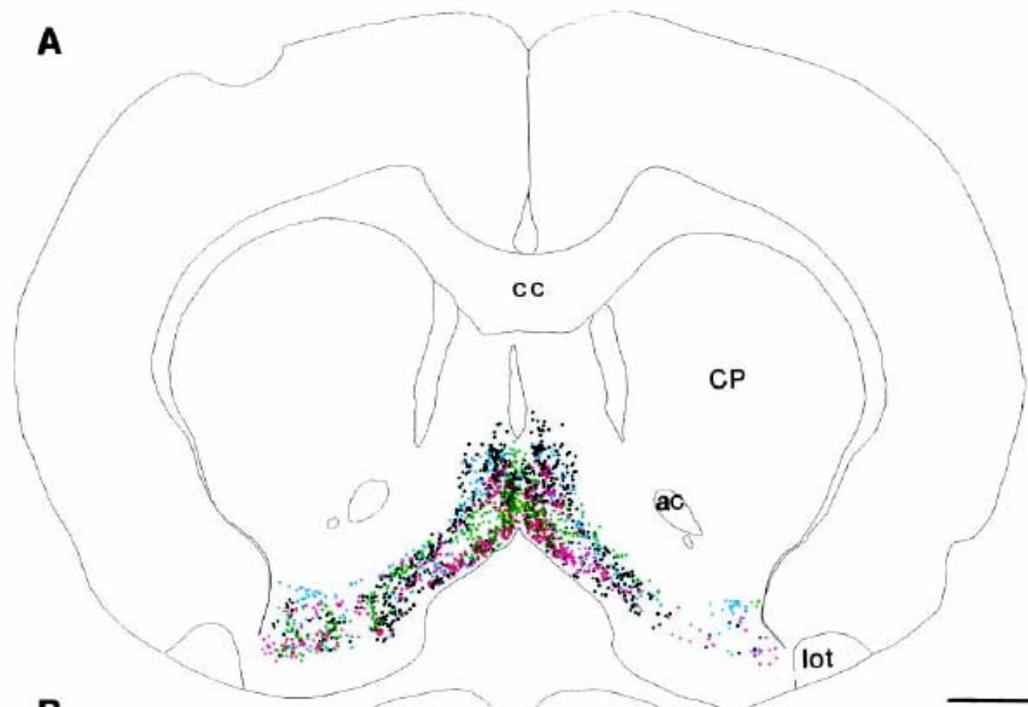


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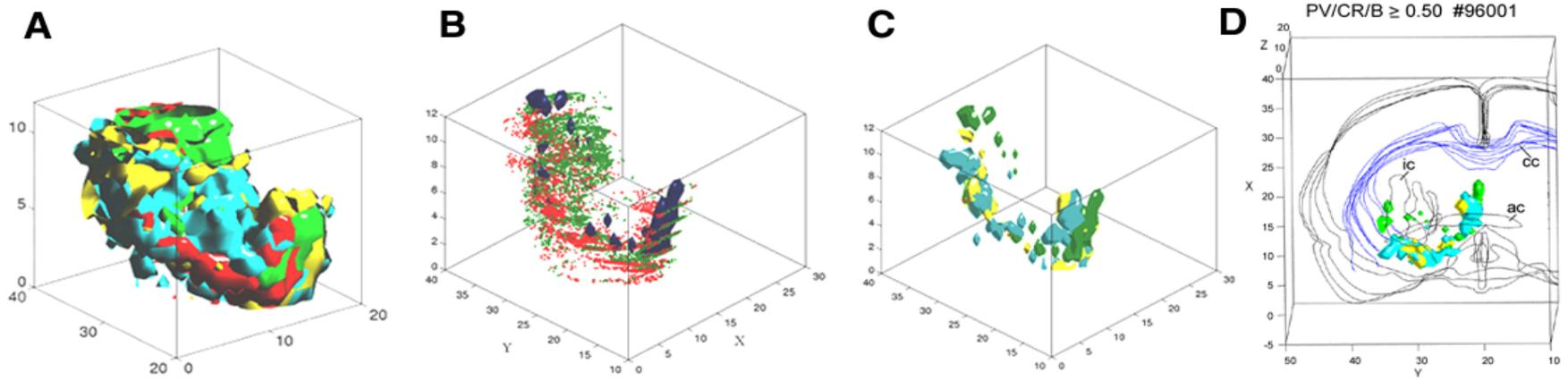
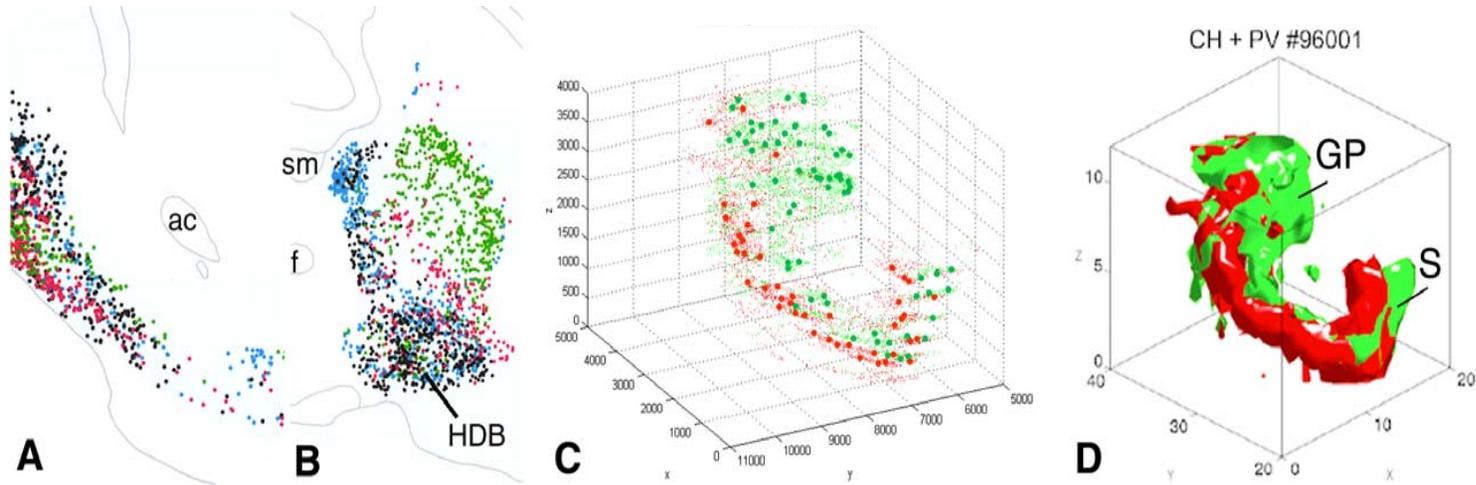


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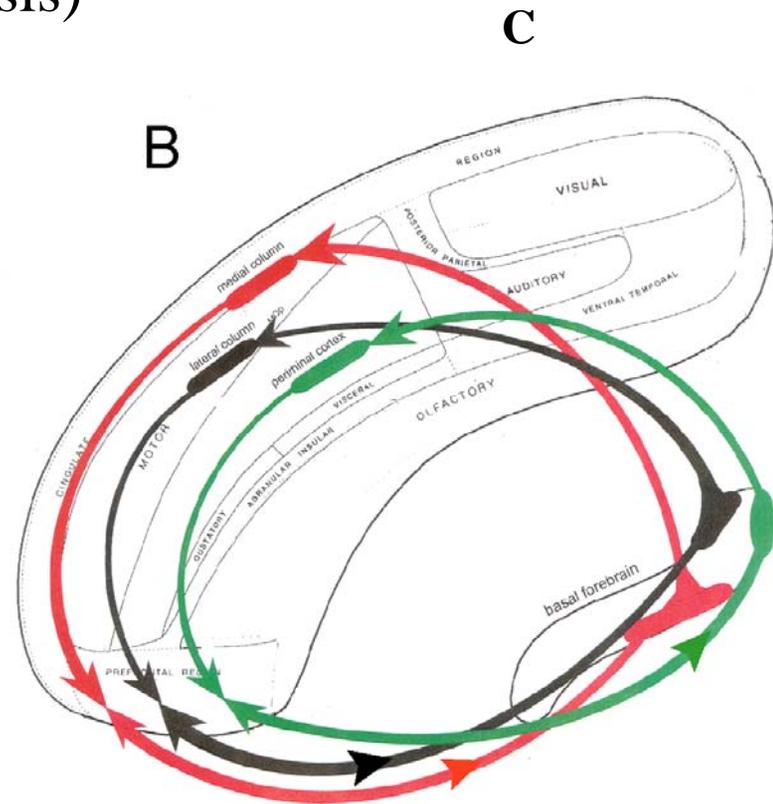
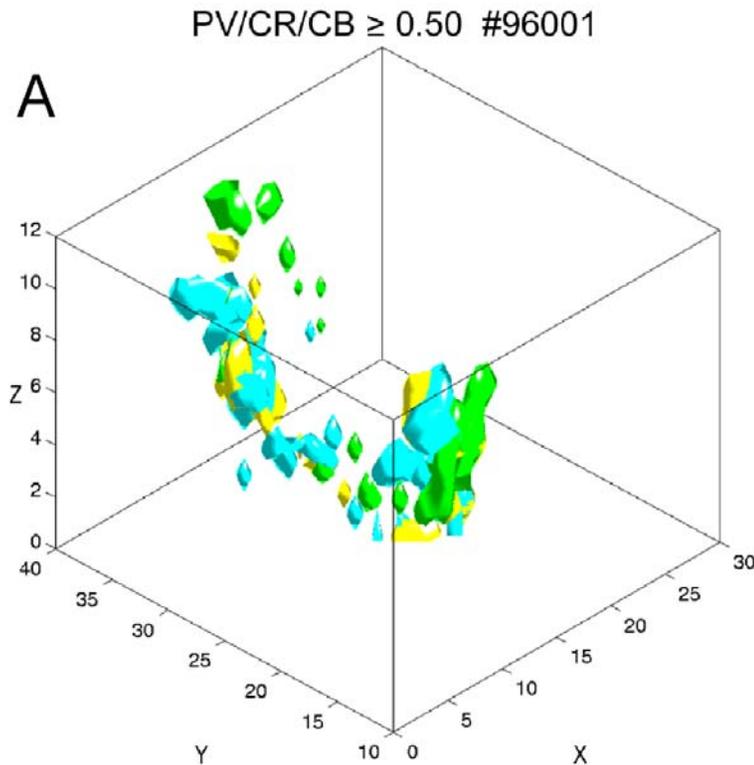
PRESENCE OF SPATIALLY SPECIFIC 'CLUSTERS' IN THE BF



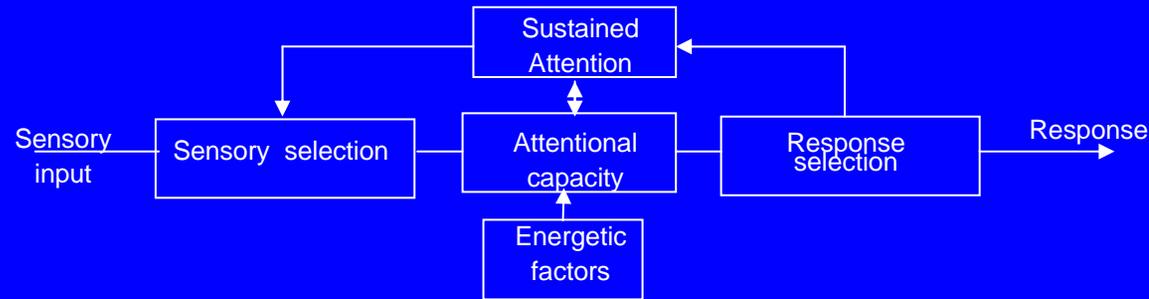


SPECIFIC CELL CLUSTERS IN THE BF TOGETHER WITH SPECIFIC PREFRONTAL AND POSTERIOR ASSOCIATIONAL CELL GROUPS FORM DISTRIBUTED FUNCTIONAL MODULES THAT MAY BE IMPORTANT IN ATTENTIONAL MECHANISMS

(Hypothesis)

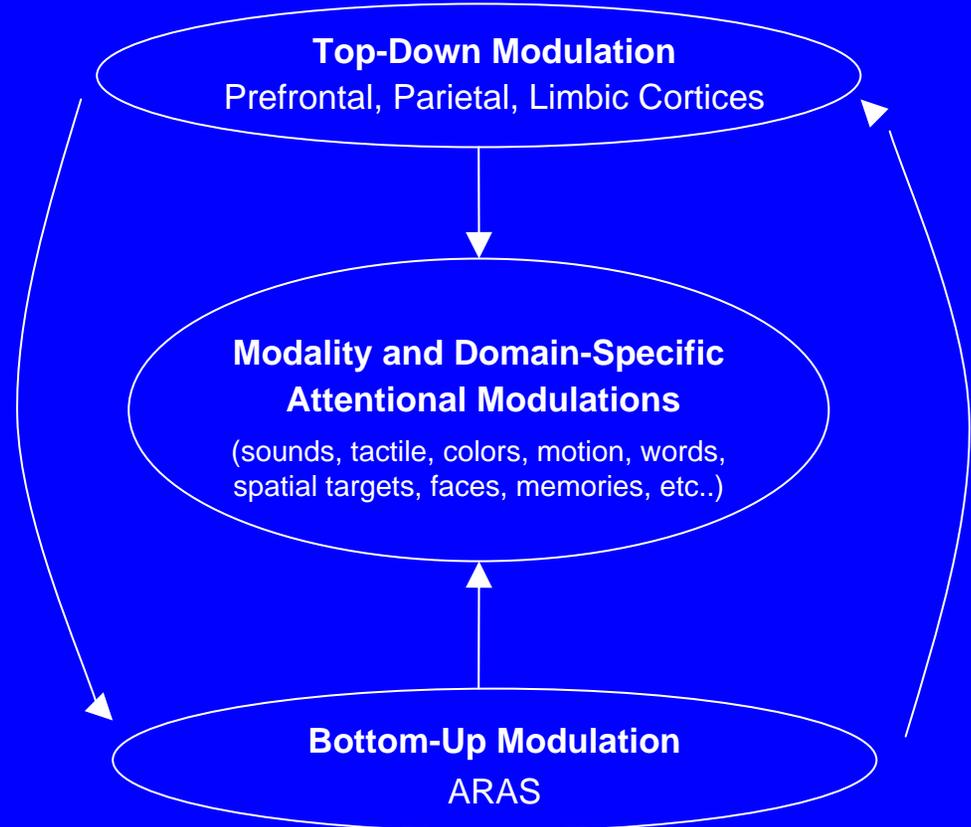


WHERE THE 'BOTTOM UP' MEETS WITH THE 'TOP DOWN'?



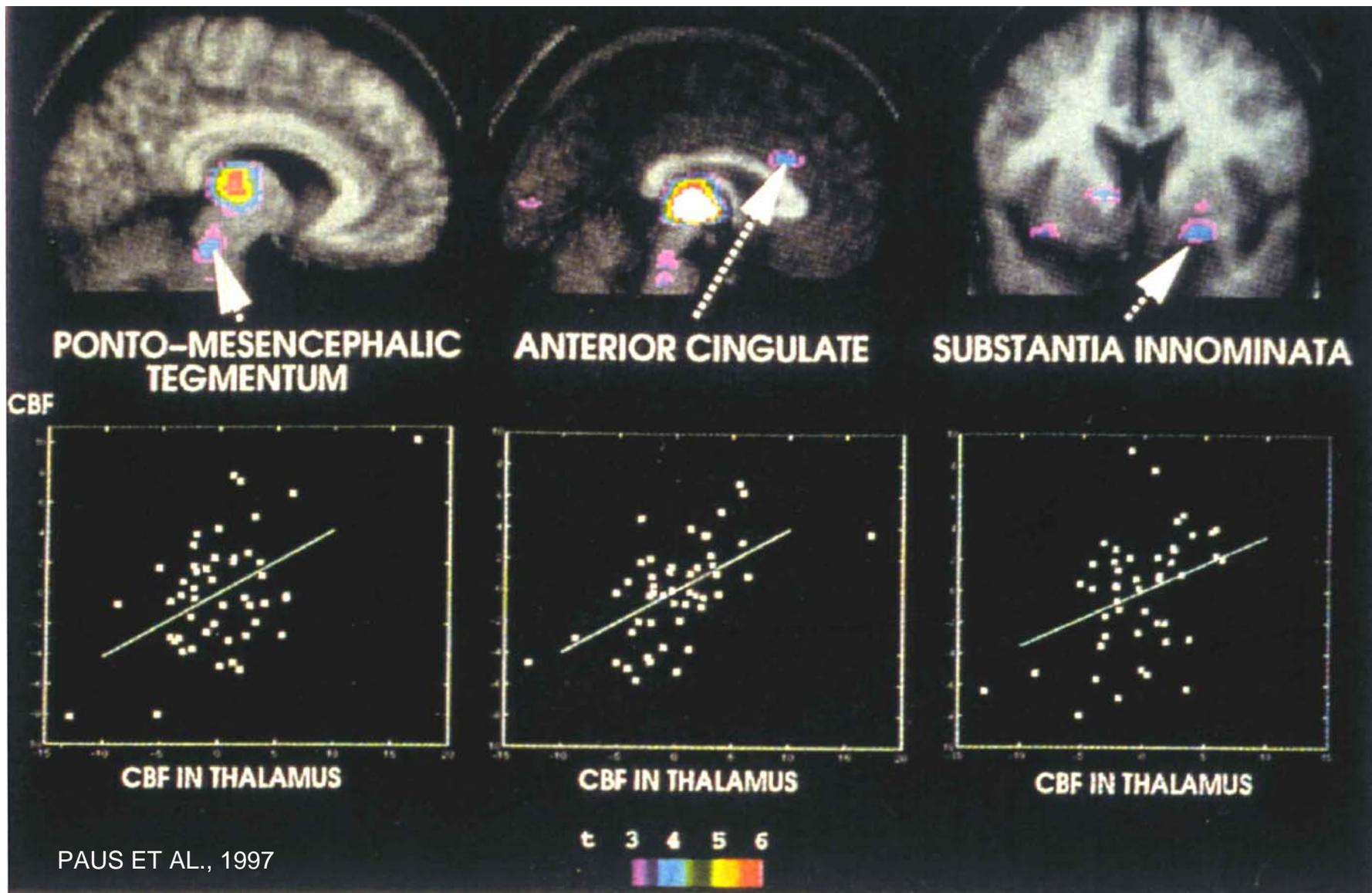
Cohen, 1993

- Attentional mechanisms can act either the input or the output levels of information processing (Broadbent, Treisman, Shiffrin, Shallice, Kahneman, Cohen).
- Attention can be divided into **Bottom-Up**, (filtering of sensory events from a large number of competing stimuli), and **Top-Down processes**, (memory for the significance of stimuli). Top-down processes, are a component of the **prefrontal** mediation of executive functions: they 'amplify computations within a particular posterior sensory area, by reentering the same area that initially performed the computation' (Posner and Raichle)

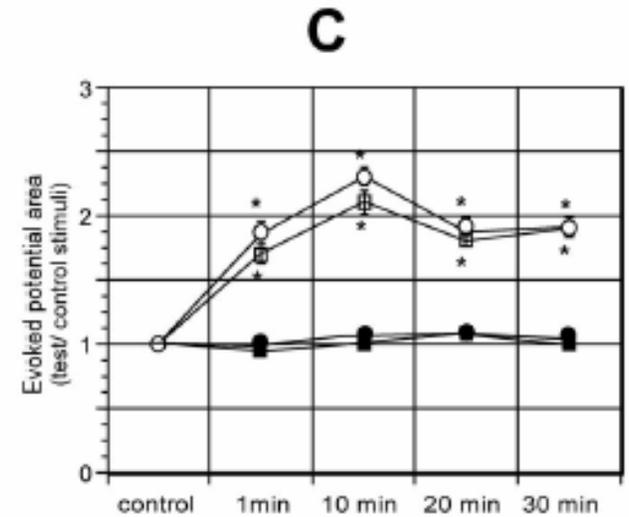
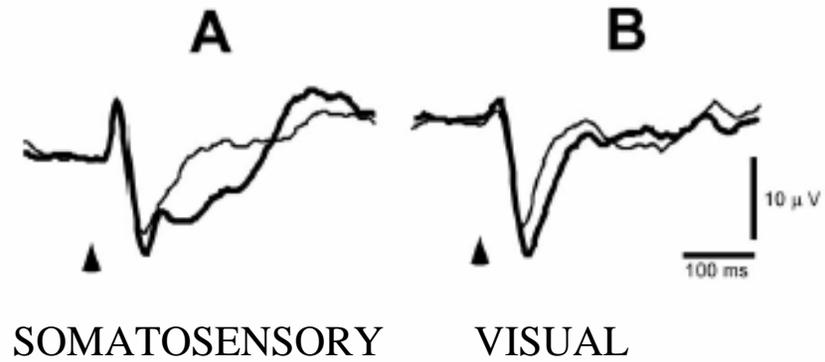
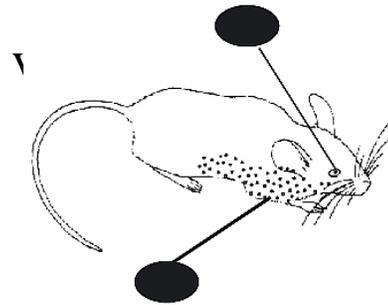
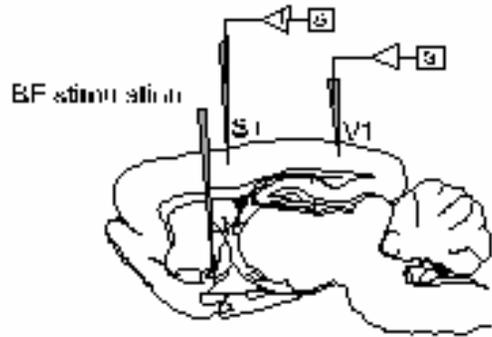


Mesulam, 1990

BASAL FOREBRAIN ACTIVATION IN A SUSTAINED ATTENTION TASK (Paus et al., 1997)

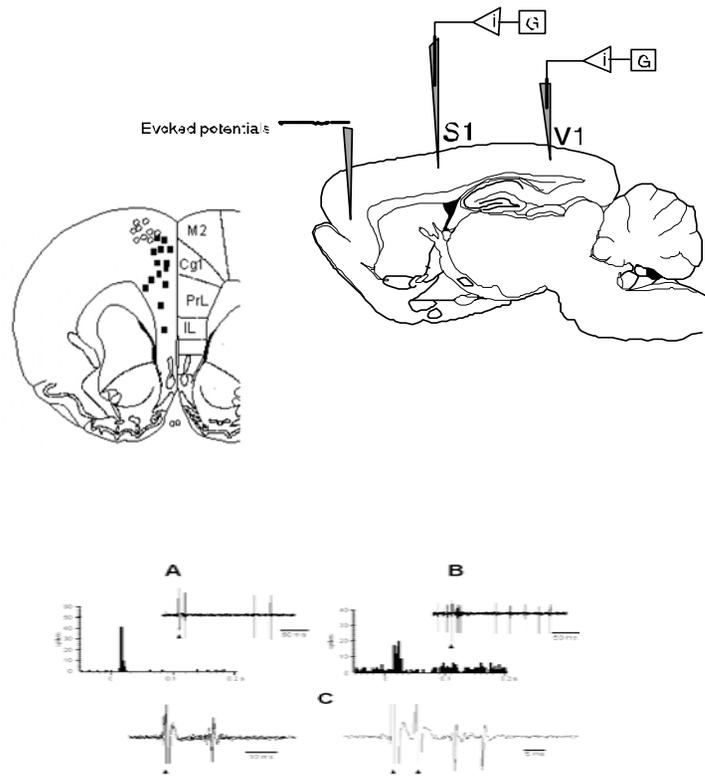


CONFIRMATION that BF STIMULATIONS ENHANCES SENSORY-EVOKED POTENTIALS



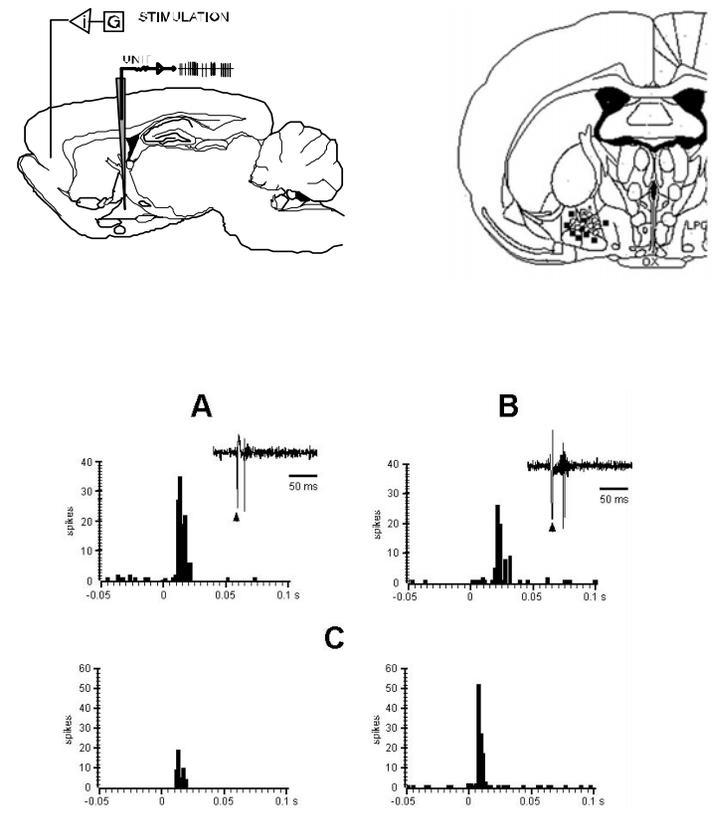
FUNCTIONAL CIRCUITR from POSTERIOR SENSORY AREAS to PFC (i) and hence to BF (ii)

i.



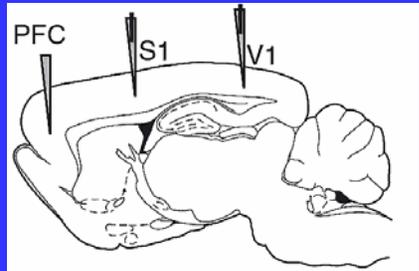
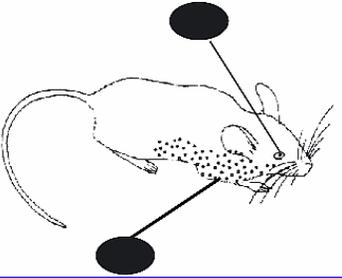
Orthodromic activation of PFC neurons from the somatosensory (A), and from the visual cortex (B).

ii.



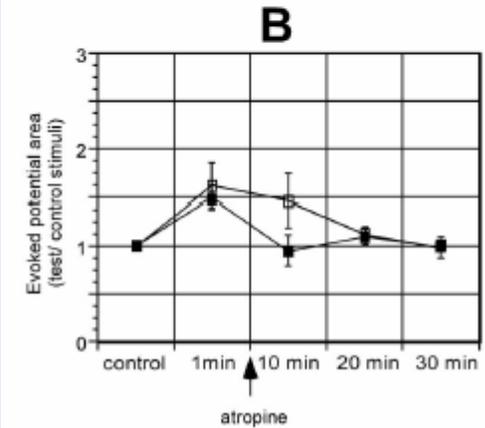
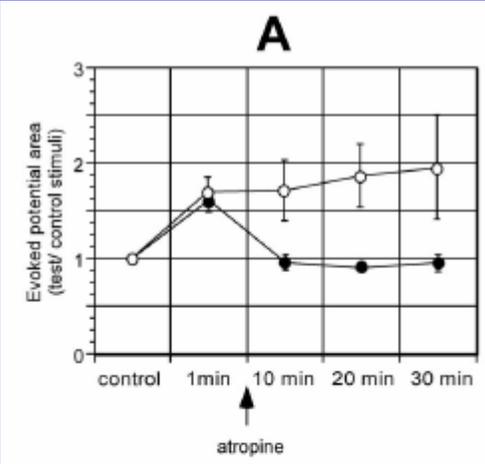
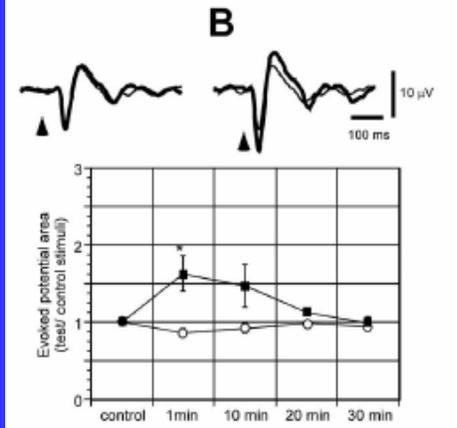
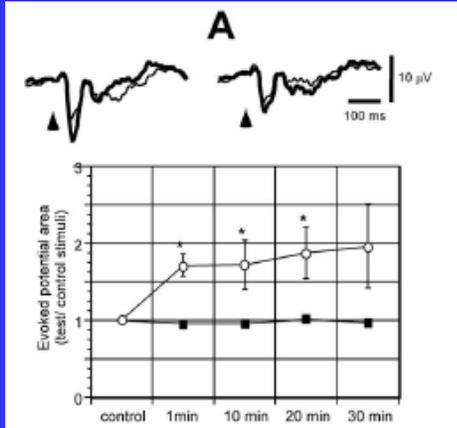
Orthodromic activation of BF neurons by PFC stimulation in the visually responsive area (A) and from the somatosensory-responsive area (B). PTSH of a BF neuron during stimulation of the visual (left) and somatosensory (right) PFC areas

PFC STIMULATIONS SELECTIVELY MODIFY EVOKED RESPONSES in SOMATOSENSORY or VISUAL CORTICES. THESE SENSORY RESPONSES ARE ATROPINE-SENSITIVE



A: Stimulation of the somatosensory-responsive PFC area facilitates SS evoked potentials (open circles and left) but does not modify the visual-evoked potentials (solid squares).

Stimulation of the visual-responsive PFC area slightly decreases SS evoked potentials (open circles) and induces a short-lasting facilitation of the visual-evoked potentials (solid squares and right)



SUMMARY

- Specific BF neurons possess distinct temporal relationships to different EEG patterns. There may be a **dynamic interplay between brainstem-BF and cortical circuitries**
- **Corticopetal neurons** in the BF are uniquely positioned to **integrate** the constant flow of cellular and homeostatic 'states' derived from the **ascending modulatory systems** and to **channel** this momentarily changing neural pattern to the cortical mantle to **modulate alertness**.
- Against the relatively 'diffuse' termination of the ascending axons, the **restricted input from the prefrontal cortex to BF projection neurons** and through local processing might be instrumental in communicating state-related changes to specific posterior sensory areas to **modulate sensory plasticity and attention**.

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